Open Access

Mohamed Zaeim Hafez*

Research Article

Interleukin (6, 10): Can be used as a Prediction tool for Multiple organ Dysfunction Syndrome in Critically ill Pediatric Patients?

Waleed Abdelmoneim Mohamed Said Abdelfattah ¹, Mohamed Zaeim Hafez ²*, Mona Eid Ahmed ³, Mohamed E EL-Refaey ², Hesham Abdelrahman Mahmoud Ahmed ⁴, Abdel Rahman Z ⁵. Abdel Rahman, Tarek Shikhon ⁶

¹Pediatric Department, HMS-Algarhoud Private Hospital, Dubai, UAE.

²Physiology department, Faculty of Medicine, Al-Azhar University (Assiut), Assiut, Egypt.

³National Nutrition Institute, Cairo, Egypt.

⁴Medical Biochemistry department, Faculty of Medicine, Al-Azhar University (Assiut), Assiut, Egypt.

⁵Forensic Medicine and Clinical Toxicology Department, Faculty of Medicine, Al-Azhar University (Assiut), Assiut, Egypt.

⁶Biochemistry Department, Faculty of Medicine, Al-Azhar University (Assiut), Assiut, Egypt.

*Corresponding Author: Mohamed Zaeim Hafez, Physiology Department, Faculty of Medicine, Al-Azhar University (Assiut), Assiut, Egypt.

Received date: December 06, 2024; Accepted date: December 16, 2024; Published date: December 23, 2024

Citation: Mohamed Said Abdelfattah WAM, Mohamed Z. Hafez, Mona E. Ahmed, Mohamed E EL-Refaey, Mahmoud Ahmed, et a, (2024), Interleukin (6, 10): Can be used as a Prediction tool for Multiple organ Dysfunction Syndrome in Critically ill Pediatric Patients? *J. Clinical Pediatrics and Mother Health*, 3(1); **DOI:10.31579/2835-2971/019**

Copyright: © 2024, Mohamed Zaeim Hafez. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract:

Objectives: This study aims to determine the best cut-off level of interleukin 6,10 levels for the prediction of multiple organ dysfunction syndrome in critically ill pediatric patients.

Methods: A case-control study included 110 critical pediatric patients who were admitted to the PICU at Al Azhar University Hospitals from March 2023 to February 2024 and 68 approximate-general children matched Sex and age were recruited from other departments of our hospital as a control group. Full history taken, Thorough clinical examination, the incidences of organ dysfunctions in the studied critically ill pediatric patients, Determined the levels of Interleukin 6 and 10 measured using for both critically ill children's patients and general patients using ELISA Kit Human IL-6, 10.

Results: critically ill patients who suffered from coagulation abnormalities were more common than other incidences (n=62, 56.36%) followed by those who suffered from acute liver injury (n=59, 53.64%), Interlukine-6 and 10 levels were significantly higher in the critically ill patients (252.3 ± 171 , 185.12 ± 72.16 pg/mL respectively) than those in the general patients (64.15 ± 50.01 , 35.40 ± 11.90 pg/mL respectively), Also, interleukin-10 levels were significantly lower among children who had post-CAR T cell/chemotherapy, and post-bone marrow transplantation (Median=144.76, and 163.44 respectively). ROC (receiver operating characteristic) analysis showed that interleukin10 levels could be used as good tool for diagnosis of critically ill patients with an area under a curve of 0.879 (p <.001) at 95% CI of (0.830-0.927), using a cut-off of >239 pg/mL with 92.5% sensitivity and 79.4% specificity as compared to interleukin 6 which at 95% CI (0.968-1), using a cut-off of >178.5 pg/mL with 97.4% sensitivity and 62% specificity. In conclusion, The IL-6 and IL-10 levels were significantly different between critically ill patients with organ injury. The IL-6 and IL-10 levels were higher in critically ill patients with organ injury compared to general patients. The interleukin-10 levels could be used as a good tool for the diagnosis of critically ill patients with a cut-off of >239 pg/mL with 92.5% sensitivity and 79.4% specificity. The IL-6 and IL-10 levels were higher in critically ill patients with organ injury compared to general patients. The interleukin-10 levels could be used as a good tool for the diagnosis of critically ill patients with a cut-off of >239 pg/mL with 92.5% sensitivity and 79.4% specificity as compared to interleukin-10 levels could be used as a good tool for the diagnosis of critically ill patients with a cut-off of >239 pg/mL with 92.5% sensitivity and 79.4% specificity as compared to interleukin-10 levels could be used as a good tool for the diagnosis of critically ill patients with a cut-of

Keywords: diagnostic tool; organ dysfunction; interleukin 6, 10; Pediatric critical illness

Introduction

Pediatric critical illness encompasses severe conditions affecting the airways, breathing, circulation, or consciousness. These life-threatening

situations often arise abruptly, making early intervention crucial. (Kortz et al., 2022). While the global impact of pediatric acute critical illness is significant, its true extent remains undetermined due to various

challenges (**Ruth et al., 2020**). Pediatric Multiple Organ Dysfunction Syndrome (MODS) is a serious condition characterized by the simultaneous impairment of two or more organs. This systemic inflammatory response can lead to widespread organ dysfunction (**Arias et al., 2024**).

Pediatric multiple organ dysfunction syndrome (MODS) has been defined as the presence of two or more concurrent organ dysfunctions (**Badke et al., 2022; Southard-Goebel et al., 2023**). The pathophysiology of MODS is characterized by a severe, systemic, uncontrolled inflammatory process that leads to multiple organ system dysfunctions (**Fan et al., 2022**). Various underlying conditions can contribute to MODS, with sepsis being the most common. Other risk factors include trauma, burns, pancreatitis, metabolic disorders, and post-transplant complications (**Bose et al., 2021**). MODS is a significant concern in pediatric intensive care, affecting over 25% of admissions and contributing to a substantial number of deaths (**Sanchez-Pinto et al., 2020; Killien et al., 2022; Gaugler et al., 2023**).

Excess humoral mediators leak into the bloodstream, triggering a systemic inflammatory response syndrome (SIRS). This dysregulated immune response is a key factor in the development of multiple organ dysfunction (**Balkrishna et al., 2023**). Early prediction of MOD could enhance critical care and patient outcomes. (**Ishikawa et al., 2021**). A cytokine storm, characterized by the excessive release of various cytokines, plays a crucial role in the inflammatory response and the development of MOD in critically ill patients (**Shimazui et al., 2021**).

The interleukin-1 (IL-1) family of cytokines and their receptors are unique players in the immune system (**Migliorini et al., 2020**). They are primarily linked to innate immunity and form a complex network regulating innate and adaptive immune responses (**Teufel et al., 2022**). This diversity allows IL-1 family members to influence a wide range of immune responses, from maintaining homeostasis to contributing to various diseases such as cancer, autoimmune disorders, infections, inflammation, and metabolic dysfunctions (**Behzadi et al., 2022**, **Fernández-Sarmiento et al., 2022**). Elevated plasma levels of IL-1 and nitric oxide metabolites have been associated with increased mortality in pediatric MODS (**Broderick and Hoffman, 2022**). Additionally, genetic variations in TNF- α , IL-1 β , and IL-1ra have been linked to the severity of sepsis (**Liang et al., 2024**).

Interleukin-6 (IL-6) is a crucial immune regulator involved in various diseases, including autoimmune disorders, chronic inflammation, and cancer (**Rose-John et al., 2023**). Normally, IL-6 levels in the blood are very low, but they can dramatically increase during inflammation or severe conditions like sepsis (**Rose-John, 2023**). IL-6 is produced by immune cells in response to infection or injury, and its production is amplified by other inflammatory cytokines like IL-1 β and TNF- α . High levels of IL-6 are associated with severe illness and the risk of multiple organ failure (**Andruszkow et al., 2014, Gidon et al., 2021, Akkaya et al., 2022**). This study aims to identify the optimal IL-6 level that can accurately predict the development of multiple organ dysfunction syndrome in critically ill children.

Patients and Methods

Study design

A case-control study was conducted involving 110 critically ill pediatric patients admitted to the PICU at Al Azhar University Hospitals between March 2023 and February 2024. A control group of 68 children, matched

for sex and age, was recruited from other departments within the same hospital.

Patients' selection criteria

This study included critically ill children aged 1 month to 18 years who met the criteria for severe disease and were expected to stay in the PICU for more than 7 days. Complete clinical information was essential for inclusion. Children were excluded if they refused blood tests, were receiving immunomodulatory drugs prior to hospitalization, had a preexisting inflammatory disease, or lacked parental consent.

Patients diagnoses

The diagnosis of multiple organ dysfunction syndrome (MODS) in the study participants was made based on international guidelines and/or criteria established by the Chinese Abdominal Intensive Care Association, (2021), McDonag et al. (2021), Pickkers et al. (2021), Liu et al. (2022).

All patients were subjected to the following:

Full history datils included the history of age, sex, present illness, history, nutritional history, and the presence of chronic illness e.g. a genetic syndrome predisposing to mechanical ventilation and VAP.

Thorough clinical examination: This included local and general examination with emphasis on detecting any change in chest examination and respiratory secretions or change in vital signs. Initial leukemia, relapsed leukemia, Post-CAR T cell/chemotherapy, post-bone marrow transplantation, sepsis, severe pneumonia, severe encephalitis, myocarditis, arthritis and hemophagocytic syndrome, and solid tumors.

The incidences of organ dysfunctions in the studied critically ill pediatric patients such as acute gastrointestinal injury (AGI), acute respiratory distress syndrome (ARDS), acute liver injury (ALI), acute liver failure (ALF), acute kidney injury (AKI), and acute myocardial injury (AMI).

Determined the levels of Interleukin 6 and 10 measured using as Sandwich ELISA for both critically ill children's patients and general patients using ELISA Kit Human IL-6, 10 (Fine Biotech comp, Wuhan, 430074, Hubei, China).

Outcomes of the study

This study aimed to evaluate the levels of interleukin-6 (IL-6) and interleukin-10 (IL-10) in critically ill pediatric patients and assess their association with clinical findings and organ dysfunction. The primary objective was to determine the optimal cutoff values for IL-6 and IL-10 to predict the development of multiple organ dysfunction syndrome (MODS) in this population.

Ethics approval and consent to participate.

Informed consent was obtained from parents or caregivers of all study participants, who were informed about the potential benefits and risks of the study. The study protocol was approved by the ethical committee of Faculty of Medicine, Al-Azhar University (Assiut), (RESERACH/AZ.AST./5/207/12/2024) and was conducted in accordance with the Declaration of Helsinki and relevant national and institutional guidelines.

Sample size estimation

Indeed, Zhu et al. et al. (2024) reported that the best cutoff value for IL-10 was 279.45 pg/mL and IL-6 was 1,425.6 pg/mL for predicting cardiac

arrest patients with a sensitivity of 0.875 and 0.625, respectively. To detect the null hypothesis, a sample size of 178 child patients (critically ill children's patients and general patients) would be required, with a drop-out rate of 10% and a research power of 80%.

Statistical analysis

The statistical analysis was performed using Microsoft Excel 2019 and SPSS v. 25. Descriptive statistics, including relative frequencies, were used to summarize categorical variables. To compare groups, independent t-tests, Mann-Whitney U tests, and Chi-square tests (including Fisher's exact test and Monte Carlo simulations) were employed. Receiver operating characteristic (ROC) curve analysis was utilized to determine the optimal cutoff values for IL-6 and IL-10 in predicting MODS. Statistical significance was set at a p-value of less than 0.05.

Results

In our study, Figure 1 depicts a flowchart of 186 pediatric patients. 8 patients were excluded from the research, with 5 of them not meeting inclusion criteria and 3 patients declined to participate. Out of the 178 patients who were willing to participate, 110 Critically ill children's patients and 68 general patients matched age and sex as the control group, (Figure 1).

In our study between the critically ill patients and general patients according to age and gender, we observed that there was no significant difference among the studied groups (P>0.05), (Table 1).

In the current study according to initial leukemia, relapsed leukemia, postcart cell/chemotherapy, post-bone marrow transplantation, sepsis, severe pneumonia, severe encephalitis, myocarditis, arthritis, and hemophagocytic syndrome, and solid tumors. there was no significant difference between the studied groups (P>0.05), (Table 2)



Figure 1: Flowchart of the studied pediatric patients.

	Group C (n=110)	Group G (n=68)	test	P value
Age/year				
Mean ±SD	6.28 ± 2.09	6.81±2.71	t=49.50	0.142
Median (Range)	6 (2.35-11.41)	6.90 (1.80-12.02)		
Gender				
Male	71(64.55%)	43 (63.24%)	X ² =0.031	0.860
Female	39 (35.45%)	25 (36.76%)		

Group C (critically ill patients), Group G (general patients), Independent t-test (t), Chi-square test (X²)

Copy rights @ Mohamed Zaeim Hafez. et al,

Table 1: Demographic data among the studied patients.

	Group C (n=110)	Group G (n=68)	X ²	P value
Initial leukemia	7 (6.36%)	6 (8.82%)	0.37	0.540
Relapsed leukemia	10 (9.09%)	4 (5.88%)	FE=0.59	0.441
Post-CAR T cell/chemotherapy	22 (20%)	17 (25%)	0.61	0.433
Post-bone marrow transplantation	5 (4.55%)	3 (4.41%)	FE=0.002	0.967
Sepsis	28 (25.45%)	16 (23.53%)	0.08	0.772
Severe pneumonia	1 (0.91%)	0	FE=0.61	0.432
Severe encephalitis	20 (18.18%)	7 (10.29%)	2.03	0.154
Myocarditis	3 (2.73%)	2 (2.94%)	FE=0.007	0.933
Arthritis and hemophagocytic syndrome	19 (17.27%)	8 (11.76%)	0.99	0.320
Solid tumors	1 (0.91%)	0	FE=0.61	0.432

Group C (critically ill patients), Group G (general patients), Chi-square test (X²), Fisher's exact test (FE)

Table 2: Thorough clinical examination among the studied patients.

In the current study, critically ill patients who suffered from coagulation abnormalities were more common than other incidences (n=62, 56.36%) followed by those who suffered from acute liver injury (n=59, 53.64%), also, acute myocardial injury, acute gastrointestinal injury, neurological abnormalities, and acute kidney injury were observed in 43 (39.09%), 37

(33.63%), 35 (31.81%) and 23 (20.9%) critically ill patients respectively, while, the lowest common were Cardiac arrest (n=5, 5.54%), acute liver failure (n=4, 3.63%), and acute respiratory distress syndrome (n=2, 1.81%), (Figure 2).



Figure 2: The incidences of organ dysfunction in the studied critically ill pediatric patients. Group C (critically ill patients), Group G (general patients), Coagulation abnormalities (CAN), acute liver injury (ALI), acute myocardial injury (AMI), acute gastrointestinal injury (AGI), Neurological abnormalities (NAN), acute kidney injury (AKI), Cardiac arrest (CA), acute liver failure (ALF), acute respiratory distress syndrome (ARDS).

In the present study, there was a significant difference between the studied patient groups regarding interleukin-6 and 10 levels (p<0.001). Interlukine-6 and 10 levels were significantly higher in the critically ill

patients (252.3 ± 171 , 185.12 ± 72.16 pg/mL respectively) than those in the general patients (64.15 ± 50.01 , 35.40 ± 11.90 pg/mL respectively), (Figure 3).



In the current studies, there was no significant association between interleukin-6,10 levels and gender, relapsed leukemia, sepsis, myocarditis, arthritis, hemophagocytic syndrome, and solid tumors (p>0.05). Interleukin-6 levels were significantly higher among children who had Initial leukemia and severe encephalitis (Median=525, 282.50 respectively) compared to those children who hadn't initial leukemia and

severe encephalitis (Median=144.76243, 234.50 respectively). Also, interleukin-10 levels were significantly lower among children who had post-CAR T cell/chemotherapy, and post-bone marrow transplantation (Median=144.76, and 163.44 respectively) compared to those who didn't post-CAR T cell/chemotherapy, and post-bone marrow transplantation (Median=203.50, 200, respectively), (Table 4)

		Group C (n=110)				
	No	Interleukin-6	UDmalus	Interleukin-10	^U P value	
		Median (range)	[•] P value	Median (range)		
Gender						
Male	71	243(10-743)	0.488	187(7.55-300)	0.389	
Female	39	252(28-703)		205.45(57.74-335.74)		
Initial leukemia						
No	103	243(10-743)	0.030*	193.25(7.55-335.74)	0.497	
Yes	7	525(76-703)		205.45(133-273.51)		
Relapsed leukemia						
No	100	239(10-743)	0.055	191.88(7.55-335.74)	0.632	
Yes	10	275(127-703)		207.22(47-295)		
Post-CAR T cell/chemotherapy						
No	88	245(10-743)	0.673	203.50(7.55-335.74)	0.024*	
Yes	22	244(24.50-555)		144.76(45-283)		
Post-bone marrow transplantation						
No	105	235(10-743)	0.080	200(7.55-335.74)	0.045*	
Yes	5	278(245.22-555)		163.44(45-189.36)		
Sepsis						
No	82	243.50(10-743)	0.714	203.26(7.55-335.74)	0.371	
Yes	28	255.50(19-629)		174.87(45-295)		
Severe pneumonia						
No	109	244(10-743)	0.582	197.91(7.55-335.74)	0.092	
Yes	1	278.00		145.00		
Severe encephalitis						
No	90	234.50(15-743)	0.045*	198.07(7.55-335.74)	0.895	
Yes	20	282.50(10-703)		186.72(45-295)		
Myocarditis						
No	107	245.22(15-743)	0.716	190.52(7.55-335.74)	0.204	
Yes	3	219(10-502)		223(205.45-274.71)		
Arthritis and hemophagocytic syndrome						
No	91	235(10-743)	0.819	189.36(7.55-335.74)	0.953	
Yes	19	256.23(24.50-703)		206.71(45-289.35)		

J. Clinical Pediatrics and Mother Health Copy rights @ Mohamed Zaeim Hafez					ed Zaeim Hafez. et	t al,	
	Solid tumors						1
	No	109	244(10-743)	0.236	193.25(7.55-335.74)	0.655	1
	Yes	1	527.25		217.98		I.

For Group C (critically ill patients), the association between interleukin 6, 10 levels and clinical examination was performed using the Mann-Whitney U test, *Significant.

Table 4: The association between interleukin 6, 10 levels and clinical examination among the studied critically ill pediatric patients.

In the present study, there was no significant relationship between Interlukine-6 and 10 levels in critically ill patients and organ functions (acute respiratory distress syndrome, acute liver injury, acute liver failure, acute kidney injury, acute myocardial injury, neurological abnormalities, coagulation abnormalities, and cardiac arrest), (p>0.05), (Table 5).

		Group C (n=110)				
	No	Interlukine-6	UDualua	Interlukine-10	^U P value	
		Median (range)	^r Value	Median (range)		
Acute gastrointestinal injury	1					
No	73	234(10-743)	0.302	190.52(47-335.74)	0.894	
Yes	37	260(15-703)		198.24(7.55-300)		
Acute respiratory distress syndrome	1					
No	108	244.6(10-743)	0.014^{*}	195.58(7.55-335.7)	0.647	
Yes	2	332.5(219-446)		152.58(82.16-223)		
Acute liver injury	Ī					
No	51	246(10-743)	0.636	187(7.55-289.65)	0.673	
Yes	59	243(23-703)		197.91(45-335.74)		
Acute liver failure						
No	106	245.61(10-743)	0.737	198.07(7.5-335.72)	0.177	
Yes	4	215(127-278)		122.14(45-223)		
Acute kidney injury	Ī					
No	87	246(10-743)	0.673	190.52(47-335.74)	0.971	
Yes	23	235(28-629)		204.53(7.55-269)		
Acute myocardial injury						
No	67	244(15-743)	0.776	204.53(7.55-313)	0.205	
Yes	43	245.22(10-593)		184.09(45-335.74)		
Neurological abnormalities	Ì					
No	75	235(10-703)	0.633	202(47-335.7)	0.220	
Yes	35	252(19-743)		181(7.55-295)		
Coagulation abnormalities	Ī					
No	48	243.5 (19-743)	0.756	178.5(7.55-335.74)	0.488	
Yes	62	250.5 (10-703		203.265(45-313)		
Cardiac arrest						
No	105	244 (10-743)	0.824	193.25(7.55-335.74)	0.813	
Yes	5	255 (178-333)		229.91(85.49-260)		

Group C (critically ill patients), acute gastrointestinal injury (AGI), acute respiratory distress syndrome (ARDS), acute liver injury (ALI), acute liver failure (ALF), acute kidney injury (AKI), and acute myocardial injury (AMI). The association between interleukin 6, 10 levels and organ dysfunctions were performed using the Mann-Whitney U test.

Table 5: The association between interleukin 6, 10 levels and organ dysfunctions among the studied critically ill pediatric patients.

ROC (receiver operating characteristic) analysis showed that interleukin10 levels could be used as good tool for diagnosis of critically ill patients with an area under a curve of 0.879 (p <.001) at 95%CI of (0.830-0.927), using a cut-off of >239 pg/mL with 92.5% sensitivity and

79.4% specificity as compared to interleukin 6 which at 95% CI (0.968-1), using a cut-off of >178.5 pg/mL with 97.4% sensitivity and 62% specificity (Table 6, Figure 4).

J. Clinical Pediatrics and Mother Health Copy rights @ Mohamed Zaeim Hafez. et al,								al,
		Area	Cutoff value >	P value	Sensitivity %	Specificity %	95%CI	
	Interlukine-6	0.879	239	< 0.001*	92.5%	79.4%	(0.830-0.927)	
	Interlukine-10	0.987	178.5	< 0.001*	97.4%	62%	(0.968-1)	

Confidence interval (CI), *significant

Table 6: The cut-off value interleukin 6,10 levels for a prediction of multiple organ dysfunction syndrome in critically ill pediatric patients.



Figure 4: ROC curve analysis of interleukin 6,10 levels for a prediction of Multiple organ dysfunction syndrome in critically ill pediatric patients.

Discussion

Multiple organ dysfunction syndrome (MODS) is a significant cause of morbidity and mortality in children undergoing cardiac surgery with cardiopulmonary bypass (CPB), (Benscoter et al. 2023). Early prediction of MODS is crucial for timely intervention and improved patient outcomes (Typpo et al. 2009, Gourd and Nikitas, 2020). While various biomarkers have shown promise (Pandey et al. 2015, Novak et al. 2023), this study focuses on the diagnostic potential of interleukin-6 (IL-6) and interleukin-10 (IL-10) levels in critically ill children with MODS.

In the current study according to initial leukemia, relapsed leukemia, postcart cell/chemotherapy, post-bone marrow transplantation, sepsis, severe pneumonia, severe encephalitis, myocarditis, arthritis, and hemophagocytic syndrome, and solid tumors. there was no significant difference between the studied groups. Also, coagulation abnormalities were more common (56.36%), followed by acute liver injury (53.64%), acute myocardial injury, acute gastrointestinal injury, neurological abnormalities, and acute kidney injury were observed in 39.09%, 33.63%, 31.81%, and 20.9% patients respectively. Zhu et al. (2024) showed that AKI and ALI happened in critically ill patients with the occurrence of 23.5% and 56%, in line with previous studies (Jettonet al. 2017, Kaddourah et al. 2017). Badke et al. (2022) found three common combinations of organ dysfunctions were associated with a significant portion of mortality and persistent MODS in our cohort. These included Cardiovascular and liver dysfunction: Linked to 75% of deaths, Respiratory and hematologic dysfunction: Linked to 75% of deaths, Respiratory and renal dysfunction: Linked to 75% of deaths and has been previously associated with fluid overload in critically ill children, potentially leading to respiratory failure and increased mortality.

Additionally, the following combinations were linked to persistent MODS, Neurologic and respiratory dysfunction: Linked to 36.3% of persistent MODS cases and Hematologic and immunologic dysfunction: Linked to 36.3% of persistent MODS cases, Endocrinologic and respiratory dysfunction: Linked to 36.3% of persistent MODS cases (Alobaidi et al. 2018, Barhight et al. 2022).

Another study by Weiss et al. (2015) found that 30% of children developed new or worsening MODS after sepsis diagnosis. Similarly, Villeneuve et al. (2016) reported an incidence of 19.6% and 20.1% for new and progressive MODS, respectively, using different diagnostic criteria. This suggests that the choice of criteria may not significantly impact the sample size of a randomized controlled trial focused on new or progressive MODS. Benscoter et al. (2023) have developed a new risk prediction model to assess the likelihood of MODS after pediatric cardiac surgery with CPB. If validated prospectively, this model could help identify high-risk patients and guide interventions to improve outcomes by preventing postoperative organ dysfunction.

The current study found significantly higher levels of interleukin-6 (IL-6) and interleukin-10 (IL-10) in critically ill patients compared to a control group. These findings align with previous research demonstrating elevated levels of various cytokines, including IL-6, IL-10, and IL-17A, in critically ill patients compared to healthy individuals. This suggests a strong association between cytokine levels and disease severity (Dhar et al. 2021, Jarczak et al. 2022, Zhang et al. 2022). In the context of sepsis, elevated IL-6 and IL-10 levels have been linked to progressive organ dysfunction, but they may not help identify specific bacterial types. However, these cytokines have been shown to correlate with the severity of organ dysfunction and can be useful predictors of sepsis prognosis (Zhang et al. 2022).

In the current study, Interleukin-6 levels were significantly higher among children who had Initial leukemia and severe encephalitis. Also, interleukin-10 levels were substantially lower among children who had post-CART cell/chemotherapy, and post bone marrow transplantation. Also, there was no significant relationship between Interlukine-6 and 10 levels in critically ill patients and organ functions (acute respiratory distress syndrome, acute liver injury, acute liver failure, acute kidney injury, acute myocardial injury, neurological abnormalities, coagulation abnormalities, and cardiac arrest). A previous study by Mu et al. (2021) suggested that IL-10 may contribute to the development of acute kidney injury (AKI), consistent with Zhu et al.'s (2024) findings of elevated IL-10 in patients with acute kidney injury. Additionally, research has focused on the "IL-6/IL-10 ratio" as a potential biomarker for disease severity. de Brito et al. (2016) demonstrated the clinical value of this ratio in early diagnosis of severe pneumonia. The ratio can reflect the balance between pro-inflammatory and anti-inflammatory responses during sepsis, which is crucial for understanding disease progression. IL-6, a multifunctional cytokine, is produced by macrophages and endothelial cells in response to inflammatory stimuli. It plays a role in various immune processes, including B-cell differentiation, T-cell activation, and the regulation of innate immune cell responses. Elevated IL-6 levels have been linked to increased disease severity and poor outcomes in clinical settings (Zhang et al. 2022).

Several studies have investigated the diagnostic potential of early IL-6 and IL-10 levels in severe infections. Vanska et al. (2012) found that a combination of IL-10 and procalcitonin (PCT) was effective in predicting severe febrile neutropenia. Persson et al. (2005) suggested that lower IL-6 levels in the early stages of neutropenic fever could indicate a milder infection.

In patients without progressive organ failure, elevated IL-6 and IL-10 levels were observed after both Gram-positive and Gram-negative bacterial infections, suggesting that these cytokines may not be highly specific for distinguishing between bacterial types in non-severe sepsis (Zhang et al. 2022). Similarly, in the non-NPMODS group, IL-6 elevation was primarily associated with worsening organ failure rather than specific bacterial types. While IL-6 elevation was linked to organ failure, IL-10 levels showed some differences between Gram-positive and Gramnegative bacterial infections, suggesting a potential role in identifying bacterial types (Zahar et al. 2011). However, other factors such as disease severity and organ dysfunction often have a greater impact on sepsis prognosis than the specific type of infecting organism.

ROC analysis showed that interleukin10 levels could be used as good tool for diagnosis of critically ill patients with an area under a curve of 0.879 (p <.001) at 95%CI of (0.830-0.927), using a cut-off of >239 pg/mL with 92.5% sensitivity and 79.4% specificity as compared to interleukin 6 which at 95% CI (0.968-1), using a cut-off of >178.5 pg/mL with 97.4% sensitivity and 62% specificity. ROC curves were plotted to assess the prediction value of the selected cytokines for organ injury. Another study reported significant differences in IL-10 between critically ill patients and general patients were indicated (Zhu et al. 2024). The AUC for IL-10 (more than 279.45 pg/mL) in predicting cardiac arrest was the largest (AUC =0.913, sensitivity =0.875, specificity =0.927), followed by the IL-6 (more than 1,425.6 pg/mL) for cardiac arrest (AUC =0.754, sensitivity =0.625, specificity =0.844). L-10, according to existing research, is a cytokine that suppresses the immune response (Fiorentino et al. 1991, Del Prete et al. 1993, Couper et al. 2008, Deng et al. 2012, Hombach et al. 2012).

Copy rights @ Mohamed Zaeim Hafez. et al,

Interestingly, while IL-10 is generally considered an anti-inflammatory cytokine, in this context, elevated levels were associated with severe tissue damage and predicted cardiac arrest. This suggests a complex role for IL-10 in the inflammatory response, potentially involving both protective and harmful effects (Steensberg et al. 2003, Tanaka et al. 2016). IL-6, on the other hand, is typically considered a pro-inflammatory cytokine. While it can induce anti-inflammatory responses through negative feedback mechanisms, excessive IL-6 levels can contribute to severe inflammation and adverse outcomes. The combination of IL-6 and IL-10 has been explored as a potential biomarker for disease severity. While some studies have shown promise, the results may vary depending on the specific disease and patient population (Dhar et al. 2021). Further research is needed to fully understand the complex interplay between these cytokines and their role in critical illness.

Conclusions

In conclusion, coagulation abnormalities and acute liver injury were the most common organ dysfunctions observed in the critically ill patient group. Significantly higher levels of IL-6 and IL-10 were detected in critically ill patients compared to the general patient group. Notably, IL-10 emerged as a valuable biomarker for the diagnosis of critical illness, with a cutoff value of >239 pg/mL demonstrating high sensitivity (92.5%) and specificity (79.4%). These findings highlight the potential of IL-10 as a diagnostic tool for identifying critically ill patients and guiding clinical decision-making.

Consent for publication: all authors have read and revised well for the manuscript and agree to publish.

Availability of data and material: All data supporting the study are presented in the manuscript or available upon request.

Competing interests: There is no conflict of interest.

Funding: The author received no financial support for this article's research, authorship, and publication.

Acknowledgments: Not applicable

Authors' information (optional): Not applicable

References

- Akkaya O, Mutlu N, Koylu R, Akilli NB, Koylu O, Eryilmaz MA. Assessment of the correlations between interleukin-6 and 10 levels and mortality in patients with sepsis. Annals of Medical Research. 2022 Nov 3;30(1):0-.
- Alobaidi R, Morgan C, Basu RK, Stenson E, Featherstone R, Majumdar SR, et al. Association between fluid balance and outcomes in critically ill children: a systematic review and meta-analysis. JAMA Pediatr. (2018) 172:257–68. doi: 10.1001/jamapediatrics.2017.4540
- Andruszkow H, Fischer J, Sasse M, Brunnemer U, Andruszkow JH, Gansslen A, Hildebrand F, Frink M. Interleukin-6 as inflammatory marker referring to multiple organ dysfunction syndrome in severely injured children. Scandinavian Journal of Trauma, Resuscitation and Emergency Medicine. 2014 Mar 3;22(1);1-8.
- Arias AV, Lintner-Rivera M, Shafi NI, Abbas Q, Abdelhafeez AH, Ali M, Ammar H, Anwar AI, Appiah JA, Attebery JE, Villalobos WE. A research definition and framework for acute paediatric critical illness across resource-variable settings: a

modified Delphi consensus. The Lancet Global Health. 2024 Feb 1;12(2): e331-40.

- Badke CM, Mayampurath A, Sanchez-Pinto LN. Multiple organ dysfunction interactions in critically ill children. Frontiers in Pediatrics. 2022 Apr 25; 10:874282.
- Badke CM, Mayampurath A, Sanchez-Pinto LN. Multiple organ dysfunction interactions in critically ill children. Frontiers in Pediatrics. 2022 Apr 25; 10:874282.
- Balkrishna A, Sinha S, Kumar A, Arya V, Gautam AK, Valis M, Kuca K, Kumar D, Amarowicz R. Sepsis-mediated renal dysfunction: pathophysiology, biomarkers and role of phytoconstituents in its management. Biomedicine & Pharmacotherapy. 2023 Sep 1; 165:115183.
- Barhight MF, Nelson D, Chong G, Basu RK, Sanchez-Pinto LN. Non-resuscitation fluid in excess of hydration requirements is associated with higher mortality in critically Ill children. Pediatr Res. (2022) 91:235–240.
- Behzadi P, Sameer AS, Nissar S, Banday MZ, Gajdács M, García-Perdomo HA, Akhtar K, Pinheiro M, Magnusson P, Sarshar M, Ambrosi C. The Interleukin-1 (IL-1) superfamily cytokines and their single nucleotide polymorphisms (SNPs). Journal of immunology research. 2022;2022(1):2054431.
- Benscoter AL, Alten JA, Atreya MR, Cooper DS, Byrnes JW, Nelson DP, Ollberding NJ, Wong HR. Biomarker-based risk model to predict persistent multiple organ dysfunctions after congenital heart surgery: a prospective observational cohort study. Critical Care. 2023 May 20;27(1):193.
- Bose SN, Greenstein JL, Fackler JC, Sarma SV, Winslow RL, Bembea MM. Early prediction of multiple organ dysfunction in the pediatric intensive care unit. Frontiers in pediatrics. 2021 Aug 16; 9:711104.
- 12. Broderick L, Hoffman HM. IL-1 and autoinflammatory disease: biology, pathogenesis and therapeutic targeting. Nature Reviews Rheumatology. 2022 Aug;18(8):448-463.
- 13. Chinese Abdominal Intensive Care Association. Asia society for emergency and critical care medicine. Expert consensus on enteral nutrition for gastrointestinal dysfunction in critically ill patients. Chin J Dig Surg 2021; 20:1123-1136.
- 14. Couper KN, Blount DG, Riley EM. IL-10: the master regulator of immunity to infection. J Immunol 2008;180:5771-5777.
- de Brito RC, Lucena-Silva N, Torres LC, Luna CF, Correia JB, da Silva GA. The balance between the serum levels of il-6 and il-10 cytokines discriminates mild and severe acute pneumonia. BMC Pulm Med (2016) 16(1):170. doi: 10.1186/ s12890-016-0324-z
- Del Prete G, De Carli M, Almerigogna F, et al. Human IL-10 is produced by both type 1 helper (Th1) and type 2 helper (Th2) T cell clones and inhibits their antigen-specific proliferation and cytokine production. J Immunol 1993; 150:353-360.
- 17. Deng B, Wehling-Henricks M, Villalta SA, et al. IL-10 triggers changes in macrophage phenotype that promote muscle growth and regeneration. J Immunol 2012; 189:3669-3680
- 18. Dhar SK, K V, Damodar S, et al. IL-6 and IL-10 as predictors of disease severity in COVID-19 patients: results from metaanalysis and regression. Heliyon 2021;7: e06155.
- Fan B, Klatt J, Moor MM, Daniels LA, Sanchez-Pinto LN, Agyeman PK, Schlapbach LJ, Borgwardt KM. Prediction of recovery from multiple organ dysfunction syndrome in

Copy rights @ Mohamed Zaeim Hafez. et al,

pediatric sepsis patients. Bioinformatics. 2022 Jul 1;38(Supplement_1):i101-108.

- 20. Fernández-Sarmiento J, Schlapbach LJ, Acevedo L, Santana CR, Acosta Y, Diana A, Monsalve MC, Carcillo JA. Endothelial damage in sepsis: the importance of systems biology. Frontiers in Pediatrics. 2022 Mar 9; 10:828968.
- 21. Fiorentino DF, Zlotnik A, Vieira P, et al. IL-10 acts on the antigen-presenting cell to inhibit cytokine production by Th1 cells. J Immunol 1991; 146:3444-3451.
- 22. Gaugler M, Swinger N, Rahrig AL, Skiles J, Rowan CM. Multiple organ dysfunction and critically ill children with acute myeloid leukemia: Single-center retrospective cohort study. Pediatric Critical Care Medicine. 2023 Apr 1;24(4): e170-178.
- 23. Gidon A, Louet C, Røst LM, Bruheim P, Flo TH. The tumor necrosis factor alpha and interleukin 6 auto-paracrine signaling loop controls Mycobacterium avium infection via induction of IRF1/IRG1 in human primary macrophages. MBio. 2021 Oct 26;12(5):110-128.
- 24. Gourd NM, Nikitas N. Multiple Organ Dysfunction Syndrome. J Intensive Care Med 2020; 35:1564-1575.
- 25. Hombach AA, Heiders J, Foppe M, et al. OX40 costimulation by a chimeric antigen receptor abrogates CD28 and IL-2 induced IL-10 secretion by redirected CD4(+) T cells. Oncoimmunology 2012; 1:458-466.
- 26. Ishikawa S, Teshima Y, Otsubo H, Shimazui T, Nakada TA, Takasu O, Matsuda K, Sasaki J, Nabeta M, Moriguchi T, Shibusawa T. Risk prediction of biomarkers for early multiple organ dysfunction in critically ill patients. BMC Emergency Medicine. 2021 Nov 8;21(1):132.
- 27. Jarczak D, Nierhaus A. Cytokine Storm-Definition, Causes, and Implications. Int J Mol Sci 2022; 23:11740.
- Jetton JG, Boohaker LJ, Sethi SK, et al. Incidence and outcomes of neonatal acute kidney injury (AWAKEN): a multicentre, multinational, observational cohort study. Lancet Child Adolesc Health 2017; 1:184-194.
- 29. Kaddourah A, Basu RK, Bagshaw SM, et al. Epidemiology of Acute Kidney Injury in Critically Ill Children and Young Adults. N Engl J Med 2017; 376:11-20.
- Killien EY, Zahlan JM, Lad H, Watson RS, Vavilala MS, Huijsmans RL, Rivara FP. Epidemiology and outcomes of multiple organ dysfunction syndrome following pediatric trauma. Journal of Trauma and Acute Care Surgery. 2022 Dec 1;93(6):829-837.
- 31. Kortz TB, Nielsen KR, Mediratta RP, Reeves H, O'Brien NF, Lee JH, Attebery JE, Bhutta EG, Biewen C, Coronado Munoz A, DeAlmeida ML. The burden of critical illness in hospitalized children in low-and middle-income countries: protocol for a systematic review and meta-analysis. Frontiers in pediatrics. 2022 Mar 16; 10:756643.
- 32. Liang J, Su Y, Wang N, Wang X, Hao L, Ren C. A metaanalysis of the association between inflammatory cytokine polymorphism and neonatal sepsis. Plos one. 2024 Jun 7;19(6):e0301859.
- Liu S, Wang C, Guo J, et al. Serum Cytokines Predict the Severity of Coronary Artery Disease Without Acute Myocardial Infarction. Front Cardiovasc Med 2022; 9:896810.
- 34. McDonagh TA, Metra M, Adamo M, et al. 2021 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure. Eur Heart J 2021; 42:3599-3726.

Copy rights @ Mohamed Zaeim Hafez. et al,

J. Clinical Pediatrics and Mother Health

- Migliorini P, Italiani P, Pratesi F, Puxeddu I, Boraschi D. The IL-1 family cytokines and receptors in autoimmune diseases. Autoimmunity reviews. 2020 Sep 1;19(9):102617.
- 36. Mu H, Zheng Q, Hao L. IL-10 -1082 A/G polymorphism is related with the risk and clinical characteristics of acute kidney injury: a case-control study. BMC Nephrol 2021; 22:212.
- 37. Novak T, Crawford JC, Hahn G, et al. Transcriptomic profiles of multiple organ dysfunction syndrome phenotypes in pediatric critical influenza. Front Immunol 2023; 14:1220028
- 38. Pandey N, Jain A, Garg RK, et al. Serum levels of IL-8, IFN γ , IL-10, and TGF β and their gene expression levels in severe and non-severe cases of dengue virus infection. Arch Virol 2015; 160:1463-1475.
- Persson L, Soderquist B, Engervall P, Vikerfors T, Hansson LO, Tidefelt U. Assessment of systemic inflammation markers to differentiate a stable from a deteriorating clinical course in patients with febrile neutropenia. Eur J Haematol (2005) 74(4):297–303.
- 40. Pickkers P, Darmon M, Hoste E, et al. Acute kidney injury in the critically ill: an updated review on pathophysiology and management. Intensive Care Med 2021; 47:835-850
- Rose-John S, Jenkins BJ, Garbers C, Moll JM, Scheller J. Targeting IL-6 trans-signalling: past, present and future prospects. Nature Reviews Immunology. 2023 Oct;23(10):666-681.
- 42. Rose-John S. Interleukin-6 signaling in health and disease. F1000Research. 2020 Aug 20;9(1013):1013.
- 43. Ruth AR, Boss RD, Donohue PK, Shapiro MC, Raisanen JC, Henderson CM. Living in the hospital: the vulnerability of children with chronic critical illness. The Journal of clinical ethics. 2020 Dec 1;31(4):340-352.
- 44. Sanchez-Pinto LN, Stroup EK, Pendergrast T, Pinto N, Luo Y. Derivation and validation of novel phenotypes of multiple organ dysfunction syndrome in critically ill children. JAMA network open. 2020 Aug 3;3(8): e209-271.
- 45. Shimazui T, Nakada TA, Yazaki M, Mayumi T, Takasu O, Matsuda K, Sasaki J, Otsubo H, Teshima Y, Nabeta M, Moriguchi T. Blood interleukin-6 levels predict multiple organ dysfunction in critically ill patients. Shock. 2021 Jun 1;55(6):790-795.
- Southard-Goebel C, Pike F, Rowan CM, Cater DT. Risk Factors Associated with Development of Multiple-Organ Dysfunction Syndrome After Pediatric Drowning. Pediatric emergency care. 2023 Dec 1;39(12):902-906.
- Steensberg A, Fischer CP, Keller C, et al. IL-6 enhances plasma IL-1ra, IL-10, and cortisol in humans. Am J Physiol Endocrinol Metab 2003;285: E433-437.

- Tanaka T, Narazaki M, Kishimoto T. Immunotherapeutic implications of IL-6 blockade for cytokine storm. Immunotherapy 2016; 8:959-970.
- 49. Teufel LU, Arts RJ, Netea MG, Dinarello CA, Joosten LA. IL-1 family cytokines as drivers and inhibitors of trained immunity. Cytokine. 2022 Feb 1;150:155773.
- Typpo KV, Petersen NJ, Hallman DM, et al. Day 1 multiple organ dysfunction syndrome is associated with poor functional outcome and mortality in the pediatric intensive care unit. Pediatr Crit Care Med 2009; 10:562-570.
- Vanska M, Koivula I, Jantunen E, Hamalainen S, Purhonen AK, Pulkki K, et al. II-10 combined with procalcitonin improves early prediction of complications of febrile neutropenia in hematological patients. Cytokine (2012) 60(3):787–792.
- 52. Villeneuve A, Joyal JS, Proulx F, Ducruet T, Poitras N, Lacroix J. Multiple organ dysfunction syndrome in critically ill children: clinical value of two lists of diagnostic criteria. Annals of Intensive Care. 2016 Dec; 6:1-7.
- 53. Weiss SL, Fitzgerald JC, Pappachan J, Wheeler D, Jaramillo-Bustamante JC, Salloo A, Singhi SC, Erickson S, Roy JA, Bush JL, Nadkarni VM. Global epidemiology of pediatric severe sepsis: the sepsis prevalence, outcomes, and therapies study. American journal of respiratory and critical care medicine. 2015 May 15;191(10):1147-1157.
- 54. Zahar JR, Timsit JF, Garrouste-Orgeas M, Francais A, Vesin A, Descorps-Declere A, et al. Outcomes in severe sepsis and patients with septic shock: Pathogen species and infection sites are not associated with mortality. Crit Care Med (2011) 3 9(8): 1886–1895.
- Zhang J, Wang J, Gong Y, et al. Interleukin-6 and granulocyte colony-stimulating factor as predictors of the prognosis of influenza-associated pneumonia. BMC Infect Dis 2022; 22:343.
- 56. Zhang Y, Li B, Ning B. Evaluating IL-6 and IL-10 as rapid diagnostic tools for Gram-negative bacteria and as disease severity predictors in pediatric sepsis patients in the intensive care unit. Front Immunol 2022; 13:1043968.
- 57. Zhu R, Cao L, Wu T, Zhang Z, Han M, Liu H, Huang S, Bai Z, Wu S. The evaluation of cytokines in predicting the organ injury of critically pediatric patients: a retrospective study. Translational Pediatrics. 2024 Jul 25;13(7):1169.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article Click Here:

Submit Manuscript

DOI: 10.31579/2835-2971/019

Ready to submit your research? Choose Auctores and benefit from:

- ➢ fast, convenient online submission
- > rigorous peer review by experienced research in your field
- rapid publication on acceptance
- > authors retain copyrights
- > unique DOI for all articles
- immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <u>https://auctoresonline.org/journals/clinical-pediatrics-and-mother-health</u>