

Application of Pyrimidine Derivatives as New Regulators to Enhance Wheat Growth in The Vegetative Phase

Tsygankova V.A.*, Andrushevich Ya.V., Vasylenko N.M., Kopich V.M., Solomyannyi R.M., Kachaeva M.V., Pilyo S.G., Bondarenko O.M., Popilnichenko S.V., Brovarets V.S.

Department for Chemistry of Bioactive Nitrogen-Containing Heterocyclic Compounds, V.P. Kukhar Institute of Bioorganic Chemistry and Petrochemistry, National Academy of Sciences of Ukraine, 1, Academician Kukhar str., 02094, Kyiv-94, Ukraine.

Corresponding author: Tsygankova Victoria Anatolyivna, Dr. Biol. Sci., Principal researcher, Senior Staff Scientist, Ukraine.

Received date: March 17, 2025; **Accepted date:** March 31, 2025; **Published date:** April 21, 2025

Citation: Tsygankova V.A., Andrushevich Ya.V., Vasylenko N.M., Kopich V.M., Solomyannyi R.M., et al., (2025), Application of Pyrimidine Derivatives as New Regulators to Enhance Wheat Growth in The Vegetative Phase, *J. Nutrition and Food Processing*, 8(6); DOI:10.31579/2637-8914/306

Copyright: © 2025, Tsygankova V.A. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract:

The regulatory effect of synthetic low-molecular-weight azaheterocyclic compounds, pyrimidine derivatives on the growth and photosynthesis of winter wheat (*Triticum aestivum* L.) variety Taira in the vegetative phase was studied. Wheat growth parameters such as average shoot length (mm) and average root length (mm) and wheat photosynthesis parameters such as chlorophyll and carotenoid content (mg/g fresh weight) under the regulatory effect of pyrimidine derivatives were measured 4 weeks after seed germination and compared with those of control wheat grown in distilled water or wheat grown under the regulatory effect of auxin IAA. The study showed that the wheat growth parameters and photosynthetic parameters under the regulatory effect of synthetic compounds, pyrimidine derivatives at a concentration of 10^{-6} M were similar to or higher than the parameters of wheat under the regulatory effect of auxin IAA at a similar concentration of 10^{-6} M, and also exceeded that of the control wheat grown in distilled water. The selectivity of the regulatory effect of synthetic compounds, pyrimidine derivatives on wheat growth and photosynthesis depended on the substituents in their chemical structure. The most physiologically active synthetic compounds, pyrimidine derivatives have been proposed for use to enhance the growth and photosynthesis of wheat in the vegetative phase.

Key words: Wheat; azaheterocyclic compounds; pyrimidine derivatives; IAA

Introduction

Wheat (*Triticum aestivum* L.) is a major cereal crop grown worldwide [1, 2]. Wheat grain is a source of biologically active compounds beneficial to human nutrition and health, including proteins, fatty acids, carbohydrates, dietary fiber, minerals such as calcium, magnesium, phosphorus, potassium, zinc, iron, and copper, vitamins such as thiamin (vitamin B1), riboflavin (vitamin B2), niacin (vitamin B3), pyridoxine (vitamin B6), folic acid (vitamin B9), tocopherol (vitamin E), and phytochemicals such as flavonoids, glycosides, alkaloids, steroids, saponins, terpenoids, and tannins [1–5].

Growing wheat in today's unfavorable climatic conditions, abiotic and biotic stresses reduce wheat yields, which require the development of new environmentally friendly technologies that can prevent environmental pollution and not harm human health [2, 6–10]. Currently, phytohormones, synthetic plant growth regulators, natural biostimulants

and biopesticides are increasingly used to increase wheat productivity and adaptation to abiotic and biotic stresses [11–21].

Plant productivity and stress tolerance depends on the development of the root system, which provides plants with organic matter, micro- and macroelements from the soil, affecting the formation and growth of vegetative and reproductive organs of plants [22–24]. As is known, phytohormones auxins and cytokinins play an important role in regulating the growth and development of the root system, shoots, leaves, flowers and seeds of plants, in photosynthesis in plant leaves, as well as in the adaptation of plants to biotic and abiotic stress factors [25–29]. The use of phytohormones has a positive effect on the interaction of plants with soil microorganisms; data on the therapeutic effects of phytohormones on mammals have also been obtained [30].

During plant ontogenesis and under stressful conditions, changes in the metabolism and homeostasis of endogenous plant auxins and cytokinins occur [31-33]. Natural phytohormones auxins and cytokinins or their synthetic analogues, as well as biostimulants, when applied exogenously, are capable of exerting a direct regulatory effect on plant growth or manipulating the biosynthesis and metabolism of endogenous phytohormones, improving plant growth, increasing their productivity and immune-mediated resistance to abiotic and biotic stresses [16-20, 34-43].

In recent years, significant progress has been achieved in the development of new effective and environmentally friendly plant growth regulators based on synthetic low-molecular-weight azaheterocyclic compounds that have a regulatory effect on plant growth similar to the phytohormones auxins and cytokinins [44, 45]. Among the various classes of synthetic azaheterocyclic compounds, pyrimidines, which are used in medicine as therapeutic agents for the treatment of various diseases [46-52], as well as in agriculture as plant growth regulators, herbicides and insecticides [53-60], have the most similar physiological effect to the phytohormones auxins and cytokinins [44, 45].

Over the past decade, numerous studies have been conducted devoted to the screening for new auxin- and cytokinin-related substances among synthetic low-molecular-weight azaheterocyclic compounds, pyrimidine derivatives that regulate plant growth [44, 45]. It has been shown that synthetic azaheterocyclic compounds, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), as well as other pyrimidine derivatives, are capable of

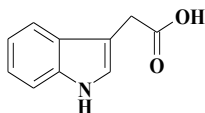
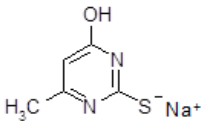
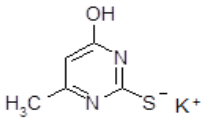
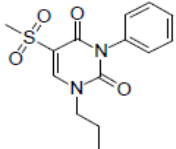
exhibiting a broad regulatory effect on various agricultural crops in low concentrations that are non-toxic to human health and the environment, improving plant growth throughout the vegetative phase and increasing their yield [61-74]. A comparative analysis of plant growth regulatory activity indicates that the regulatory effect of these synthetic azaheterocyclic compounds, pyrimidine derivatives is equivalent to, or exceeds, the regulatory effect of auxins and cytokinins on plant growth and development. It is thanks to these unique properties that pyrimidine derivatives can find practical use in agriculture as new effective and environmentally friendly plant growth regulators.

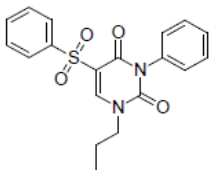
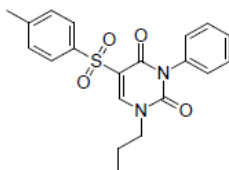
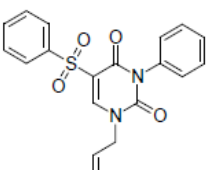
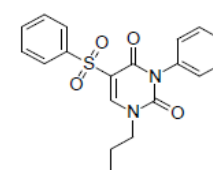
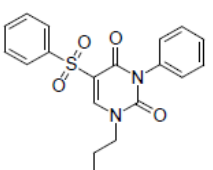
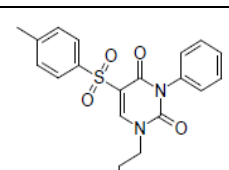
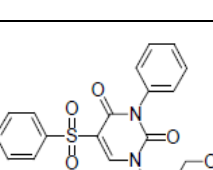
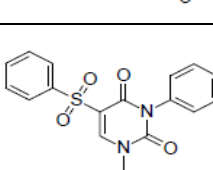
The aim of this work is screening of physiologically active synthetic low-molecular-weight azaheterocyclic compounds, pyrimidine derivatives, capable of regulating the growth and photosynthesis of winter wheat (*T. aestivum* L.) variety Taira.

Materials and Methods

Chemical name and structure of the studied compounds

Synthetic low-molecular-weight azaheterocyclic compounds, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur) were synthesized at the Department for Chemistry of Bioactive Nitrogen-Containing Heterocyclic Compounds, V.P. Kukhar Institute of Bioorganic Chemistry and Petrochemistry of the National Academy of Sciences of Ukraine; auxin IAA (1*H*-indol-3-yl) acetic acid) was manufactured by Sigma-Aldrich, USA (Table 1).

Chemical compound	Chemical structure	Chemical name and relative molecular weight (g/mol)
IAA		1 <i>H</i> -indol-3-ylacetic acid MW=175.19
Methyur		Sodium salt of 6-methyl-2-mercapto-4-hydroxypyrimidine MW=165.17
Kamethur		Potassium salt of 6-methyl-2-mercapto-4-hydroxypyrimidine MW=181.28
1		5-Methanesulfonyl-3-phenyl-1-propyl-1 <i>H</i> -pyrimidine-2,4-dione MW=308.3586

2		5-Benzenesulfonyl-3-phenyl-1-propyl-1 <i>H</i> -pyrimidine-2,4-dione MW=370.4303
3		3-Phenyl-1-propyl-5-(toluene-4-sulfonyl)-1 <i>H</i> -pyrimidine-2,4-dione MW=384.4574
4		1-Allyl-5-benzenesulfonyl-3-phenyl-1 <i>H</i> -pyrimidine-2,4-dione MW=368.4144
5		5-Benzenesulfonyl-1-butyl-3-phenyl-1 <i>H</i> -pyrimidine-2,4-dione MW=384.4574
6		5-Benzenesulfonyl-1-(3-hydroxypropyl)-3-phenyl-1 <i>H</i> -pyrimidine-2,4-dione MW=386.4297
7		1-(3-Hydroxypropyl)-3-phenyl-5-(toluene-4-sulfonyl)-1 <i>H</i> -pyrimidine-2,4-dione MW=400.4568
8		5-Benzenesulfonyl-1-(2,3-dihydroxypropyl)-3-phenyl-1 <i>H</i> -pyrimidine-2,4-dione MW=402.4291
9		5-Benzenesulfonyl-3-phenyl-1-(tetrahydrofuran-2-ylmethyl)-1 <i>H</i> -pyrimidine-2,4-dione MW=412.4680

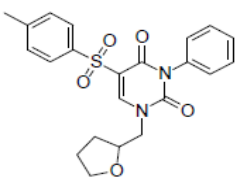
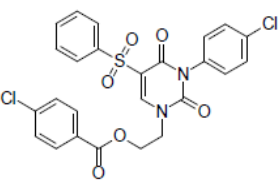
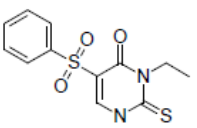
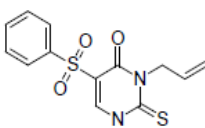
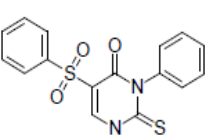
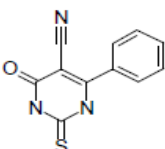
10		3-Phenyl-1-(tetrahydrofuran-2-ylmethyl)-5-(toluene-4-sulfonyl)-1H-pyrimidine-2,4-dione MW=426.4950
11		4-Chlorobenzoic acid 2-[5-benzenesulfonyl-3-(4-chlorophenyl)-2,4-dioxo-3,4-dihydro-2H-pyrimidin-1-yl]-ethyl ester MW=545.4020
12		5-Benzenesulfonyl-3-ethyl-2-thioxo-2,3-dihydro-1H-pyrimidin-4-one MW=296.3690
13		3-Allyl-5-benzenesulfonyl-2-thioxo-2,3-dihydro-1H-pyrimidin-4-one MW=308.3802
14		5-Benzenesulfonyl-3-phenyl-2-thioxo-2,3-dihydro-1H-pyrimidin-4-one MW=344.4136
15		4-Oxo-6-phenyl-2-thioxo-1,2,3,4-tetrahydro-pyrimidine-5-carbonitrile MW=229.2619

Table 1: Chemical structure of IAA (1H-indol-3-yl) acetic acid), sodium salt of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur), potassium salt of 6-methyl-2-mercapto-4-hydroxypyrimidine (Kamethur), and pyrimidine derivatives № 1–15

Plant growing conditions

The seeds of winter wheat (*T. aestivum* L.) variety Taira were sterilized with 1 % KMnO₄ solution for 10 min, then treated with 96 % ethanol solution for 1 min, after which they were washed three times with sterile distilled water. After this procedure, seeds were placed in the plastic cuvettes (each containing 20-25 seeds) on the perlite moistened with distilled water (control sample) or water solutions of auxin IAA (1H-indol-3-yl)acetic acid or synthetic low-molecular-weight azaheterocyclic compounds, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), or pyrimidine derivatives (compounds № 1–15) at a concentration of 10⁻⁶M (experimental samples). Then seeds were placed in the thermostat for germination in darkness at the temperature 20-22 °C during 48 h. The germinated seeds were placed in a climatic chamber, in which the plants were grown for 4 weeks under a light/dark regime of 16/8 h, a temperature of 20-22 °C, a light intensity of 3000 lux, and an air humidity of 60-80 %. Comparative analysis of plant growth parameters, such as average shoot

length (mm), average root length (mm), was performed at the end of the 4-week period according to the methodical manual [75]. Plant growth parameters determined on experimental plants, in comparison with similar parameters of control plants, were expressed as %.

Determination of chlorophyll and carotenoid content

The content of photosynthetic pigments such as chlorophylls and carotenoids (mg/g fresh weight) in wheat leaves was analyzed according to methodological recommendations [76, 77]. To perform the extraction of photosynthetic pigments, we homogenized a sample (500 mg) of wheat leaves in the porcelain mortar in a cooled at the temperature 10 °C 96 % ethanol at the ratio of 1:10 (weight:volume) with addition of 0.1-0.2 g CaCO₃ (to neutralize the plant acids). The 1 ml of obtained homogenate was centrifuged at 8000 g in a refrigerated centrifuge K24D (MLW, Engelsdorf, Germany) during 5 min at the temperature 4 °C. The obtained precipitate was washed three times, with 1 ml 96 % ethanol and centrifuged at above mentioned conditions. After this procedure, the optical density of chlorophyll a, chlorophyll b and carotenoid in the

obtained extract was measured using spectrophotometer Specord M-40 (Carl Zeiss, Germany).

The content of chlorophyll a, chlorophyll b, and carotenoids in plant leaves was calculated in accordance with formula [76, 77]:

$$\text{Cchl a} = 13.36 \times A_{664.2} - 5.19 \times A_{648.6},$$

$$\text{Cchl b} = 27.43 \times A_{648.6} - 8.12 \times A_{664.2},$$

$$\text{Cchl (a + b)} = 5.24 \times A_{664.2} + 22.24 \times A_{648.6},$$

$$\text{Ccar} = (1000 \times A_{470} - 2.13 \times \text{Cchl a} - 97.64 \times \text{Cchl b}) / 209,$$

Where, Cchl – concentration of chlorophylls (µg/ml), Cchl a – concentration of chlorophyll a (µg/ml), Cchl b – concentration of chlorophyll b (µg/ml), Ccar – concentration of carotenoids (µg/ml), A – absorbance value at a proper wavelength in nm.

The chlorophyll and carotenoids content per 1 g of fresh weight of extracted from leaves was calculated by the following formula (separately for chlorophyll a, chlorophyll b and carotenoids):

$$A_1 = (C \times V) / (1000 \times a_1),$$

where, A₁ – content of chlorophyll a, chlorophyll b, or carotenoids (mg/g fresh weight), C – concentration of pigments (µg/ml), V – volume of extract (ml), a₁ – sample of leaves (g).

The content of photosynthetic pigments determined in the leaves of experimental wheat plants in relation to control plants was expressed as %.

Statistical data analysis

Each experiment was performed three times. Statistical processing of the experimental data was carried out using Student's t-test with a significance level of $P \leq 0.05$; mean values \pm standard deviation (\pm SD) [78].

Results and Discussion

Regulatory effect of pyrimidine derivatives on wheat growth

A comparative analysis of the regulatory effect of auxin IAA (1*H*-indol-3-yl)acetic acid, synthetic low-molecular-weight azaheterocyclic compounds, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), and pyrimidine derivatives № 1–15 in a concentration of 10^{-6} M on the growth and development of winter wheat (*T. aestivum* L.) variety Taira was conducted. Wheat growth parameters such as average shoot length (mm), average root length (mm), measured at the end of 4 weeks after seed germination, were compared with those of control wheat grown in distilled water.

The study showed that the growth parameters of wheat shoots and roots under the regulatory effect of derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), or pyrimidine derivatives № 1–15 were similar to or higher than the growth parameters of wheat under the regulatory effect of auxin IAA, and also significantly exceeded the growth parameters of control wheat grown in distilled water (Figure 1).

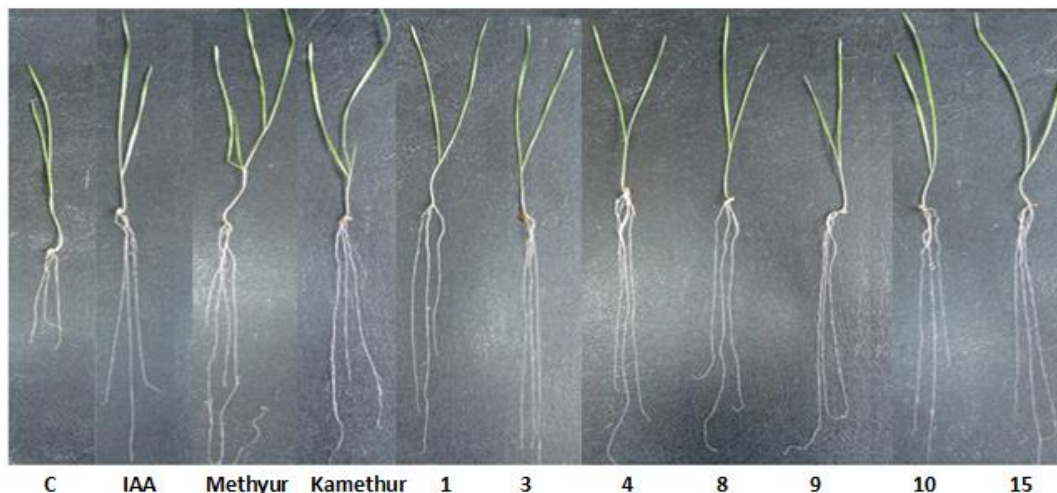


Figure 1: The regulatory effect of auxin IAA, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), as well as the most physiologically active pyrimidine derivatives № 1, 3, 4, 8, 9, 10, 15 at a concentration of 10^{-6} M on the growth parameters of shoots and roots of 4-week-old winter wheat (*T. aestivum* L.) variety Taira compared to control (C) plants.

It was shown that the highest regulatory effect on the average shoot length (mm) is exhibited by derivative of sodium salt of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur) and pyrimidine derivatives № 4, 5, 6, 8, 9, 10, 11, 13, 14, 15, under the effect of which these parameters increase: by 36.38% - under the effect of Methyur, by 43.5–71.75% - under the effect of pyrimidine derivatives № 4, 5, 6, 8, 9, 10, 11, 13, 14, 15, compared to similar parameters of control plants (Figure 2).

The lower regulatory effect on the average shoot length (mm) is exhibited by auxin IAA, derivative of potassium salt of 6-methyl-2-mercapto-4-hydroxypyrimidine (Kamethur) and pyrimidine derivatives № 1, 2, 3, 7, 12, under the effect of which these parameters increase: by 9.6% - under the effect of IAA, by 20.9% - under the effect of Kamethur, by 12.88–30.85% - under the effect of pyrimidine derivatives № 1, 2, 3, 7, 12, compared to similar parameters of control plants (Figure 2).

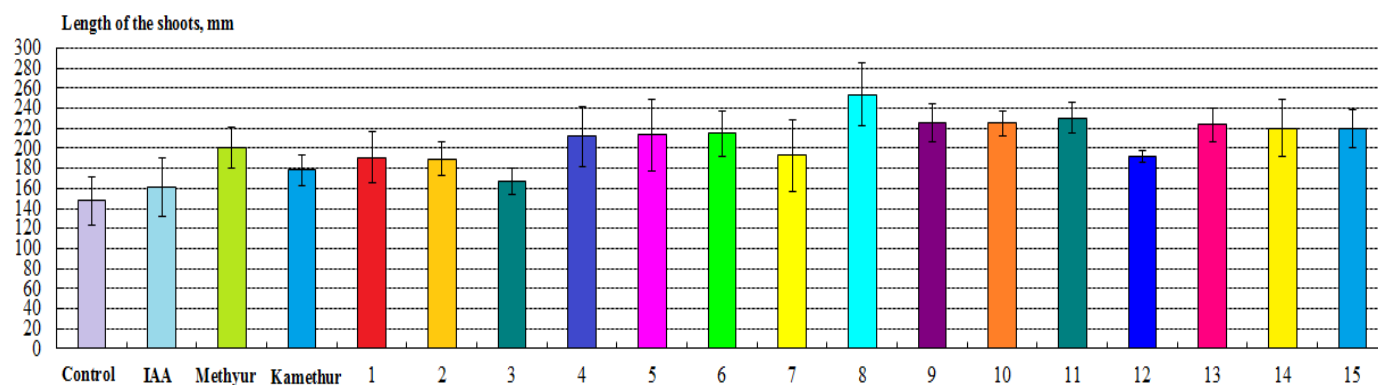


Figure 2: The regulatory effect of auxin IAA, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), and pyrimidine derivatives № 1-15 at a concentration of 10^{-6} M on the average shoot length (mm) of 4-week-old winter wheat (*T. aestivum* L.) variety Taira compared to control plants.

The highest regulatory effect on the average root length (mm) is exhibited by derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur) and pyrimidine derivatives № 1, 3, 4, 9, 10, 15, under the effect of which these parameters increase:

by 133.33% - under the effect of Methyur, by 118.61% - under the effect of Kamethur, by 92.56–113.11 % - under the effect of pyrimidine derivatives № 1, 3, 4, 9, 10, 15, compared to similar parameters of control plants (Figure 3).

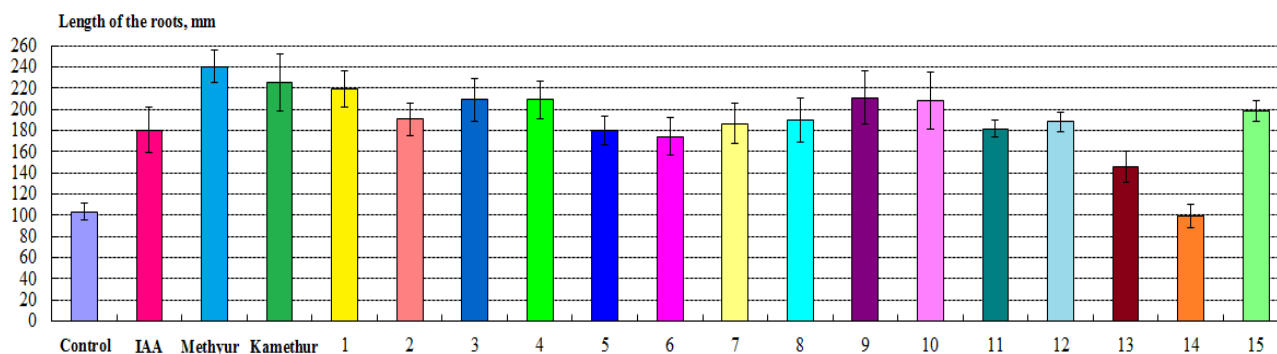


Figure 3: The regulatory effect of auxin IAA, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), and pyrimidine derivatives № 1-15 at a concentration of 10^{-6} M on the average root length (mm) of 4-week-old winter wheat (*T. aestivum* L.) variety Taira compared to control plants.

The lower regulatory effect on the average root length (mm) is exhibited by auxin IAA and pyrimidine derivatives № 2, 5, 6, 7, 8, 11, 12, 13, 14, under the effect of which these parameters increase: by 75.24% - under the effect of IAA, by 41.59–84.95% - under the effect of pyrimidine derivatives № 2, 5, 6, 7, 8, 11, 12, 13, 14, compared to similar parameters of control plants (Figure 3).

Summarizing the obtained data, it should be noted that the highest regulatory effect on the average shoot length (mm) and average root length (mm) was revealed by derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), and pyrimidine derivatives № 1, 3, 4, 5, 6, 8, 9, 10, 11, 15. Their regulatory effect was similar to or higher than that of auxin IAA. A lower regulatory effect on the average shoot length (mm) and average root length (mm) was found in pyrimidine derivatives № 2, 7, 12, 13, 14

Regulatory effect of pyrimidine derivatives on wheat photosynthesis

A comparative analysis of the regulatory effect of auxin IAA (1*H*-indol-3-yl) acetic acid, synthetic low-molecular-weight azaheterocyclic compounds, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), and pyrimidine

derivatives № 1–15 in a concentration of 10^{-6} M on the photosynthetic parameters of winter wheat (*T. aestivum* L.) variety Taira was carried out. Wheat photosynthetic parameters such as content of chlorophylls and carotenoids (mg/g fresh weight), measured 4 weeks after seed germination, were compared with those of control wheat grown in distilled water.

The highest regulatory effect on the content of chlorophylls and carotenoids in wheat leaves is revealed by derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur) and pyrimidine derivatives № 1, 2, 3, 11, 14, 15, under the effect of which the content of chlorophyll a increases: by 23.48% - under the effect of Methyur, by 28.1% - under the effect of Kamethur, by 25.98–65.39% - under the effect of pyrimidine derivatives № 1, 2, 3, 11, 14, 15; the content of chlorophyll b increases: by 23.67% - under the effect of Methyur, by 39.02% - under the effect of Kamethur, by 25.71–29.01% - under the effect of pyrimidine derivatives № 1, 2, 3; the content of chlorophylls a+b increases: by 23.54% - under the effect of Methyur, by 31.5% - under the effect of Kamethur, by 17.58–44.52% - under the effect of pyrimidine derivatives № 1, 2, 3, 11, 14, 15; the content of carotenoids

increases: by 18.98% - under the effect of Methyur, by 48% - under the effect of Kamethur, by 24.74–37.86% - under the effect of pyrimidine

derivatives № 1, 2, 11, 14 and 15, compared to similar parameters of control plants (Figure 4).

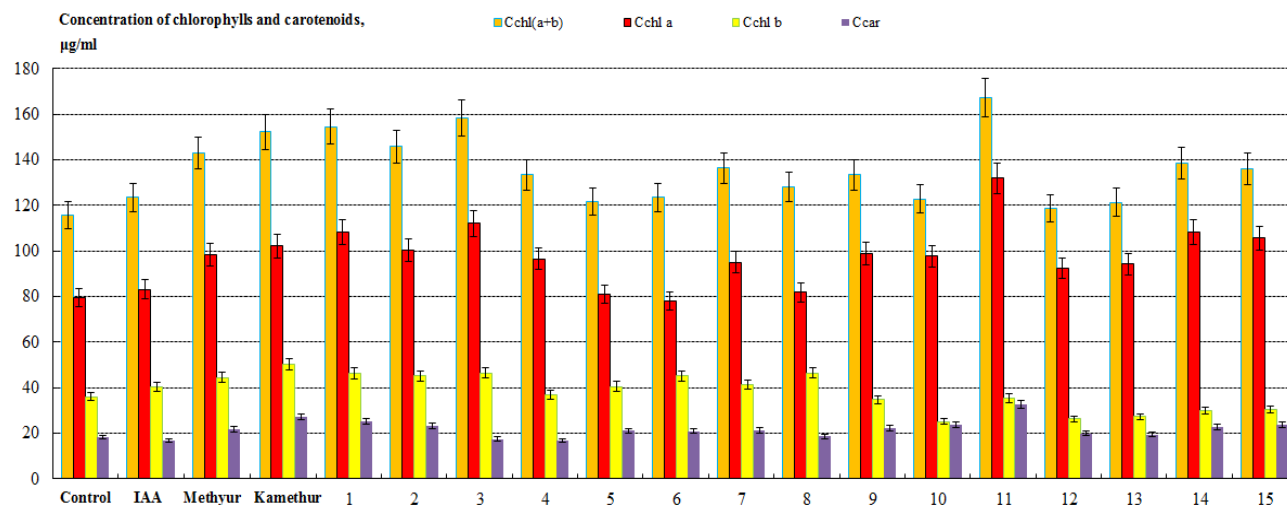


Figure 4. The regulatory effect of auxin IAA, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), and pyrimidine derivatives № 1-15 at a concentration of 10^{-6} M on the concentration of chlorophylls a, b, a+b and carotenoids (µg/ml) in the leaves of 4-week-old winter wheat (*T. aestivum* L.) variety Taira compared to control plants.

The lower regulatory effect on the content of chlorophylls and carotenoids in wheat leaves is revealed by auxin IAA and pyrimidine derivatives № 4, 5, 6, 7, 8, 9, 10, 12, 13, under the effect of which the content of chlorophyll a increases: by 4.34 % - under the effect of IAA, by 2.57–23.74% - under the effect of pyrimidine derivatives № 4, 5, 6, 7, 8, 9, 10, 12, 13; the content of chlorophyll b increases: by 11.69% - under the effect of IAA, by 2.57–28.89% - under the effect of pyrimidine derivatives № 4, 5, 6, 7, 8, 9, 10, 12, 13; the content of chlorophylls a+b increases: by 6.63% - under the effect of IAA, by 2.63–17.78% - under the effect of pyrimidine derivatives № 4, 5, 6, 7, 8, 9, 10, 12, 13; the content of carotenoids increases: by 2.5–28.81% - under the effect of pyrimidine derivatives № 4, 5, 6, 7, 8, 9, 10, 12, 13, compared to similar parameters of control plants (Figure 4).

Analyzing the relationship between the chemical structure and selectivity of regulatory effect on wheat growth parameters of synthetic compounds, pyrimidine derivatives №1-15, it can be assumed that their effect, similar to or exceeding the effect of the auxin IAA or derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methyur and Kamethur), is associated with the presence of substituents in the chemical structure of these compounds (Table 1).

The synthetic compounds, pyrimidine derivatives № 1, 3, 4, 5, 6, 8, 9, 10, 11, 15 showed the highest regulatory effect on wheat growth parameters, these compounds contain: compound №1 contains methylsulfonyl group in position 5, propyl group in position 1, phenyl group in position 3 of the 1H-pyrimidine-2,4-dione ring; compound №3 contains tolylsulfonyl group in position 5, propyl group in position 1, phenyl group in position 3 of the 1H-pyrimidine-2,4-dione ring; compound №4 contains allyl substituent in position 1, phenylsulfonyl group in position 5, phenyl group in position 3 of the 1H-pyrimidine-2,4-dione ring; compound №5 contains benzenesulfonyl group in position 5, phenyl group in position 3, butyl group in position 1 of the 1H-pyrimidine-2,4-dione ring; compound №6 contains benzenesulfonyl group in position 5, phenyl group in position 3, 3-hydroxypropyl group in position 1 of the 1H-pyrimidine-

2,4-dione ring; compound №8 contains phenylsulfonyl group in position 5, 2,3-dihydroxypropyl group in position 1, phenyl group in position 3 of the 1H-pyrimidine-2,4-dione ring; compound №9 contains benzenesulfonyl group in position 5, phenyl group in position 3, tetrahydrofuran-2-ylmethane group in position 1 of the 1H-pyrimidine-2,4-dione ring; compound №10 contains *para*-tolylsulfonyl group in position 5, phenyl group in position 1, tetrahydrofuran-2-ylmethyl group in position 1 of the 1H-pyrimidine-2,4-dione ring; compound №11 contains phenylsulfonyl group in position 5, 4-chlorobenzoic acid ethyl ester residue in position 3, 4-chlorophenyl group in position 1 of the 2,4-dioxo-3,4-dihydro-2H-pyrimidine ring; compound № 15 contains a phenyl group in position 6, a cyano group in position 5 of the 4-oxo-2-thioxo-1,2,3,4-tetrahydropyrimidine ring.

At the same time, the synthetic compounds, pyrimidine derivatives № 2, 7, 12, 13, 14, showed the lower regulatory effect on wheat growth parameters, these compounds contain: compound №2 contains benzenesulfonyl group in position 5, phenyl group in position 3, propyl group in position 1 of the 1H-pyrimidine-2,4-dione ring; compound №7 contains *para*-tolylsulfonyl group in position 5, phenyl group in position 1, 3-hydroxypropyl group in position 1 of the 1H-pyrimidine-2,4-dione ring; compound №12 contains a benzenesulfonyl group in position 5, an ethyl group in position 3 of the 2-thioxo-2,3-dihydro-1H-pyrimidin-4-one ring; compound №13 contains an allyl substituent in position 3, a phenylsulfonyl group in position 5 of the 2-thioxo-2,3-dihydro-1H-pyrimidin-4-one ring; compound №14 contains a phenyl group in position 3, a benzenesulfonyl group in position 5 of the 2-thioxo-2,3-dihydro-1H-pyrimidin-4-one ring.

The selected most physiologically active synthetic compounds, pyrimidine derivatives № 1, 3, 4, 5, 6, 8, 9, 10, 11, 15 that stimulate wheat growth and increase photosynthesis are promising for use in agricultural practice.

The results obtained in this work correlate with the data of our previous studies, which indicate that synthetic low-molecular-weight

azaheterocyclic compounds, derivatives of sodium and potassium salts of 6-methyl-2-mercapto-4-hydroxypyrimidine (Methur and Kamethur), and other pyrimidine derivatives, have a stimulating effect on the growth and development of different wheat varieties, similar to the phytohormones auxins and cytokinins, enhance photosynthesis in wheat leaves and increase wheat productivity [63, 64, 79-84]. Based on the obtained data, it was suggested that the regulatory effect of synthetic compounds, pyrimidine derivatives on wheat growth is due to their specific auxin- and cytokinin-like regulatory effect on the proliferation, elongation and differentiation of root and shoot meristem cells during the plant embryogenesis and vegetative phase, improved metabolism, increased protein biosynthesis and prevention of degradation of chlorophylls and carotenoids in plant leaves, which play an important role in plant productivity [25-33, 85-87]. It was also concluded that synthetic compounds, pyrimidine derivatives regulate plant growth similarly to synthetic auxin or cytokinin analogues, through auxin or cytokinin signaling pathways or by increasing the level of endogenous auxins and cytokinins in plants by modulating the activity of key enzymes involved in the biosynthesis, transport, metabolism, conjugation and oxidation of endogenous auxins and cytokinins in plant cells [34-42, 88-99].

Conclusions

A comparative analysis of the regulatory effect of pyrimidine derivatives and auxin IAA on the growth and photosynthesis of wheat (*T. aestivum* L.) variety Taira in the vegetative phase was carried out. A significant intensification of growth of wheat shoots and roots, as well as photosynthesis in wheat leaves, was observed under the regulatory effect of pyrimidine derivatives compared to control wheat grown in distilled water or wheat grown under the regulatory effect of auxin IAA. A correlation has been found between the chemical structure and the selectivity of the regulatory action of synthetic compounds, pyrimidine derivatives. The obtained data indicate a similar effect of pyrimidine derivatives with phytohormones auxins and cytokinins on the growth and photosynthesis of wheat. This fact confirms the prospects of practical application of selected most physiologically active synthetic compounds, pyrimidine derivatives № 1, 3, 4, 5, 6, 8, 9, 10, 11, 15 in agriculture as new wheat growth regulators.

Statement of conflict of interest:

The authors are declared that they have no conflict with this research article.

References

- Shiferaw B., Smale M., Braun H.J., Duveiller E., Reynolds M., Muricho G. (2013). Crops that feed the world. Past successes and future challenges to the role played by wheat in global food security. *Food Sec.* 5(3): 291-317.
- Reynolds M.P., Braun H.J. (Eds.). (2022). *Wheat Improvement. Food Security in a Changing Climate.* Springer, Cham. 629 p.
- Shewry P.R., Hey S.J. (2015). The contribution of wheat to human diet and health. *Food Energy Secur.* 4(3):178-202.
- Pathak V., Shrivastav S. (2015). Biochemical studies on wheat (*Triticum aestivum* L.). *Journal of Pharmacognosy and Phytochemistry.* 4(3):171-175.
- Javid Iqbal M., Shams N., Fatima K. (2022). Nutritional Quality of Wheat. In: *Wheat - Recent Advances* / Ed. Mahmood-ur-Rahman Ansari. IntechOpen, 342 p.
- Elahi I., Saeed U., Wadood A., Abbas A., Nawaz H., Jabbar S. (2022). Effect of Climate Change on Wheat Productivity. In: *Wheat - Recent Advances* / Ed. Mahmood-ur-Rahman Ansari. IntechOpen, 342 p.
- Akram S., Ghaffar M., Wadood A., Abdur Rehman Arif M. (2022). Development of Better Wheat Plants for Climate Change Conditions. In: *Wheat - Recent Advances* / Ed. Mahmood-ur-Rahman Ansari. IntechOpen, 342 p.
- Sabagh A.E., Islam M.S., Skalicky M., Ali Raza M., Singh K., Anwar Hossain M., Hossain A., Mahboob W., Iqbal M.A., Ratnasekera D., Singhal R.K., Ahmed S., Kumari A., Wasaya A., Sytar O., Brestic M., ÇIG F., Erman M., Habib Ur Rahman M., Ullah N. and Arshad A. (2021). Salinity Stress in Wheat (*Triticum aestivum* L.) in the Changing Climate: Adaptation and Management Strategies. *Front. Agron.* 3: 661932.
- Mu Q., Xu J., Yu M., Guo Z., Dong M., Cao Y., Zhang S., Sun S., Cai H. (2022). Physiological response of winter wheat (*Triticum aestivum* L.) during vegetative growth to gradual, persistent and intermittent drought. *Agricultural Water Management.* 274: 107911.
- Tiwari V. and Shoran J. Growth and production of wheat. Soils, plant growth and crop production. Vol. I. *Encyclopedia of Life Support Systems (EOLSS).*
- Pal P., Ansari S.A., Jalil S.U., Ansari M.I. (2023). Regulatory role of phytohormones in plant growth and development. Chapter 1. Pp. 1-13. In: *Plant Hormones in Crop Improvement.* Khan M. I.R., Singh A., Poór P. (Eds.). London, San Diego, Cambridge, Kidlington: Academic Press. 358 p.
- Ahammed G.J., Yu J. (Eds.) *Plant Hormones and Climate Change.* Springer Nature, Singapore. 2023. 372 p.
- Shang Q., Wanga Y., Tang H., Sui N., Zhang X., Wanga F. (2021). Genetic, hormonal, and environmental control of tillering in wheat. *The Crop Journal.* 9: 986-991.
- Lalarukh I., Arslan A.M., Azeem M., Akbar M., Yasin A., Tariq M. & Iqbal N. (2014). Growth stage-based response of wheat (*Triticum aestivum* L.) to kinetin under water-deficit environment: pigments and gas exchange attributes. *Acta Agriculturae Scandinavica, Section B — Soil & Plant Science.* 64(6): 501-510.
- Zhang H., Sun X., Dai M. (2022). Improving crop drought resistance with plant growth regulators and rhizobacteria: Mechanisms, applications, and perspectives. *Plant Communications.* 3(1): 100228.
- Blyuss K.B., Fatehi F., Tsygankova V.A., Biliavska L.O., Iutynska G.O., Yemets A.I., Blume Y.B. (2019). RNAi-based biocontrol of wheat nematodes using natural polycapillary biostimulants. *Frontiers in Plant Science.* 10: 483.
- Tsygankova V.A., Blyuss K.B., Shysha E.N., Biliavska L.A., Iutynska G.A., Andrushevich Ya.V., Ponomarenko S.P., Yemets A.I. and Blume Ya.B. (2020). Using Microbial Biostimulants to Deliver RNA Interference in Plants as an Effective Tool for Biocontrol of Pathogenic Fungi, Parasitic Nematodes and Insects. Chapter 6. P. 205-319. In: "Research Advances in Plant biotechnology". Series: *Plant Science Research and Practices* / Ed. Ya.B. Blume. USA: Nova Science Publishers, Inc., 270 p.
- Tsygankova V.A., Spivak S.I., Shysha E.N., Pastukhova N.L., Biliavska L.A., Iutynska G.A., Kyrylenko V.M., Yemets A.I., Blume Ya.B. (2023). The role of polycapillary biostimulants

- in increasing plant resistance to the biotic and abiotic stress factors. Chapter 1. Pp. 1 – 86. In: Agricultural Research Updates. Vol. 46. Editor(s): **Prathamesh Gorawala and Srushti Mandhatri**. Nova Science Publishers, Inc., NY, USA. 307 p.
19. Farhat F., Arfan M., Wang X., Tariq A., Kamran M., Tabassum H.N., Tariq I., Mora-Poblete F., Iqbal R., El-Sabroun A.M. and Elansary H.O. (2022). The Impact of Bio-Stimulants on Cd-Stressed Wheat (*Triticum aestivum* L.): Insights into Growth, Chlorophyll Fluorescence, Cd Accumulation, and Osmolyte Regulation. *Front. Plant Sci.* 13:850567.
 20. Kasim W.A., Osman M.E., Omar M.N., El-Daim I.A.A., Bejai S., Meijer J. (2013) Control of Drought Stress in Wheat Using Plant-Growth-Promoting Bacteria. *J Plant Growth Regul.* 32(1): 122-130.
 21. Lamlom S.F., Irshad A., Mosa W.F.A. (2023). The biological and biochemical composition of wheat (*Triticum aestivum*) as affected by the bio and organic fertilizers. *BMC Plant Biol.* 23: 111.
 22. Fageria N.K. (2013). The Role of Plant Roots in Crop Production. 1st Edition. CRC Press, Taylor @ Francis Group, LLC, NY, 467 p.
 23. Khan M.A., Gemenet D.C. and Villordon A. (2016). Root System Architecture and Abiotic Stress Tolerance: Current Knowledge in Root and Tuber Crops. *Front. Plant Sci.* 7:1584.
 24. Anbarasan S. and Ramesh S. (2021). The Role of Plant Roots in Nutrient Uptake and Soil Health. *Plant Science Archives.* 05-08.
 25. Su Y.H., Liu Y.B., Zhang X.S. (2011). Auxin–Cytokinin Interaction Regulates Meristem Development. *Molecular Plant.* 4(4): 616 – 625.
 26. Schaller G.E., Bishopp A., Kieber J.J. (2015). The Yin-Yang of Hormones: Cytokinin and Auxin Interactions in Plant Development. *Plant Cell.* 27: 44–63.
 27. Lee Z.H., Hirakawa T., Yamaguchi N., Ito T. (2019). The Roles of Plant Hormones and Their Interactions with Regulatory Genes in Determining Meristem Activity. *Int J Mol Sci.* 20(16):4065.
 28. Sosnowski J, Truba M, Vasileva V. (2023). The Impact of Auxin and Cytokinin on the Growth and Development of Selected Crops. *Agriculture.* 13(3): 724.
 29. Tsygankova V.A. (2015). Genetic Control and Phytohormonal Regulation of Plant Embryogenesis. *Int. J. Med. Biotechnol. Genetics (IJMBG).* 3(1): 9-20.
 30. Mukherjee A., Gaurav A.K., Singh S., Yadav S., Bhowmick S., Abeyasinghe S., Verma J.P. (2022). The bioactive potential of phytohormones: A review. *Biotechnology Reports.* 35: e00748.
 31. Ljung K. (2013). Auxin metabolism and homeostasis during plant development. *Development.* 140(5): 943–950.
 32. Hu Y. and Shani E. (2023). Cytokinin activity – transport and homeostasis at the whole plant, cell, and subcellular levels. *New Phytologist.* 239: 1603 - 1608.
 33. Mok D.W.S. and Mok M.C. (2001). Cytokinin metabolism and action. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 52: 89 – 118.
 34. Novickienė L., Asakavičiūtė R. (2006). Analogues of auxin modifying growth and development of some monocot and dicot plants. *Acta Physiol Plant.* 28(6): 509–515.
 35. Fukui K., Hayashi K., (2018). Manipulation and Sensing of Auxin Metabolism, Transport and Signaling. *Plant and Cell Physiology.* 59(8): 1500–1510.
 36. Nowicka B. (2022). Modifications of Phytohormone Metabolism Aimed at Stimulation of Plant Growth, Improving Their Productivity and Tolerance to Abiotic and Biotic Stress Factors. *Plants.* 11(24): 3430.
 37. Savaldi-Goldstein S., Baiga T.J., Pojer F., Dabi T., Butterfield C., Parry G., Santner A., Dharmasiri N., Tao Y., Estelle M., Noel J.P., Chory J. (2008). New auxin analogs with growth-promoting effects in intact plants reveal a chemical strategy to improve hormone delivery. *Proc Natl Acad Sci USA.* 105(39): 15190-5.
 38. Rigal A., Ma Q., Rober S. (2014). Unraveling plant hormone signaling through the use of small molecules. *Frontiers in Plant Science.* 5(Article 373): 1–20.
 39. Vylčilová H., Bryksová M., Matušková V., Doležal K., Plíhalová L., Strnad M. (2020). Naturally Occurring and Artificial N9-Cytokinin Conjugates: From Synthesis to Biological Activity and Back. *Biomolecules.* 10(6): 832.
 40. Jameson P. E. Zeatin: The 60th anniversary of its identification. *Plant Physiology.* 2023. 192(1): 34 – 55.
 41. Podlešáková K., Zalabák D., Čudejčková M., Plíhal O., Szűčová L., Doležal K, et al. (2012). Novel Cytokinin Derivatives Do Not Show Negative Effects on Root Growth and Proliferation in Submicromolar Range. *PLoS ONE.* 7(6): e39293.
 42. Naseem A., Mohammad F. (2020). Thidiazuron: From Urea Derivative to Plant Growth Regulator. Singapore: Springer, 2018. 491 p.
 43. Geelen D. and Xu L. (Eds). The Chemical Biology of Plant Biostimulants. Book Series Wiley Series in Renewable Resources. New Jersey, USA: John Wiley & Sons Ltd. 305 p.
 44. Tsygankova V.A., Brovarets V.S., Yemets A.I., Blume Y.B. Prospects of the development in Ukraine of the newest plant growth regulators based on low molecular heterocyclic compounds of the azole, azine and their condensed derivatives. P. 246 – 285. In Book: Synthesis and bioactivity of functionalized nitrogen-containing heterocycles / Eds. A.I. Vovk. Kyiv: Interservice, 2021.
 45. Tsygankova V.A., Andrushevich Ya.V., Shtompel O.I., Solomyanny R. M., Hurenko A.O., Frasinuk M.S., Mrug G.P., Shablykin O.V., Pilyo S.G., Kornienko A.M. & Brovarets V. S. (2022). New Auxin and Cytokinin Related Compounds Based on Synthetic Low Molecular Weight Heterocycles, Chapter 16, In: Aftab T. (Ed.) Auxins, Cytokinins and Gibberellins Signaling in Plants, Signaling and Communication in Plants, Springer Nature Switzerland AG, 353-377.
 46. Farghaly T.A., Harras M.F., Alsaedi A.M.R., Thakir H.A., Mahmoud H.K., Katowah D.F. (2023). Antiviral Activity of Pyrimidine Containing Compounds: Patent Review. *Mini Rev Med Chem.* 23(7): 821-851.
 47. Somkuwar S., Chaubey N. (2023). Pyrimidine derivatives: Their significance in the battle against malaria, cancer and viral infections. *GSC Biological and Pharmaceutical Sciences* 25(2): 076-083.
 48. Patil S.B. (2023). Recent medicinal approaches of novel pyrimidine analogs: A review. *Heliyon.* 9(6):e16773.

49. Finger V., Kufa M., Soukup O., Castagnolo D., Roh J., Korabecny J. (2023). Pyrimidine derivatives with antitubercular activity. *Eur J Med Chem.* 246: 114946.
50. Khalid T., Kalsoom S., Anwar S., Farrukh A., Gao L., Jafri L., et al. (2024). Molecular Docking, Synthesis and Anti-diabetic Studies of Pyrimidine Derivatives. *Ann Pharmacol Pharm.* 9(1): 1211.
51. Verma P.K., Bhutani G., Saini R. and Rani S. (2016). Experimental evaluation of effects of a pyrimidine derivative 4CPTP on cardiovascular system. *International Journal of Biomedical and Advance Research.* 7(9): 448-451.
52. Amr A.G., Mohamed A.M., Mohamed S.F., Abdel-Hafez N.A., Hammam Ael-F. (2006). Anticancer activities of some newly synthesized pyridine, pyrane, and pyrimidine derivatives. *Bioorg Med Chem.* 14(16): 5481-5488.
53. Ota C., Kumata S., Kawaguchi S. Novel herbicides, usage thereof, novel thienopyrimidine derivatives, intermediates of the same, and process for production thereof. Patent US20070010402A1. 2007.
54. Cansev A., Gülen H., Zengin M.K., Ergin S., Cansev M. Use of Pyrimidines in Stimulation of Plant Growth and Development and Enhancement of Stress Tolerance. WIPO Patent WO 2014/129996A1. 2014.
55. Boussemghoune M.A., Whittingham W.G., Winn C.L., Glithro H., Aspinall M.B. Pyrimidine derivatives and their use as herbicides. Patent US20120053053 A1. 2012.
56. Cansev A., Gülen H., Zengin M.K., Ergin S., Cansev M. (2014). Use of Pyrimidines in Stimulation of Plant Growth and Development and Enhancement of Stress Tolerance, WIPO Patent WO 2014/129996A1.
57. Li J.H., Wang Y., Wu Y.P., Li R.H., Liang S., Zhang J., Zhu Y.G., Xie B.J. (2021). Synthesis, herbicidal activity study and molecular docking of novel pyrimidine thiourea. *Pestic Biochem Physiol.* 172: 104766.
58. Wang D.W., Li Q., Wen K., Ismail I., Liu D.D., Niu C.W., Wen X., Yang G.F., Xi Z. (2017). Synthesis and Herbicidal Activity of Pyrido[2,3-d]pyrimidine-2,4-dione-Benzoxazinone Hybrids as Protoporphyrinogen Oxidase Inhibitors. *J Agric Food Chem.* 65(26): 5278 - 5286.
59. Kamal El-Dean A. M., Abd-Ella A. A., Hassanien R., El-Sayed M. E. A., Zaki R. M., and Abdel-Raheem Sh.A.A. (2019). Chemical design and toxicity evaluation of new pyrimidothienotetra hydroisoquinolines as potential insecticidal agents. *Toxicol. Rep.* 6: 100-104.
60. Kut M., Kut D., Mariychuk R.T. (2024). Synthesis of N-alkenyl(alkynyl)-5,6-dimethyl-2-(thiophen-2-yl)thieno[2,3-d]pyrimidin-4-amines and their *in silico* study on group II chitinase (ChtII) inhibition. *Sci. Bull. Uzhh. Univ. Ser. Chem.* 2(52): 75-82.
61. Tsygankova V.A., Voloshchuk I.V., Kopich V.M., Pilyo S.G., Klyuchko S. V., Brovarets V.S. Studying the effect of plant growth regulators Ivin, Methyur and Kamethur on growth and productivity of sunflower. (2023). *Journal of Advances in Agriculture.* 14: 17–24.
62. Tsygankova V.A., Voloshchuk I.V., Pilyo S.H., Klyuchko S.V., Brovarets V.S. (2023). Enhancing Sorghum Productivity with Methyur, Kamethur, and Ivin Plant Growth Regulators. *Biology and Life Sciences Forum.* 27(1): 36.
63. Tsygankova V.A., Kopich V.M., Vasylenko N.M., Golovchenko O.V., Pilyo S.G., Malienco M.V., Brovarets V.S. (2024). Increasing the productivity of wheat using synthetic plant growth regulators Methyur, Kamethur and Ivin. *Znanstvena misel journal.* 94: 22 - 26.
64. Tsygankova V.A., Andreev A.M., Andrushevich Ya.V., Pilyo S.G., Klyuchko S.V., Brovarets V.S. (2023). Use Of Synthetic Plant Growth Regulators In Combination With Fertilizers to Improve Wheat Growth. *Int J Med Biotechnol Genetics.* S1:02:002:9-14.
65. Tsygankova V.A., Andreev A.M., Andrushevich Ya.V., Pilyo S.G., Brovarets V.S. (2023). Effect of plant growth regulators and fertilizers on the vegetative growth of sunflower (*Helianthus annuus* L.). *The scientific heritage.* 116(116): 3–9.
66. Tsygankova V.A., Andreev A.M., Andrushevich Ya.V., Kopich V.M., Klyuchko S.V., Pilyo S.G., Brovarets V.S. (2023). Use of Ivin, Methyur, Kamethur and microfertilizers to improve the growth of oilseed flax (*Linum usitatissimum* L.). *Annali d'Italia.* 48: 3-10.
67. Tsygankova V.A., Andreev A.M., Andrushevich Ya.V., Kopich V.M., Pilyo S.G., Klyuchko S.V., Brovarets V.S. (2023). Synergistic effect of synthetic plant growth regulators and microfertilizers on the growth of canola (*Brassica napus* L.). *Danish Scientific Journal (DSJ).* 1(77): 8 - 12.
68. Tsygankova V.A., Kopich V.M., Voloshchuk I.V., Pilyo S.G., Klyuchko S. V., Brovarets V.S. (2023). New growth regulators of barley based on pyrimidine and pyridine derivatives. *Sciences of Europe.* 124: 13 – 23.
69. Tsygankova V.A., Andrushevich Ya.V., Vasylenko N.M., Pilyo S.G., Klyuchko S.V., Brovarets V.S. (2023). Screening of Auxin-like Substances among Synthetic Compounds, Derivatives of Pyridine and Pyrimidine. *J Plant Sci Phytopathol.* 7: 151-156.
70. Tsygankova V.A., Andrushevich Ya.V., Kopich V.M., Voloshchuk I.V., Pilyo S.G., Klyuchko S. V., Brovarets V.S. (2023). Application of pyrimidine and pyridine derivatives for regulation of chickpea (*Cicer arietinum* L.) growth. *International Journal of Innovative Science and Research Technology (IJISRT).* 8(6): 19 – 28.
71. Tsygankova V.A., Andrushevich Ya.V., Kopich V.M., Voloshchuk I.V., Bondarenko O.M., Pilyo S.G., Klyuchko S.V., Brovarets V.S. (2023). Effect of pyrimidine and pyridine derivatives on the growth and photosynthesis of pea microgreens. *Int J Med Biotechnol Genetics (IJMBG).* S1:02:003:15-22.
72. Tsygankova V.A., Kopich V.M., Vasylenko N.M., Andrushevich Ya.V., Pilyo S.G., Brovarets V.S. (2024). Phytohormone-like effect of pyrimidine derivatives on the vegetative growth of haricot bean (*Phaseolus vulgaris* L.). *Polish Journal of Science.* 1(71): 6–13.
73. Tsygankova V.A., Andrushevich Ya.V., Vasylenko N.M., Kopich V.M., Solomyanni R.M., Popilnichenko S.V., Kozachenko O.P., Pilyo S.G., Brovarets V.S. (2024). The use of thioxypyrimidine derivatives as new regulators of growth and photosynthesis of barley. *J Plant Sci Phytopathol.* 8(2): 090-099.

74. Tsygankova V.A., Vasylenko N.M., Andrushevich Ya.V., Kopich V.M., Kachaeva M.V., Popilnichenko S.V., Pilyo S.G. and Brovarets V.S. (2025). Use of Thienopyrimidine Derivatives to Optimize Sorghum Growth and Photosynthesis during the Vegetation Period. *Journal of Biomedical Research & Environmental Sciences*. 6(1): 071 - 080.
75. Voytsehovska O.V., Kapustyan A.V., Kosik O.I. (2010). *Plant Physiology: Praktykum*, Parshikova T.V. (Ed.), Lutsk: Teren, 420 p.
76. Lichtenthaler H. (1987). Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. *Methods in Enzymology*. 148: 331 – 382.
77. Lichtenthaler H.K., Buschmann C. (2001). Chlorophylls and Carotenoids: Measurement and Characterization by UV-VIS Spectroscopy *Current Protocols in Food Analytical Chemistry (CPFA)*: John Wiley and Sons, New York, F4.3.1-F4.3.8.
78. Bang H., Zhou X.K., van Epps H.L., Mazumdar M. (Eds.). (2010). *Statistical Methods in Molecular Biology. Series: Methods in molecular biology*, New York: Humana press., 13(620): 636 p.
79. Tsygankova V., Vasylenko N., Andrushevich Ya., Kopich V., Kachaeva M., Popilnichenko S., Kozachenko O., Pilyo S., Brovarets V. (2024). Application of thienopyrimidine derivatives as new eco-friendly wheat growth regulators. *Sciences of Europe*. 146: 8–18.
80. Tsygankova V.A., Vasylenko N.M., Andrushevich Ya.V., Kopich V.M., Solomyannyi R.M., Pilyo S.G., Bondarenko O.M., Popilnichenko S.V., Brovarets V.S. (2024). New Wheat Growth Regulators Based On Thioxopyrimidine Derivatives. *Int J Med Biotechnol Genetics*. S1:02:004:23-30.
81. Tsygankova V.A., Andrushevich Ya.V., Vasylenko N.M., Kopich V.M., Pilyo S.G., Solomyannyi R.M., Popilnichenko S.V., Bondarenko O.M., Brovarets V.S. (2024). The use of thioxopyrimidine derivatives for the regulation of vegetative growth of wheat. *Journal of Medicinal Botany*. 8: 1-7.
82. Tsygankova V., Andrushevich Ya., Kopich V., Vasylenko N., Solomyannyi R., Popilnichenko S., Kachaeva M., Kozachenko O., Pilyo S., & Brovarets V. (2024). Wheat growth in the vegetative phase under the regulatory effect of furoypyrimidine derivatives. *The scientific heritage*. 140: 3-12.
83. Tsygankova V.A., Andrushevich Ya.V., Vasylenko N.M., Kopich V.M., Popilnichenko S.V., Pilyo S.G., Brovarets V.S. (2024). Auxin-like and cytokinin-like effects of new synthetic pyrimidine derivatives on the growth and photosynthesis of wheat. *J Plant Sci Phytopathol*. 8(1): 015–024.
84. Tsygankova V.A., Vasylenko N.M., Andrushevich Ya.V., Kopich V.M., Solomyannyi R.M., Kachaeva M.V., Bondarenko O.M., Pilyo S.G., Popilnichenko S.V., Brovarets V.S. (2025). Screening of Synthetic Auxin-Like and Cytokinin-Like Compounds, Derivatives of Thioxopyrimidine as New Plant Growth Regulators. *Significances Bioeng Biosci*. 7(2). SBB. 000657.
85. Paque S., Weijers D. (2016). Q & A: Auxin: the plant molecule that influences almost anything. *BMC Biol*. 14, 67: 1-5.
86. Hönig M., Plíhalová L., Husíčková A., Nisler J., Doležal K. (2018). Role of Cytokinins in Senescence, Antioxidant Defence and Photosynthesis. *Int J Mol Sci*. 19(12): 4045.
87. Wu W., Du K., Kang X. and Wei H. (2021). The diverse roles of cytokinins in regulating leaf development. *Hortic Res*. 8:118, 1-13.
88. Lavy M., Estelle M. (2016). Mechanisms of auxin signaling. *Development*. 143(1): 3226–3229.
89. Casanova-Sáez R., Mateo-Bonmatí E., Ljung K. (2021). Auxin Metabolism in Plants. *Cold Spring Harb Perspect Biol*. 13(3): a039867.
90. Hayashi K.I. (2021). Chemical Biology in Auxin Research. *Cold Spring Harb Perspect Biol*. 13(5): a040105.
91. Hwang I., Sheen J., Muller B. (2012). Cytokinin Signaling Networks. *Annu. Rev. Plant Biol*. 63: 353–380.
92. Kieber J.J., Schaller G.E. (2018). Cytokinin signaling in plant development. *Development*. 145(4): dev149344: 1–7.
93. Blázquez M.A., Nelson D.C., Weijers D. (2020). Evolution of Plant Hormone Response Pathways. *Annu. Rev. Plant Biol*. 71: 327-353.
94. Fàbregas N., Alisdair R. Fernie A.R. (2021). The interface of central metabolism with hormone signaling in plants. *Current Biology*. 31(23): R1535-R1548.
95. Müller K., Dobrev P.I., Pěňčík A., Hošek P., Vondráková Z., Filepová R., Malínská K., Brunoni F., Helusová L., Moravec T., Retzer K., Harant K., Novák O., Hoyerová K., Petrášek J. (2021). Dioxxygenase for auxin oxidation 1 catalyzes the oxidation of IAA amino acid conjugates. *Plant Physiol*. 187(1): 103-115.
96. Zhang J., Peer W. A. (2017). Auxin homeostasis: the DAO of catabolism. *Journal of Experimental Botany*. 68(12): 3145–3154.
97. Mellor N., Band L.R., Pěňčík A., Novák O., Rashed A., Holman T., Wilson M.H., Voß U., Bishopp A., King J.R., Ljung K, Bennett M.J., Owen M.R. (2016). Dynamic regulation of auxin oxidase and conjugating enzymes AtDAO1 and GH3 modulates auxin homeostasis. *PNAS*. 113(39): 11022–11027.
98. Hayashi Ki., Arai K., Aoi Y. *et al.* (2021). The main oxidative inactivation pathway of the plant hormone auxin. *Nat Commun*. 12: 6752.
99. Khablak S.H., Spivak S.I., Pastukhova N.L., Yemets A.I., and Blume Ya.B. (2024). Cytokinin Oxidase/Dehydrogenase as an Important Target for Increasing Plant Productivity. *Cytology and Genetics*. 58(2): 115–125.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article Click Here:

Submit Manuscript

DOI:[10.31579/2637-8914/306](https://doi.org/10.31579/2637-8914/306)

Ready to submit your research? Choose Auctores and benefit from:

- fast, convenient online submission
- rigorous peer review by experienced research in your field
- rapid publication on acceptance
- authors retain copyrights
- unique DOI for all articles
- immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <https://auctoresonline.org/journals/nutrition-and-food-processing>