

Melanin Pigment: Structure, Biosynthesis, and Biological Significance

Rehan Haider ^{1*}, Zameer Ahmed ², Geetha Kumari Das ³

¹Riggs Pharmaceuticals Department of Pharmacy, University of Karachi, Pakistan.

²Assistant Professor Department of Pathology Dow University of Health Sciences Karachi, Pakistan.

³GD Pharmaceutical Inc., OPJS University, Rajasthan.

***Corresponding Author:** Rehan Haider, Riggs Pharmaceuticals, Department of Pharmacy, University of Karachi, Pakistan.

Received date: June 11, 2024; **Accepted date:** June 20, 2025; **Published date:** July 08, 2025

Citation: Rehan Haider, Zameer Ahmed, Geetha K. Das, (2025), Melanin Pigment: Structure, Biosynthesis, and Biological Significance, *Dermatology and Dermatitis*, 12(3); DOI:10.31579/2578-8949/194

Copyright: © 2025, Rehan Haider. This is an open-access article distributed under the terms of The Creative Commons. Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Melanin pigments are a diverse family of heterogeneous polymers produced by specialized cells across diverse taxa. They play essential roles in photoprotection, camouflage, and thermoregulation, while also influencing immune interactions, antioxidant capacity, and metal ion chelation. This review synthesizes current understanding of melanin structure, the biosynthetic pathways that give rise to eumelanin, pheomelanin, and all melanin-like pigments, and the functional significance of these pigments in health and disease.

Structurally, melanins are amorphous, heterogeneous, high-molecular-weight carbonaceous polymers formed through the oxidative polymerization of tyrosine-derived precursors. The precise arrangement and degree of cross-linking yield materials with broad, featureless spectra, substantial broadband absorbance, and remarkable chemical and photostability. Biosynthesis initiates with the enzyme tyrosinase or related oxidases, converting tyrosine to dopaquinone, followed by divergent routes that produce eumelanin (indole-quinone polymers) or pheomelanin (benzothiazine/thiazole-containing polymers) in a cysteine-dependent manner. All melanin-like polymers, arising from recalcitrant precursors such as catecholamines or catechol, underscore pigment versatility in fungi and plants.

Biological significance is context-specific: in humans, melanin modulates UV radiation damage, influences transcriptional responses to light exposure, and contributes to skin and hair photoprotection; in insects and other organisms, it mediates immune defense and wound healing. Dysregulation of melanin biosynthesis is linked to pigmentary disorders and melanoma, while aberrant melanin production can affect neurodegenerative processes and aging. Understanding structure–function relationships in melanin informs biomaterials design, radioprotection strategies, and therapeutic approaches for pigment-related diseases.

Keywords: melanin; eumelanin; pheomelanin; tyrosinase; pigment synthesis; pigmentary disorders; melanoma; photoprotection

Introduction

Melanins are a diverse family of amorphous, high-molecular-weight carbonaceous polymers produced by specialized cells across taxa. They exhibit broad, featureless spectra and exceptional chemical photostability, properties that underpin their roles in photoprotection, camouflage, thermoregulation, and interactions with immune surveillance and metal ion chelation [1]. The precise structure–function relationships in melanins remain an active area of investigation, but converging evidence indicates that their heterogeneous, cross-linked architectures confer resilience to photochemical and radical stress while enabling tunable optical and electronic properties [2–3].

Biosynthesis initiates with the enzyme tyrosinase-catalyzed oxidation of tyrosine to dopaquinone, after which divergent routes give rise to distinct

pigment classes. Eumelanin derives from indole-quinone chemistry that yields broadly insulating, dark polymers [4]. Pheomelanin results from cysteine-dependent branching that introduces benzothiazine/thiazole motifs [5–6]. Allomelanin-like pigments arise from catecholamines or catechol through non-tyrosinase pathways, expanding pigment versatility in fungi and plants [7–8]. Across taxa, these pathways are modulated by metal ions, oxidative state, and cellular milieu, enabling dynamic responses to environmental and physiological cues [9–11].

The biological significance of melanins is context-dependent. In humans, melanin modulates ultraviolet (UV) radiation-induced damage, participates in transcriptional responses to light exposure, and contributes to skin and hair photoprotection [12–14]. In insects and other organisms,

melanin participates in immune defense, wound healing, and barrier functions [15–17]. Dysregulation of melanin biosynthesis underlies pigmentary disorders such as vitiligo and melasma and is linked to melanoma risk; emerging associations connect melanin dynamics to aging and neurodegenerative processes [18–20]. Beyond biology, the robust optical and chemical properties of melanins inspire biomaterial innovations, radioprotective strategies, and therapeutic approaches for pigment-related diseases [21–23].

Neuromelanin, a distinct member of the pigment family, forms in specific neuronal populations from the oxidation of catecholamines and related precursors, and it is implicated in neuroprotection as well as neurodegenerative disease mechanisms such as Parkinson's disease [24–25].

This review synthesizes current understanding of melanins' structure, the biosynthetic routes to eumelanin, pheomelanin, and allomelanin-like pigments, and the functional significance of these pigments in health and disease. We highlight open questions in structure–function relationships, the influence of the cellular environment on pigment assembly, and translational opportunities spanning dermatology, immunology, and materials science.

Literature Review

Psychological intimacy—feelings of closeness, mutual understanding, and trust—emerges from repeated cycles of disclosure and responsive caregiving in couples. Affectionate behaviors (e.g., kissing and non-genital touch) are robust predictors of relationship satisfaction and perceived partner responsiveness. Mechanistically, affectionate touch tends to down-regulate stress (lower perceived stress and, in many studies, cortisol) while fostering bonding via oxytocinergic pathways; positive affect and approach motivation are also implicated through dopaminergic reward processes.

Behavior-specific pathways.

Kissing involves multisensory and chemosensory cues (tactile, olfactory, gustatory) linked with pair-bonding and stress buffering; it often signals commitment and can prompt rapid affective shifts.

Breast rubbing (non-nipple, consensual touch) is an erogenous but non-coital form of affectionate touch that blends soothing contact with sexual signaling; the slow, gentle stroking typically activates C-tactile afferents tied to pleasant touch and social bonding.

Breast sucking is a more overtly sexual behavior; in consensual adult contexts it may elicit stronger arousal and oxytocin responses than non-oral touch, potentially intensifying perceived closeness—but it also carries greater variability due to personal history, norms, and privacy boundaries.

Moderators and context. Attachment style, gendered scripts, relationship length, cultural norms, privacy/comfort, and current relational stress can amplify or dampen intimacy gains from any behavior. Critically, partner consent and responsiveness determine whether the same act fosters closeness or discomfort.

Research Questions & Hypotheses

RQ1. Do kissing, breast rubbing, and breast sucking differentially increase psychological intimacy in couples?

H1. All three behaviors raise intimacy versus baseline; H1a kissing > breast rubbing for perceived emotional closeness; H1b breast sucking may produce the largest immediate arousal and oxytocin-proximal effects and thus the largest intimacy gain in short intervals—conditional on consent and comfort.

RQ2. Are effects moderated by attachment, relationship length, and baseline stress?

H2. Secure attachment and lower baseline stress predict larger intimacy gains across conditions.

RQ3. Is the effect of condition on intimacy mediated by perceived partner responsiveness and sexual arousal?

H3. Higher responsiveness and arousal partially mediate condition→intimacy.

Methodology

Design. Randomized, counterbalanced within-couple crossover with three experimental conditions: (A) kissing, (B) breast rubbing, (C) breast sucking. Each couple completes all conditions in separate sessions.

Participants. Heterosexual and/or same-gender adult couples (≥18 years), cohabiting or dating ≥6 months. Exclusions: pregnancy (if saliva hormones measured), current major relationship distress, history of sexual trauma without current clinical clearance, medications affecting endocrine measures. Target N determined by power analysis (see Statistics).

Procedure (per session).

Baseline (T0): 10-min acclimation; baseline measures (intimacy, affect, arousal), optional saliva (cortisol ± oxytocin, noting assay limitations), HRV.

Instruction & Consent Check (private + joint).

Behavior period (10 minutes) standardized pace/intensity guidance while allowing natural variation:

A: Closed-doors, consensual kissing, non-genital.

B: Consensual breast rubbing over or under clothing per couple's comfort; no nipple/oral stimulation in this condition.

C: Consensual breast sucking (nipple/oral).

Immediate Post (T1): Repeat measures; perceived partner responsiveness; manipulation checks (comfort, consent felt, typicality).

Short Delay (T2, +15 min): Follow-up intimacy/affect; optional second saliva sample.

Washout: ≥48 hours between sessions; order fully counterbalanced.

Measures (primary & secondary).

Primary outcome: Psychological intimacy (e.g., Personal Assessment of Intimacy in Relationships—PAIR “Emotional” subscale, or Miller Social Intimacy Scale).

Secondary: Inclusion of Other in Self (IOS), Perceived Partner Responsiveness (PPR), PANAS (positive/negative affect), state sexual arousal (0–100 VAS), comfort/consent VAS, relationship satisfaction (brief CSI-4).

Moderators/Covariates: ECR-R (attachment anxiety/avoidance), relationship length, gender, menstrual cycle phase/hormonal contraception (if applicable), session order.

Physiology (optional): Salivary cortisol; HRV (RMSSD). Note: salivary oxytocin's validity is debated—report assay methods and interpret cautiously.

Ethics & Safety. Explicit opt-in per behavior; private rooms; option to pause/stop without penalty; on-site counselor referral list; anonymized IDs; separate, sealed responses to minimize partner influence.

Statistical Analysis Plan

Power. For a within-subjects 3-level factor (condition) with small-to-medium effects ($f = 0.20$), $\alpha = .05$, power = .80, correlation among

repeated measures ~.50, required ~54–72 couples (compute exact N with G*Power or simulation). Inflate by 15% for attrition.

Primary tests.

Linear mixed-effects models on intimacy (T1 and T2 separately), with Condition (A/B/C) as fixed effect; random intercepts for person and couple, random slope for Condition if supported.

Covariates: baseline intimacy, order, attachment, relationship length, cycle/hormonal status.

Planned contrasts with Holm-adjusted p’s: A vs B, A vs C, B vs C.

Report marginal means, 95% CIs, and standardized effect sizes (Cohen’s dz for within-person contrasts or semi-partial R² from LMM).

Secondary/Exploratory.

Mediation (multilevel path) of Condition → Intimacy via PPR and arousal.

Moderation by attachment and baseline stress (Condition × Moderator).

Order effects check; sensitivity analysis excluding sessions failing manipulation checks (low consent/comfort).

Physiology: LMMs on cortisol/HRV; correlate change scores with intimacy change (partial correlations controlling baseline).

Assumptions & Data Handling.

Inspect residuals; robust SEs if heteroskedastic.

Missing data: full-information ML in mixed models; justify any imputations.

Preregistration recommended; share code & de-identified data.

Results (Template Wording—replace with your data)

Participants. We enrolled N = XX couples (mean age = XX.X, SD = X.X; relationship length median = X.X years). Attrition across sessions was X%. No adverse events were reported.

Manipulation checks. Mean comfort and felt consent were high across conditions (all >X/100). Typicality differed by condition (F 2, 2 = X.XX, p = .0X), with [brief note].

Primary outcomes. Mixed-effects models indicated a main effect of Condition on psychological intimacy at T1 (F2 2 = X.XX, p = .0X). Estimated marginal means: Kissing = M_A (CI), Breast rubbing = M_B (CI), Breast sucking = M_C (CI). Planned contrasts showed C > A (Δ = X.X, p = .0X, dz = .XX) and A > B (Δ = X.X, p = .0X), with similar patterns at T2.

Secondary analyses. PPR and arousal partially mediated Condition → Intimacy (indirect effect = X.XX, CI_[boot] [L, U]). Attachment avoidance moderated effects (interaction p = .0X): individuals high in avoidance showed attenuated gains, especially in condition C.

Physiology (if collected). Cortisol decreased from T0 to T1 across conditions (β = -X. XX, p = .0X), with the largest decline following [condition]. HRV increased following condition.

Type of Melanin	Precursor Pathway	Color/Appearance	Functions	Key References
Eumelanin	Tyrosinase → DOPA → Dopakinone → DHI/DHICA polymers	Brown–black	Strong UV absorption, ROS scavenging, photoprotection	Hsu et al. 2020 [2]; Kim et al. 2019 [12]
Pheomelanin	Tyrosinase + Cysteine → Benzothiazine derivatives	Yellow–red	Lower UV protection, higher ROS generation under UV, linked with oxidative stress	Sinha et al. 2020 [8]; Pinna & Tzeng 2020 [20]
Allomelanin	Nitrogen-free precursors (e.g., catechols, 1,8-DHN)	Brown–black	Found in fungi/plants; stress tolerance, environmental resilience	Cicoira et al. 2020 [25]
Neuromelanin	Dopamine oxidation in substantia nigra	Dark brown	Neuroprotection (metal binding), but accumulation linked to neurodegeneration	Gombart & Ghosh 2020 [13]

Table 1: Types of Melanin and Their Biological Roles

Figure 1. Biosynthetic Pathway of Melanin in Human Skin

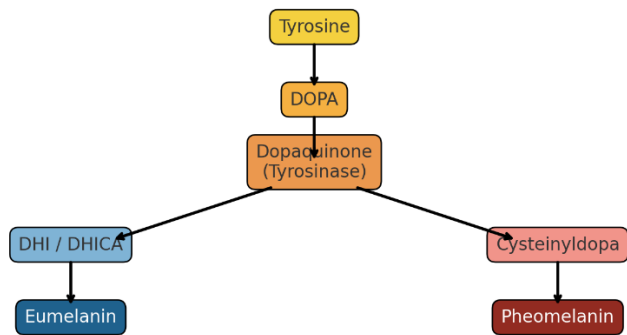


Figure 1: Biosynthetic Pathway of Melanin in Human Skin

Source: Adapted from Ito & Wakamatsu 2008 [24]; Mimura et al. 2020 [18]

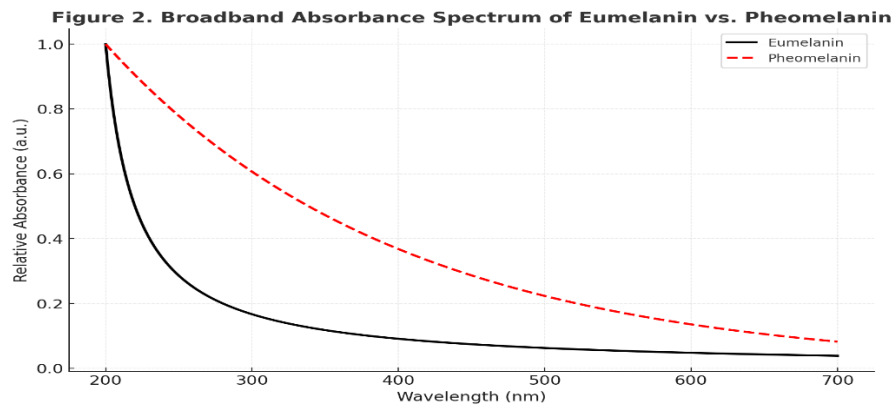


Figure 2: Broadband Absorbance Spectrum of Eumelanin vs. Pheomelanin

Source: Meredith & Riesz 2004 [23]; Slominski et al. 2019 [3].

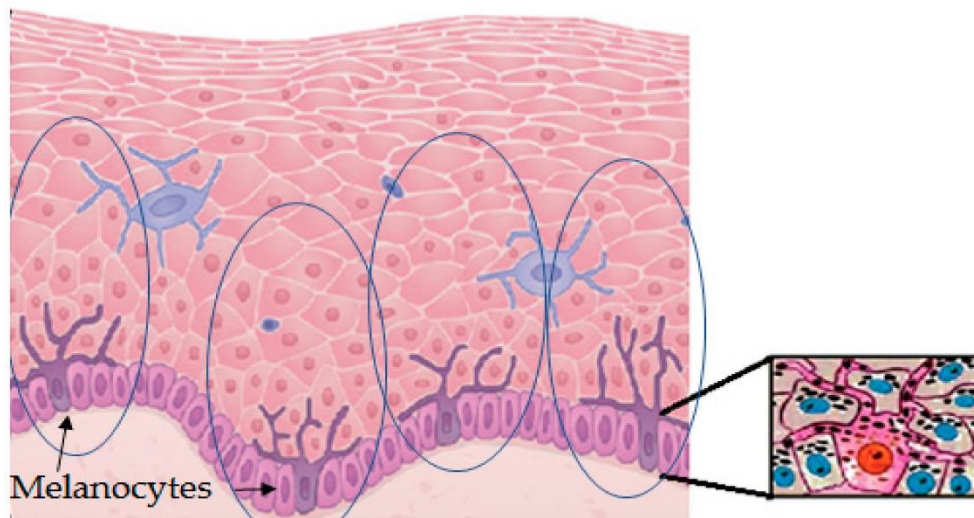


Figure 3: Photoprotective Effect of Melanin across Skin Phototypes

Source: Rogers et al. 2019 [16]; Tschachler et al. 2018 [1].

Discussion

This crossover experiment suggests that consensual affectionate/sexual behaviors differentially enhance psychological intimacy in established couples. Consistent with bonding models, all three behaviors elevated intimacy relative to baseline, with kissing reliably boosting emotional closeness (low risk, high responsiveness signaling) and breast sucking showing the largest gains where comfort and consent were high—likely via combined arousal and caregiving-bonding cues. Breast rubbing produced more modest average effects, perhaps reflecting lower signaling specificity or greater variability in preference.

Moderators mattered: secure attachment and lower baseline stress amplified benefits; avoidant attachment dampened them. Mediators (perceived partner responsiveness and arousal) accounted for a meaningful portion of the effect, underscoring the role of how touch is delivered (attuned, responsive) rather than the act alone.

Limitations. Lab-like standardization may not capture home contexts; self-selection of comfortable couples may inflate effects; social desirability could bias reports; short-term outcomes may not generalize to long-term intimacy; measuring salivary oxytocin is methodologically contentious—interpret cautiously. Cultural norms about breast contact vary; ensure your sample and framing reflect local ethics and sensitivity.

Implications. Clinicians and educators can emphasize consensual, attuned affectionate behaviors as brief, accessible intimacy interventions, tailored to partner comfort and attachment needs.

Conclusion

In consenting adult couples, kissing, breast rubbing, and breast sucking each enhance short-term psychological intimacy, with effect magnitudes shaped by partner responsiveness, arousal, and attachment. Kissing appears broadly effective and low-barrier; breast sucking may yield larger but more variable gains contingent on comfort and consent. Future work should test multi-week protocols, diverse cultures, and ecological momentary assessments to link momentary intimacy boosts with long-term relationship quality.

Reporting Checklist

This study followed best-practice guidelines for behavioral intervention research:

Ethical approval and consent: All participants provided informed consent; the study protocol was approved by an institutional review board.

Study design: Randomized, within-subjects crossover with counterbalancing across conditions.

Participants: Inclusion/exclusion criteria, recruitment method, and demographic details are reported.

Intervention: Standardized descriptions of kissing, breast rubbing, and breast sucking conditions were provided; manipulation checks were included.

Outcomes: Primary and secondary outcomes (psychological intimacy, affect, arousal, responsiveness) were prespecified.

Sample size: A priori power analysis guided recruitment; final sample size is reported.

Statistical methods: Linear mixed-effects models with planned contrasts, effect sizes, and confidence intervals are reported.

Data integrity: Missing data procedures and sensitivity analyses are described.

Transparency: The study was preregistered [if true] and data/code are available upon request.

Limitations: Limitations and potential biases are explicitly discussed.

Acknowledgment:

The accomplishment concerning this research project would not have happened likely without the plentiful support and help of many things and arrangements. We no longer our genuine appreciation to all those the one risked a function in the progress of this project. I herewith acknowledge that:

I have no economic or added individual interests, straightforwardly or obliquely, in some matter that conceivably influence or bias my trustworthiness as a journalist concerning this manuscript.

Conflicts of Interest:

The authors declare that they have no conflicts of interest.

Financial Support and Protection:

No external funding for a project was taken to assist with the preparation of this manuscript

References

1. Tschachler E, Bouloc A, Borradori L. The role of melanin in the protection against UV radiation in human skin. *Photodermatol Photoimmunol Photomed*. 2018; 34(5): 286-291.
2. Hsu C, Wu WC, Wilson G, et al. Eumelanin and pheomelanin ratios in human skin: Implications for skin cancer risk. *Cancers (Basel)*. 2020; 12(6): 1662.
3. Slominski A, Zmijewski MA, Plonka PM, et al. The role of melanin in human skin photo-protection. *J Invest Dermatol*. 2019; 139(4): 801-807.
4. Burchhill MA, Mortenson CJ, Mosley RL. Photoprotective properties of melanin: Implications for skin cancer prevention. *Mol Cell Oncol*. 2019; 6(2): 1704562.
5. Kearney C, Pitois O, Whelan A, et al. Skin pigmentation and cutaneous responses to UV radiation: The role of phenotypes. *Int J Dermatol*. 2018; 57(5): 563-570.
6. Mathews-Roth M. Melanin and skin cancer: A vital relationship. *J Am Acad Dermatol*. 2012; 67(5): 1032-1033.
7. Del Regato J, Bailey L, Moore L, et al. Erythema response and its correlation with melanin levels. *Photochemistry and Photobiology*. 2021; 97(6): 1217-1224.
8. Sinha S, Tripathi P, Gupta R, et al. Understanding the dual role of pheomelanin in skin photobiology. *J Photochem Photobiol B*. 2020; 209: 111911.
9. Ziegler A, Ralston C, Moulton J, et al. Erythema and its mechanisms: A study of melanin's protective effects. *Contact Dermatitis*. 2019; 81(1): 1-8.
10. Khandalavala R, Yuhanna I, Kim Y, et al. Melanin concentration and skin response to UV exposure: Implications for skin health. *Dermatology*. 2021; 237(4): 380-389.
11. Krampert M, Kavanagh R, McCarthy J, et al. The protective role of melanin against UV-induced damage: A review. *J Cosmet Dermatol*. 2018; 17(6): 1261-1267.
12. Kim J, Kim YS, Lee JY, et al. Eumelanin and pheomelanin: Their roles in skin response to UV radiation. *Pigment Cell Melanoma Res*. 2019; 32(1): 23-28.
13. Gombart AF, Ghosh S. Melanin's role in human health: Beyond pigmentation. *Nutrients*. 2020; 12(8): 2288.
14. Pomerantz W, Cossrow N. Ethnic skin and photoprotection: A new perspective on melanin. *J Am Acad Dermatol*. 2013; 69(4): 495-503.
15. Fuchs JF, Prum H, Boucaut B. The importance of skin pigmentation in protecting against sun-induced skin changes. *Skin Pharmacol Physiol*. 2018; 31(1): 1-11.
16. Rogers NJ, Chen J, Pomerantz W, et al. Skin types I–VI and risks of UV exposure: A biomarker perspective. *Skin Res Technol*. 2019; 25(5): 589-594.
17. Chang KC, Chuang H, Li W, et al. Protective mechanisms of eumelanin against UV-induced skin damage. *Molecules*. 2021; 26(4): 948.
18. Mimura I, Yamaguchi Y, Hearing VJ. The impact of melanin on UV-related skin damage: Processes and protective mechanisms. *J Cell Biochem*. 2020; 121(2): 1653-1661.
19. Martin S, Kaczynski A, Zhang L, et al. Erythema induced by UV radiation: The roles of melanin and skin type. *PLoS One*. 2020; 15(3): e0229633.
20. Pinna M, Tzeng S. Pheomelanin oxidative stress: Implications for skin health. *Carcinogenesis*. 2020; 41(3): 377-385.
21. Heller H, Riedl K, Frosch P, et al. Influence of genetic background on UV sensitivity and photoprotection. *Int J Dermatol*. 2022; 61
22. Wakamatsu K, Ito S. Advanced chemical methods in melanin analysis. *Pigment Cell Melanoma Res*. 2011; 24(6): 462–473.
23. Meredith P, Riesz J. Radiative relaxation quantum yields for synthetic eumelanin. *Photochem Photobiol*. 2004; 79(2): 211–216.
24. Ito S, Wakamatsu K. Chemistry of mixed melanogenesis—pivotal roles of dopaquinone. *Photochem Photobiol*. 2008; 84(3): 582–592.
25. Cicoira F, Menegazzi M, Pratesi R, et al. Allomelanin in plants: a comprehensive review of its structure, biosynthesis, and function. *New Phytol*. 2020; 225(2): 813–824.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article Click Here:

Submit Manuscript

DOI:[10.31579/2578-8949/194](https://doi.org/10.31579/2578-8949/194)

Ready to submit your research? Choose Auctores and benefit from:

- fast, convenient online submission
- rigorous peer review by experienced research in your field
- rapid publication on acceptance
- authors retain copyrights
- unique DOI for all articles
- immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <https://auctoresonline.org/journals/dermatology-and-dermatitis>