

Targeting on the podocyte immune response in diabetic nephropathy: from pathogenesis to therapy

He Qin ¹, Peng Rongdong ², Li Zukai ³, Feng Junxia ⁴, Zhou Xiaoying ⁴, Su Yanyan ^{1,3*}

¹ The Third School of Clinical Medicine, Southern Medical University, Guangzhou, 510515, China,

² Medical Department, Qingbu Community Health Service Center, Huadu District, Guangzhou, Guangzhou, 510800, China,

³ Department of Nephrology, Huadu District People's Hospital of Guangzhou, Guangzhou, 510800, China,

⁴ Department of Central laboratory, Huadu District People's Hospital Guangzhou, Guangzhou, 510800, China.

***Corresponding Author:** Yanyan Su, Department of Nephrology, Huadu District People's Hospital of Guangzhou, 510800, No. 48 Xinhua Road, Guangzhou, China

Received date: August 27, 2024; **Accepted date:** September 18, 2024; **Published date:** October 01, 2024

Citation: He Qin, Peng Rongdong, Li Zukai, Feng Junxia, Zhou Xiaoying, Su Yanyan, (2024), Targeting on the podocyte immune response in diabetic nephropathy: from pathogenesis to therapy, *J Clinical Research and Reports*, 16(5); DOI:10.31579/2690-1919/398

Copyright: © 2024, Su Yanyan. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

The dysfunction of the glomerular filtration barrier (GFB) is a main feature of diabetic nephropathy (DN). Podocyte, terminally differentiated epithelial cell, is located on the outer of the glomerular basement membrane and play a pivotal role in maintaining the integrity of GFB. Damage in podocyte leads to proteinuria in the early stages of DN and eventually develops into chronic kidney disease. Podocyte expresses certain immune characteristics and this may predispose them to immune injury in diseases. Immune-mediated podocyte injury is a vital pathogenic mechanism in DN. Numerous evidence indicate that podocytes serve as a target of immune response in both innate and adaptive immunity, which mediate the podocyte injury in DN. In this mini-review, we focus on recent developments involving in the immune mechanisms of podocyte injury in DN and provide recent advances in the therapeutic strategy of podocyte protection. This review offers valuable opinions for understanding immune-related mechanisms in podocyte injury and identification of potential therapeutic targets, which have significant implications for future research and clinical treatment.

Keywords: podocyte; immune response; diabetic nephropathy

Introduction

Diabetic nephropathy (DN) is one of microvascular complications in diabetes mellitus, accounting for the major cause of end-stage renal disease (ESRD) worldwide [1]. Increasing evidence indicate that glucose-lowering agents have limited efficacy in preventing the progression of DN in its later stages. The clinical development of DN is classified according to the level of urine albumin excretion and the decline in glomerular filtration rate (GFR). In the development of DN, urine albumin to creatinine ratio (UACR) between 30 to 299 mg/g and UACR \geq 300 mg/g are separately described as incipient nephropathy and overt nephropathy, and GFR $<$ 30 ml/min/1.73m² is considered as kidney failure [2-3]. Plenty of animal blood glucose and several agents can alleviate the development of DN, a proportion of diabetic patients in ESRD have to require kidney dialysis to stay alive [4].

It has been well known that complex mechanisms are involved in the pathogenesis of DN onset and development. Podocyte injury is one of the main characteristic features in DN, occurring as a consequence of their

loss and dysfunction, which is a driver of albuminuria [5-6]. Podocytes account for major component of the GFB and are highly specialized epithelial cells with limited capacity for renewal, which means podocyte loss can hardly be fully compensated by regeneration nearby healthy podocytes [7]. Thus, understanding the mechanism of podocyte injury in DN can provide insights to therapeutic strategies in the future. Up to date, emerging studies show that multiple mechanisms are involved in podocyte injury, including abnormal apoptosis, pyroptosis, necroptosis, autophagy, inflammation and immune-related cell death, as well as several molecular signal pathways [8-10]. Moreover, podocyte exhibits some features of immune cell, such as expressing certain immune characteristics (MHC class, B7-1, FcRn) and acting as antigen-presenting cell [11-12], and the podocyte's properties in immunity may predispose it to immune injury in kidney diseases. In this review, we aim to determine the immune-related mechanisms of podocyte injury in DN and further discuss the agents recommended by guidelines on how best to protect the life of podocytes in our kidneys.

1 Inflammation in DN

Numerous studies illustrate that both systemic and local renal inflammation are associated with DN. In different stages of DN, elevated levels of inflammatory cytokines and immune mediators can be detected in serum and peripheral blood cells, such as interleukin-6 (IL-6), tumor necrosis factor- α (TNF- α), monocyte chemoattractant protein-1 (MCP-1), intracellular adhesion molecule-1 (ICAM-1) and myeloid differentiation factor 88 (MyD88) [13,14]. In patients with type 2 diabetes, chronic inflammation in adipose tissue is considered a vital contributor to insulin resistance [15]. The serum C-reactive protein (CRP), a marker of systemic inflammation, increases in early diabetic kidney injury and is associated with the development of microalbuminuria in diabetic patient [16]. Urine level of MCP-1 is upregulated in diabetic patient with albuminuria and relates to the decline of renal function [17]. Furthermore, the urine level of MCP-1 is considered a predictor of the progression of DN [18]. These evidence demonstrate the vital role of inflammation in the development of patients with DN.

Histological analysis of renal biopsy is considered a “golden standard” strategy to diagnose the features of DN. However, in real clinical activity, renal biopsies are rarely performed in diabetic patients. Those who have substantial increase in albuminuria or significant decline in renal function, are suitable patients to perform renal biopsies to determine whether there

are additional diseases [19,20]. Due to above limitations, animal model studies have unique advantages in understanding pathological characteristics of kidney injury [21]. Animal models can partially exhibit pathological characteristics of kidney injury that are similar to patients, and these strategies help researchers to well understand the process of pathology in DN [22]. Plentiful of animal studies have illustrated inflammatory cell infiltration in the kidney is associated with the progression of DN. Proinflammatory cytokines and leucocyte adhesion molecules are elevated in diabetic kidneys, while inhibition of inflammatory response can alleviate the development of renal inflammation and improve renal function [23-26]. This evidence supports inflammations in the kidney, an early feature of DN, which is required for disease progression.

2 immune responses associated with podocyte injury

Podocyte injury is one of the major characteristics in DN and plays an essential role in the progression of albuminuria [1]. Emerging evidence illustrate podocyte displays the properties of immune cell and this may predispose podocyte to damage by immune response [12]. Studies also demonstrate that innate and adaptive immune responses extensively exert effects in the onset and progression of podocyte injury, which supports the inseparably intimate relation between podocyte injury and immune response in DN (as shown in Figure1).

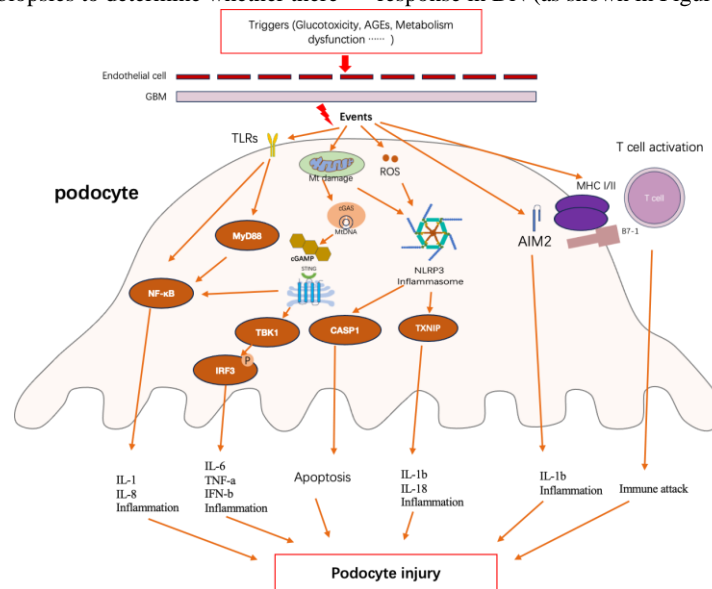


Figure 1

Innate and adaptive immune response in podocyte lead to podocyte injury in diabetic nephropathy. Triggers that induce the immune response in podocyte include various factors, including glucotoxicity, advanced glycation end-products (AGEs) and metabolism dysfunction such as lipidic toxicity [27-28]. Events are the form of actions when podocyte response to extracellular stress, include a range of molecular mechanism [29]. Podocyte injury contains podocyte effacement, apoptosis, cell structure alteration and cell dysfunction and so on [30]. GBM, glomerular basement membrane; TLRs, toll-like receptors; MyD88, myeloid differentiation factor 88; NF- κ B, nuclear factor kappa B; Mt DNA, mitochondrial DNA; cGAS, cyclic GMP-AMP synthase; cGAMP, 2'-3'-cyclic GMP-AMP; STING, stimulator of interferon gene; TBK1, TNK-binding kinase1; IRF3, interferon regulatory factor 3; ROS, reactive oxygen species; NLRP3, NLR family pyrin domain containing 3; CASP1,

caspase 1; TXNIP, thioredoxin (TRX)-interacting protein; AIM2, absent in melanoma 2; MHC, major histocompatibility complex; IL-1 β , interleukin 1 β ; IL-1, interleukin 1; IL6, interleukin 6; IL-18, interleukin 18; IL8, interleukin 8; TNF- α , tumor necrosis factor alpha; IFN- β , interferon β ;

2.1 Innate immune responses associated with podocyte injury

The innate immune system serves as the front defense of the host to eliminate invading pathogens [31]. Difference from the adaptive immune system, the innate immune system is capable of quickly detecting and removing the invaders without clonal expansion of antigen-specific lymphocytes. The innate immune system depends on pattern recognition receptors (PRRs), which can recognize pathogen-associated molecular patterns (PAMPs) and danger-associated molecular patterns (DAMPs). There are various types of PRRs in mammalian animals, including toll-

like receptors (TLRs), retinoic acid-inducible Gene-I (RIG-I)-like receptors (RLRs), Nucleotide-binding and oligomerization domain (NOD)-like receptors (NLRs), absent in melanoma 2 (AIM2)-like receptors (ALRs), and C-type lectin receptors (CLRs), as well as cyclic GMP-AMP synthase (cGas) and other intracellular DNA sensors [32-34]. According to subcellular localization, they can be divided into membrane-bound PRRs (TLRs and CLRs) or cytoplasmic PRRs (NLRs, RLRs, ALRs, and cGas) [35]. PRRs are mainly expressed in immune cells, such as macrophages and dendritic cells, as well as nonimmune cells, such as podocytes. Podocyte has been found to express various PRRs which play a vital role in response to innate immunity in DN [11].

2.1.1 Involvement with TLRs

The best-characterized PRRs are TLRs, which have been extensively explored. To date, studies show that ten kinds of TLRs (TLR1-10) in human species and twelve kinds of TLRs (TLR1-9, TLR11-13) in mouse species have been identified [36]. TLR located on the cell surface have the N-terminal and Toll/interleukin-1 receptor (TIR) domain, the former consists of the Leucine-rich repeat (LRR) with substrate binding ability towards the extracellular, and the latter is responsible for transferring signals into the cell [37]. Various types of cells can express TLRs, including immune cells, endothelia and epithelial cells. Podocyte has been found to express TLRs and the activation of TLRs in podocyte is the response to cell stress and injury, which can induce secretion of cytokines and chemokines to prompt the innate immune response [38-39]. For instance, in human immortalized podocytes, induction of podocyte injury by puromycin aminonucleoside is associated with the upregulation of TLR9 expression and activation of nuclear factor kappa-B (NF- κ B) [40]. TLR2 and TLR4 signaling are involved in the pathogenesis of several kidney diseases, such as acute kidney disease [41], kidney transplantation [42] and diabetic nephropathy [43]. These evidence demonstrate a double role of TLRs in podocytes, not only as major players in response to foreign pathogens but also as mediators of podocyte injury. Additionally, among TLRs, preclinical and clinical studies support the causative role of TLR2 and TLR4 in DN [39], thereby in this section we mainly focus on the role of TLR2 and TLR4 in podocyte injury.

It is reported that high glucose induced the overexpression of TLR2 and TLR4 in kidney tissues, the process of which were associated with the increased levels of several chemokines and immune mediators, such as NF- κ B, interleukin-8 (IL-8), MCP-1, ICAM-1, and vascular cell adhesion molecule-1 (VCAM-1) [44]. High glucose directly promoted TLR4 activation in podocytes in vitro, resulting in MyD88/NF- κ B activation and inflammatory response. Consistent with this, elevated expression of TLR4 and its downstream cytokines and chemokines were detected in diabetic nephropathy in WT mice with STZ-induced diabetes [45]. High glucose also directly promoted TLR2 activation in podocytes and tubular epithelial cells in vitro, resulting in NF- κ B activation and inflammation [46]. Interestingly, treatment with TLR2 agonists further exacerbated podocyte injury, inflammatory infiltration and serum creatinine levels in STZ-induced diabetic mice [47]. On the other hand, blocking the Toll-like receptor might have potential protection for diabetic podocyte damage. In cultured human podocytes with sera from T1D patients, high LPS activity induced cell apoptosis via the downregulation of the Akt cell survival pathway, however, these effects were prevented by inhibiting the TLR signaling pathway with immunomodulatory agent GIT27 [48]. Human umbilical cord mesenchymal stem cells decreased the inflammation of podocytes under high glucose via inhibition of TLR2 and

TLR4 signaling pathways [49], suggesting their vital role in mediating inflammation in diabetic nephropathy. Compared to diabetic WT mice, TLR4^{-/-} mice were protected against the development of diabetic nephropathy, exhibiting less podocyte injury, albuminuria and inflammation [45]. Similar to TLR4^{-/-} mice, deficient TLR2 mice also displayed protection against podocyte injury in diabetic nephropathy [46]. Collectively, both genetic deletion and pharmacological inhibition of TLR2 and TLR4 in diabetic mice attenuate podocyte injury and kidney dysfunction, suggesting that they might be potential targets for podocyte protection in DN.

2.1.2 Involvement with NLRs

It has been well known that nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs) are intracellular receptors, mainly located in the cytoplasm, which have C-terminal leucine-rich nucleotide-binding (NACHT) and N-terminal effector domain. NLRs can be further subdivided into four different subfamilies, including NLRAs, NLRBs, NLRs, and NLRPs, according to their N-terminal domains. [50] The NLRs exert multiple effects and play pivotal roles in innate immunity, such as presenting antigens, pathogen sensing and mediation in inflammation [51]. Although the links between NLRs and renal disease still need to be further elucidated, increasing pieces of evidence demonstrate that several NLRs, including NLR family pyrin domain containing 3 (NLRP3), NOD1, and NOD2 play crucial roles in the pathophysiological process of DN [52].

NLRP3. The proteins of the NLR family are mainly localized in the cell and are divided into NLRC proteins with CARD domains and NLRP proteins with pyrin domains, according to their N-terminal domain. NLRP3 is an important member of the NLRs family, which has been studied extensively [53]. The NLRP3 inflammasome and its downstream signaling molecules, such as caspase-1, interleukin -1 β (IL-1 β), and interleukin-18 (IL-18), play a pivotal role in cell death and are associated with podocyte injury in DN [54]. The process of NLRP3 inflammasome activation is regulated not only by infection and inflammation but also by nutrition and metabolism [55]. It is generally thought that the activation of NLRP3 inflammasome involves two steps, including priming and protein complex assembly, the former step is triggered by PRR signaling or cytokines which leads to the transcriptional activation of NLRP3 inflammasome components, and the later step is induced by various PAMPs and DAMPs [56-57]. A few studies support that the overactivation of NLRP3 inflammasome often leads to various diseases and cell injury [58-61]. It has also been shown that inhibition of NLRP3 inflammasome leads to the suppression of multiple proinflammatory signaling pathways and improves cell survival and tissue function, supporting that NLRP3 inflammasome might be the potential target for the treatment of podocyte damage in diabetes. [62-63]

Experimentally, study from Gao et al. showed that podocytes were capable of expressing NLRP3 inflammasome, and exposure to high glucose could activate NLRP3 inflammasome and induce IL-1 production by interacting with thioredoxin (TRX)-interacting protein (TXNIP), which eventually resulted in podocyte injury [64]. Hyperglycemia induced the expression of TXNIP and subsequently caused the activation of gp91, which was a subunit of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase. In vivo studies, genetic deletion of TXNIP abrogated NLRP3 inflammasome activation through inhibition of NADPH oxidase and alleviated podocyte injury in STZ-induced diabetic mice [65]. In the presence of high glucose or AGEs, mitochondrial-

derived reactive oxygen species (ROS) stimulated the NLRP3 inflammasome in podocytes from diabetic patients, while mice with abolishment of NLRP3 expression in non-myeloid derived cells have protection against diabetic nephropathy [66]. Apoptosis-associated speck-like protein (ASC) is a component of NLRP3 inflammasome. Inhibition of ASC by shRNA transfection in high-fat diet mice attenuated proteinuria, foot process effacement and loss of slit diaphragm molecules in podocytes [55]. In vitro cultured podocytes, pharmacological or siRNA inhibition of NLRP3 markedly decreased high glucose-induced apoptosis, cytoskeleton change and lipid accumulation [67], as well as restoration in podocyte autophagy and nephrin expression [68].

However, Sophie et al. argued that NLRP3 was poorly expressed or virtually absent in podocyte [69]. In this study, RNA-Seq data released from the Human Protein Atlas were explored by single-cell transcriptome analysis and the results showed the absence of relevant amounts of transcripts for NLRP3, IL-1b and other related molecules. They further identified the lack of mRNA expression of NLRP3, CASP1 (caspase 1) and IL-1b in cultured human podocytes, even under the presence of NLRP3 inflammasome inducers lipopolysaccharides (LPS) and adenosine triphosphate (ATP). They also raised the limitations of their study, in which the cultured human podocyte transcriptomes were not performed in diabetic conditions and the mouse model of diabetes did not develop overt diabetic nephropathy. In brief, whether podocyte express NLRP3 inflammasome seems contradictory in the published studies, suggesting that further studies are needed to identify the role of NLRP3 in podocyte injury of diabetic kidney disease.

NOD1 and NOD2. The expression of NOD2 can be identified in various types of cells, including tubular epithelial cells, glomerular endothelial cells and podocytes, and plays a crucial role in the pathogenesis of DN [70]. Clinical and animal studies demonstrate that the upregulation of NOD2 expression can be detected in the kidney biopsy samples from patients with T2DM and STZ-induced diabetic mice. Experimentally, the presence of high glucose, advanced glycation end-products (AGEs), TNF- α and transforming growth factor- β (TGF- β) significantly increased and activated the NOD2 expression in vitro cultured podocytes, and the activation of NOD2 expression induced the secretion of proinflammatory and profibrotic mediators, resulting in podocyte injury [71]. RNA-binding protein human antigen (HuR) is a key posttranscriptional regulator of NOD2 expression, acting to enhance NOD2 mRNA stability. In the kidneys of patients with DN, the HuR expression was upregulated and closely related to the progression of proteinuria [72]. It had also been found that NOD2 affected glucose handling and nephrin expression in podocytes, as well as TGF- β signaling in tubular cells and immune cells [71-73]. On the other hand, genetic NOD2 knockout in diabetic mice or inhibition of NOD2 expression in cultured podocytes displayed protection against the hyperglycemia-induced podocyte damage, leading to the alleviation of loss in nephrin and disruption of actin filament structure [71]. Moreover, the activation of NOD1 expression is related to metabolism inflammation and insulin resistance induced by obesity. Study found that the NOD1-RICK-NF- κ B inflammatory signaling pathway contributed to the pathogenesis and progression of DN. NOD1 deficiency had protection against high-fat diet-induced insulin resistance [74]. However, whether NOD1 is involved in podocyte injury in the development of DN needs to be further explored. Collectively, based on the above evidence, therapies targeting on NLRs may provide promising benefits for podocyte protection.

2.1.3 Involvement with RLRs

The retinoic acid-inducible gene I (RIG-I)-like receptors (RLRs) are RNA sensors, which are localized in the cytosol and act on important inducers of type I interferons and other antiviral immune mediators [75]. This protein family includes three members: RIG-I, melanoma differentiation-associated gene5 (MDA5) and laboratory of genetics and physiology 2 (LGP2). Generally speaking, RLRs consist of a central helicase and a carboxy-terminal domain (CTD), while RIG-I and MDA5 additionally have two N-terminal caspase recruitment domains (CARD). These domains work together in response to immunostimulatory dsRNA and exert crucial effects on viral infection. Moreover, RLRs are associated with a range of cellular processes, such as proliferation, differentiation and apoptosis [76]. It has been shown that besides immune cells, podocytes express RIG-I and MDA5, and the activation of RLR is related to podocyte damage. For instance, Yamashita et al. reported that in cultured human and mouse podocytes, the RLR activation in podocytes induced by dsRNA led to the expression of interferon I (IFN-I), IL-6 and cytoskeleton alteration, eventually resulting in podocyte effacement [77]. Apolipoprotein L1 (APOL1) is closely associated with primary kidney and kidney-related pathologies, which is identified in a recent large Phenome-Wide Association Study [78]. Animal study showed that long-term injections of recombinant APOL1 in mice increased expression of RIG-I in podocytes. Inhibition of RIG-I expression, either by using siRNA in vitro experiments or by adeno-associated virus short hairpins (AAV-sh) in vivo mouse experiments, decreased the expression of pro-inflammatory genes and alleviated podocyte injury and glomerular damage [79]. Similar results reported by Cristian et al. also showed that in human podocytes cultured with IFN- γ , inhibition of MDA5 down-regulated the expression of APOL1 and the subsequent IFN-I response [80]. The above published studies support the notion that the activation of RLRs can contribute to Sirtuin 1. However, whether RLRs exert effects or how RLRs act on podocyte in diabetes remains largely elucidated.

2.1.4 Involvement with ALRs

Absent in melanoma 2 (AIM2)-like receptors (ALRs) are mainly composed of N terminal containing pyrin domain (PYD) and C terminal containing hematopoietic interferon-inducible nuclear (HIN) domain. The AIM2 protein is highly conserved in the HIN-200 family and is recently identified as the only conserved protein in both humans and mice of all ALRs [81]. AIM2 is capable of identifying abnormal cytoplasmic dsDNA from pathogens and host cells. The activation of AIM2 initiates the assembly of AIM2 inflammasome, which is composed of AIM2, an apoptosis-associated speck-like protein containing a CARD and pro-caspase-1, and plays a pivotal role in protection against pathogen infection [82]. However, the activation of AIM2 also has harmful effects in some aseptic inflammatory diseases, such as chronic kidney disease and atherosclerosis [83,84]. Emerging studies found that the activation of AIM2 inflammasome is associated with the pathogenesis of insulin resistance in patients with T2DM and renal diseases. The mechanism of AIM2 in insulin resistance (IR) and renal diseases include promotion of chronic activation in inflammatory signaling pathway and conduction of the insulin signaling pathway. For example, in the mouse model of aldosterone-induced renal injury, overactivation of AIM2 aggravated endoplasmic reticulum stress (ERS) and inflammasome activation and fibrotic changes, while silencing of AIM2 could block inflammasome-mediated signaling pathway and relieve ERS and fibrotic changes, reducing proteinuria levels in vivo [71]. In patients with T2DM, the

increased circulating mtDNA levels that were closely related to mitochondrial dysfunction can be detected and are associated with AIM2 inflammasome-mediated chronic inflammation. The activation of AIM2 inflammasome elevates the expression of IL-1 β , promoting IR and T2DM development [85]. However, there is an argument on the harmful role of AIM2. Gong Z et al. showed that in animal experiments, compared to WT mice, AIM2 $^{-/-}$ mice tended to obesity and decreased energy expenditure, showing impaired brown fat function and increased fasting blood glucose and insulin resistance. The mechanism might be attributed to the AIM2 inhibition on the encoded protein 202, leading to the reduction of monocyte infiltration and lipogenesis, thus improving obesity and IR [86]. To date, there are few studies on AIM2 and podocyte injury in DN, and the seemingly contradictory role of AIM2 in metabolism means more studies are needed to confirm the association of AIM2 with diabetes and podocyte injury.

2.1.5 Involvement with CLRs

C-type lectin (CLEC) is originally defined as a protein that binds carbohydrates in a Ca $^{2+}$ -dependent manner, containing soluble and membrane-bound proteins. The C-type lectin receptors (CLRs) and the related C-type lectin-like receptors (CTLRs) are distinguished by their characteristics, the former have C-type lectin-like domain (CTLD) and Ca $^{2+}$ ion for glycan binding, the latter possess CTLD but lack any type of ion for ligand recognition [87]. It should be noted that the term CTLRs is also used for the broad definition containing all CLEC irrespective of carbohydrate recognition because many of these receptors' binding domain structures have not been fully elucidated [88]. CLRs can be further subdivided into three clusters based on their intracellular signaling motifs. Cluster 1 consists of activation spleen tyrosine kinase (Sky)-coupled CLRs with immunoreceptor tyrosine-based activating motifs (ITAM), such as Mincle, Dectin-1 and Dectin-2. Cluster 2 includes inhibitory CLRs with immunoreceptor tyrosine-based inhibition motif (ITIM) domains, such as DCIR. Cluster 3 contains a group of CLRs lacking typical signaling motifs, such as the Mannose receptor, DEC-205, and DC-SIGN. Upon ligand binding, CLRs stimulate intracellular cascades to induce the production of inflammatory cytokines and chemokines, consequently triggering immune response to pathogens. [89] Besides, they play essential roles in the physiological functions and pathological processes including immune homeostasis, immune defenses and immune surveillance [90].

Owing to the family's large repertoire, covering them all is beyond the scope of this review, and to date, it is difficult to make a definite conclusion on whether CLRs are protective or harmful to podocytes in kidney diseases. For instance, in a mouse model experiment, CLEC14A were expressed in podocytes and showed protection against podocyte injury via anti-inflammatory effect in mice with Adriamycin-induced focal segmental glomerular sclerosis (FSGS). In vitro cultured podocytes, overexpression of CLEC14A in podocytes have anti-inflammatory and anti-apoptosis effects through inhibition of high-mobility group box 1 protein (HMGB1) release, resulting in the suppression of NF- κ B signaling and early growth response protein 1 (EGR1) signaling [91]. Dectin-1 is another classic CTLR but displays pro-inflammatory effects. Upregulated expression of dectin-1 was observed in heart tissues in diabetic mice and localized to macrophages. Deficiency of Dectin-1 in mice displayed protection against diabetes-induced cardiac dysfunction, tissue fibrosis and inflammation, which means Dectin-1 is essential for cell activation and induction of inflammatory cytokines in diabetic

cardiomyopathy [92]. However, there are few studies concerning the effects of CLRs on podocytes in diabetic kidney disease. The differently displayed functions of CLRs may mean each member in the CLR family has its unique role in response to various diseases.

2.1.6 Involvement with cGAS/STING pathway

The cyclic GMP-AMP synthase and stimulator of interferon gene (cGAS/STING) pathway has recently been identified as a pivotal player in innate immunity. cGAS is a vital sensor of cytosolic DNA which can catalyze the second messenger 2'3-cGAMP, and consequently activates the endoplasmic reticulum membrane adapter STING (encoded by TMEM173), leading to the activation of TNK-binding kinase1 (TBK1) and interferon regulatory factor 3 (IRF3), which in turn induces the expression of type I interferons [34]. Activation of STING also leads to the activation of NF- κ B in a non-canonical mechanism and induces inflammatory cytokines expression [93,94]. In response to the abnormal leakage of cytosolic DNA, regardless from the invaded pathogen, damaged mitochondrial or genomic instability, the cGAS/STING pathway can be aberrantly activated and causes various diseases, including infectious diseases, autoimmune diseases, tumors, organ fibrosis, and neurodegenerative diseases [95-97].

The bulk of studies linking the activation of cGAS/STING pathway to kidney disease have focused on the podocyte. It has been reported that podocyte expresses components of cGAS/STING pathway at the early stage of kidney dysfunction and the activation of this pathway can cause podocyte injury [98]. For instance, in a mouse model of diabetic kidney disease (DKD), treatment with STING agonist triggered podocytes loss and resulted in susceptibility to albuminuria. Similar results in vitro cultured murine and human podocytes were observed [99]. Metabolism dysfunction in diabetes can also activate this pathway. Study from Zang et al. showed that lipotoxicity in a mouse model with DKD caused mitochondrial damage and mtDNA leakage into the cytoplasm, promoting the activation of the cGAS/STING pathway which induced the secretion of inflammatory cytokines, thereby resulting in podocyte injury [98]. On the other hand, pharmacologic inhibition or genetic deletion of this pathway protected against podocyte injury and the progression of kidney disease in mice with DKD, which supports the cGAS/STING pathway as a potential therapeutic target for DKD [99].

Several studies concern the role of cGAS/STING pathway in cell crosstalk between podocyte injury and other glomerular cell damage. Qi et al. presented evidence that in DKD-susceptible mice, mtDNA from diabetes-induced mitochondrial dysfunction and stress contributed to glomerular injury indirectly through glomerular endothelial cells (GECs) and was associated with podocytes depletion, resulting in proteinuria [100]. Casalena et al. showed that in supernatant transfer experiments, diabetic serum caused mitochondrial dysfunction and superoxide release in GECs and secreted factors from dysfunctional GECs induced podocytes apoptosis [101]. Similar phenomena in which crosstalk between podocytes and GECs, can also be found in other kidney diseases, such as focal segmental glomerulosclerosis [102] and APOL1 renal risk variants [103]. Mesangial cells (MCs) also display markers of mitochondrial damage induced by diabetes, which means the leakage of mtDNA and the subsequent activation of the cGAS/STING pathway may operate in some instances of DN, mediating with podocyte injury [104].

2.2 Adaptive immune response associated with podocyte injury

Besides their role in innate immunity, podocytes are capable of promoting the adaptive immune response. It has been known that podocytes possess properties of antigen-presenting cells (APC), such as expression of major histocompatibility complex (MHC) I/II and B7-1 [12]. MHC I and MHC II respectively contribute to the activation of CD8⁺ T cells and CD4⁺ T cells, while the B7-1 is a co-stimulatory molecule for T cells. Interestingly, B7-1 (also known as CD80) is part of the B cells and APC repertoire, providing the second signal to T cells to allow amplification of response to antigens. B7-1 also directly binds to CD28 on the T cell membrane to induce a positive co-stimulatory signal [105,106]. Podocyte's immune properties play an essential role in podocyte injury in kidney diseases. For example, Li et al. showed that B7-1 induction was related to podocyte injury in DN, while inhibition of B7-1 by an agent could efficaciously protect podocytes through prevention of systemic T-cell activation [107]. A study from Shan et al. reported that in mouse model with ovalbumin plus IFN- γ and IL-17 renal injection, increased levels of MHC-I, MHC-II and B7-1 were expressed in podocytes. Podocytes could uptake and process ovalbumin, then presented the ovalbumin peptide on the cell surface, and consequently recruited specific T cells and activated T cell proliferation and inflammatory cytokine secretion, which in turn resulted in podocyte injury and developed nephropathy [108]. Li et al. presented that renal CD8⁺ tissue-resident-memory T (CD8⁺TRM) cells were activated through the interleukin-15 (IL-15) signaling pathway and played a crucial role in mediating local immune response in diabetic kidney disease. Targeting on CD8⁺TRM cells by pharmacological inhibition of IL-15 signaling protected against podocyte injury [109]. A study performed by Chen et al. showed that in the kidney of patients with crescentic glomerulonephritis, CD8⁺ T cells invaded the disrupted Bowman's space and directly contacted podocytes, resulting in podocyte destruction. However, in a healthy mouse model with an intact Bowman's capsule, the infiltration of CD8⁺ T cells was prevented from interacting with podocytes, suggesting that the normal Bowman's space provided a protective niche for podocytes from cytotoxic CD8⁺ T cells and thereby protected renal function [110]. This study may partly explain the reality that the use of reno-protective agents in clinical activity always has prominent effects against proteinuria and renal dysfunction in the early stage, but not in the end-stage kidney disease. A recent study using pod I-Ppol mice as a model showed that DNA damage in podocytes caused alterations of DNA methylation in blood cells and was associated with the proliferation of CD8⁺ memory T cells in the kidney, leading to proteinuria and glomerulosclerosis, which suggests the presence of crosstalk between podocyte injury and cytotoxic T cell [111]. The above evidence suggest the adaptive immune response systemically and locally contributes to the immunopathological process of podocyte injury in DN.

2.3 Involvement with Sirtuin family [112-116]

Silent information regulator 2-related enzymes (Sirtuin or SIRT) are evolutionarily conserved class III histone deacetylases, which depend on nicotinamide adenine dinucleotide (NAD⁺) [117]. Studies have shown

that there are seven members (Sirtuin1-Sirtuin7) of the sirtuin family in mammals, of which Sirtuin1 is widely expressed in podocytes [118] and plays a critical role in mediation of cell apoptosis [119], autophagy [120], energetic homeostasis [121], mitochondrial biogenesis [122] as well as immune response [123]. Recent report by Mourits et al. illustrated the closed association of Sirtuin1 and immunology. Sirtuin1 genetic polymorphisms could affect inflammatory cytokine production of human peripheral blood mononuclear cells (PBMCs) in response to various stimuli via modulation of gene transcription [124]. Study from Chen et al. showed that Sirtuin1 was involved in the regulation of cell differentiation, activation and function in innate and adaptive immune cells [125]. Those evidence suggest Sirtuin1 might be relevant to the immune related mechanism in podocyte injury. Experimentally, Q et al. presented that in diabetic OVE26 mice with established albuminuria, overexpression of Sirtuin1 in podocytes attenuated the progression of diabetic glomerulopathy. Treatment with selective Sirtuin1 agonist in vitro cultured human podocytes protected against high glucose-induced mitochondrial injury by deacetylation of PGC-1 α and activation of PPAR γ -targeted gene expression, and in OVE26 mice also showed a similar result, displaying a marked reduction of albuminuria and glomerular injury [126]. Another study performed by Jiang et al. showed that in aldosterone-induced podocyte injury mice model, Sirtuin1 protected renal function through suppressing the activation of NLRP3 inflammasome, showing the greater urinary albumin excretion in podocyte-specific Sirtuin1 knockout mice than that in wild-type mice. In the cultured podocytes treated with aldosterone, overexpression of Sirtuin1 inhibited NLRP3 activation and protected against podocyte injury [127]. In addition, Sirtuin1 was also found to have renoprotective effects through improvement of the organization of actin cytoskeleton [128] and insulin resistance [129] in podocytes. Collectively, Sirtuin1 might be a potential target for podocyte protection in DN, however, more studies are needed to further elucidate the role of Sirtuin1 in the immune related podocyte injury in DN.

3 Mechanisms of agents for DN: especially focus on podocyte protection

Recently, published guideline for diabetes from the American Diabetes Association (ADA) recommend the antidiabetic agents [130]. In this guideline, sodium-glucose cotransporter 2 (SGLT2) inhibitor and glucagon-like peptide 1 receptor agonist (GLP-1RA) have direct renal protection beyond the glucose-lowering effect. ACE inhibitor (ACEI) or angiotensin II receptor blocker (ARB) are suggested for diabetic patients with hypertension, except for those who are normotensive with or without high albuminuria. Because evidence from two long-term, double-blind studies demonstrate no reno-protective effect of either ACEI or ARB among patients with type 1 and type 2 diabetes who are normotensive. Finerenone is currently the only nonsteroidal MRA with proven clinical kidney and cardiovascular benefits. The following section discusses the mechanism of these agents, especially focusing on the protection of podocyte in diabetes (as shown on table 1).

Category	Name of agents	Protective mechanism for podocyte	References
SGLT2i	canagliflozin, empagliflozin, dapagliflozin, ertugliflozin, etc.	limitation of cytoskeletal remodeling reduction of nephrin loss increase of autophagy improvement of lipid accumulation alleviation in inflammation	[105] [105] [106] [107] [111] [114][115]
GLP1-RA	dulaglutide, liraglutide, semaglutide, exenatide, lixisenatide, etc.	alleviation in inflammation reduction of oxidative stress mediation of natriuresis	[123][124] [129][130] [134]
ACEI/ARB	benazepril ,captopril, valsartan, losartan, etc.	improvement of podocyte regeneration modulation of progenitor cell proliferation promotion of podocyte repair	[143] [143] [144]
MRA	finerenone	suppression of oxidative stress prevention of podocyte effacement inhibition of Rac1-MR pathway	[153] [153] [154]

SGLT2i, sodium-glucose cotransporter 2

GLP1-RA, Glucagon-like peptide 1 receptor agonist

ACEI, Angiotensin-converting enzyme inhibitor

ARB, angiotensin II receptor blocker

MRA, Mineralocorticoid receptor antagonist

Table 1: Mechanisms of agents recommended for diabetic nephropathy by guideline

3.1 SGLT2 inhibitor

The balance between glomerular filtrate and reabsorption is one of the pivotal physiologic functions of the kidney. In euglycemia with normal GFR, the proximal tubules reabsorb almost all tubular nutrients, including glucose [131]. The bulk of glucose uptake occur in the early proximal tubule which contain high-capacity sodium-glucose cotransporter 2 (SGLT2). It has been shown that SGLT-2 accounts for all glucose reabsorption in the early proximal tubule and nearly 97% of total renal glucose reabsorption [132]. SGLT2 inhibitors are a class of antihyperglycemic drugs approved for T2DM. These drugs inhibit renal glucose reabsorption in the early proximal tubule, hence increasing urinary excretion and lowering blood glucose. Several large-scale clinical trials confirmed that SGLT2 inhibitors have pleiotropic properties beyond blood glucose-lowering effects, including beneficial renal and cardiac effects [133]. Multiple mechanisms contribute to the reno-protective effects of SGLT2 inhibitors in T2DM, such as restoration of tubule-glomerular feedback to reduce glomerular hyperfiltration, protection against inflammation, hypoxia, oxidative stress and fibrosis [134]. Direct action on podocyte is another mechanism for reno-protective function by SGLT-2 inhibitors [135].

In recent years, the expression of SGLT2 has been found in podocyte and increase in diabetic mice. For example, Cassis et al. showed that in a mouse model with proteinuria induced by bovine serum albumin (BSA), the expression of SGLT2 was upregulated in kidney after BSA injection. SGLT2 inhibition with dapagliflozin reduced the expression of SGLT2 in podocytes and protected against podocyte dysfunction through the

limitation of cytoskeletal remodeling and loss of nephrin, supporting a potential mechanism that SGLT2 inhibitors directly participated in maintaining the structure and function of podocytes to prevent the progression of DN [136].

The downregulated level of autophagy in DKD has been observed and is associated with increased albuminuria and kidney dysfunction. For example, in a mouse model of type 2 diabetes performed by Anton et al, SGLT2 inhibitor empagliflozin enhanced podocyte autophagy level, showing the increased volume density of autophagosomes and autolysosomes in podocytes, thus attenuating podocyte effacement and urinary albumin excretion in db/db mice [137]. A vitro cell experiment presented by Lei et al. also illustrated a similar result. The cultured podocytes induced by advanced glycation end products (AGEs) displayed inhibition of autophagy, showing the reduced markers of autophagosome formation such as light chain 3 phospholipid conjugate (LC3II) and Beclin-1, while treatment with SGLT2 inhibitor dapagliflozin restored the expression of Beclin-1 and LC3II, promoting autophagosome turnover and autophagosome degradation, thus exerted a protective effect on podocytes [138]. These observations support that SGLT2 inhibitors exert protective effects in podocyte via the mediation of cell autophagy. The relationship between dyslipidemia and podocyte injury is well documented. Indeed, Apolipoprotein-1 (APOL1) which is considered as a high risk genetic variant in kidney disease, is highly expressed in podocyte [139]. Analysis from experimental studies and renal biopsies of patients with DN showed that sterol regulatory element-binding proteins (SREBPs), essential transcription factors associated with lipid metabolism, are increased in lipid droplets loaded podocytes [140]. In this

regard, podocyte injury could be induced by lipid imbalance. SGLT2 inhibitors can improve podocyte lipid content to maintain podocyte health. Mengyuan et al. reported that in an experimental mouse model with Alport syndrome, SGLT2 inhibitor empagliflozin inhibited the utilization of glucose and pyruvate as metabolic substrate, leading to the reduction of podocyte lipotoxicity and renal cortical lipid deposition, thus improving kidney function and prolonged mice survival [141]. Dong et al. described that in mice with Western diet-induced obesity, SGLT2 inhibitor dapagliflozin decreased lipid accumulation in podocytes, which was associated with decreased SREBP-1c expression [142]. These observations support the role of SGLT2 inhibitors in the improvement of lipid metabolism in podocyte.

It is well known that low-grade inflammation is one of vital features of diabetic kidney disease. Several clinical trials illustrated that SGLT2 inhibitors have demonstrated the downregulation of circulating inflammatory markers such as IL-6, TNF- α , and IFN- γ in diabetic patients [143,144]. Experimentally, *in vitro* cultured macrophages isolated from type 2 diabetic patients, SGLT2 inhibitor empagliflozin suppressed the secretion of IL-1 β and the activation of NLRP3 inflammasome [145]. Treatment with SGLT2 inhibitor canagliflozin for 2 years in diabetic patients illustrated a decrease in levels of genes encoding TNF receptor1 and fibronectin1, which supported that SGLT2 inhibitors might alleviate molecular processes related to inflammation [146]. In line with this proposal, a study from nephrectomy rats with established CKD, SGLT2 inhibitor empagliflozin suppressed fibrosis-promoting M2 macrophage polarization and inhibition of profibrotic marker, thereby improving kidney function [147].

The above evidence support that SGLT2 inhibitors exert protective effects on podocyte through several key aspects, including maintenance of cell integrity, upregulated autophagy, improved lipid deposition and decreased inflammation. However, whether SGLT2 inhibitors have direct immune effects on mediation of podocyte injury in diabetes remains largely elucidated, thereby they are worthy of exploration in the future.

3.2 GLP-1RA

Glucagon-like peptide-1 (GLP-1) is a thirty-amino acid peptide, which is released from gut enteroendocrine cells in response to the stimulation of luminal nutrients after meal ingestion. It is well known that GLP-1 exerts metabolism actions in maintaining glucose homeostasis mainly via secretion of insulin and inhibition of glucagon release as well as weight loss [148]. Glucagon-like peptide 1 receptor agonists (GLP-1RAs) are commonly used in the treatment of patients with type 2 diabetes. Numerous evidence suggest that GLP-1RAs have multiple benefits to protect the function and structure of the kidney in diabetes in addition to their glucose-lowering effects [149]. Renal outcomes observed in recent large trials showed that GLP-1RAs reduced urinal albumin excretion and weight loss, which are associated with improved survival [150-152]. Studies in models of T2DM and/or obesity illustrated that GLP-1RAs exert anti-inflammatory, anti-oxidative stress and mediate natriuresis to prevent the progression of DN and podocyte injury.

Inflammatory disorder drives the development of diabetes and its complications. A study conducted in 4213 people showed that high-sensitivity C-reactive protein (hs-CRP), a marker of systemic inflammation, is elevated in patients with T2DM [153], while an analysis from systematic review illustrated that treatment with GLP-1RAs is associated with a significant reduction of CRP, as well as other

biomarkers of inflammation [154]. Anti-inflammatory activities can be found in the treatment with GLP-1RAs, including direct effects on leukocyte activation, macrophage polarization, NLRP3, and NF- κ B dependent signaling pathway [155]. It has been illustrated that podocyte expresses GLP-1 receptor and GLP-1RA can act on podocyte to exert anti-inflammatory effects and prevent podocyte injury in diabetes [156]. For example, Li et al. presented that in the cultured podocytes treated with high glucose, GLP-1RA liraglutide increased podocyte survival by inhibiting the NLRP3 inflammasome pathway and reducing the secretion of IL-1 β and IL-18 [157]. Moreover, studies also showed that GLP-1RA liraglutide inhibited NF- κ B activation mediated by TNF- α in podocytes of obese mice and could induce distinct transcriptional changes in podocytes through inhibition of the receptor for AGEs-induced inflammation [158,159].

Oxidative stress plays a pivotal role in the mechanism of podocyte injury and DN. Numerous pieces of evidence show that GLP-1RAs have antioxidative activities independent of lowering glucose. For instance, a pilot study performed in patients with type 2 diabetes displayed that treatment with GLP-1RAs was related to reduced circulating markers of oxidative stress, such as lipid hydroperoxides, nitro-tyrosine and malondialdehyde, as well as downregulated NADPH oxidase and endogenous superoxide generation [160]. It is reported that reactive oxygen species (ROS) production through NADPH oxidases damage podocyte in diabetes, which contribute to the alterations of nephrin and podocalyxin on the podocyte surface and result in glomerular filtration dysfunction. ROS production in the kidney is due to the action of NOXs, including NOX1, NOX2, and NOX4. Experimentally, *in vitro* high glucose cultured podocytes, inhibition of the mammalian target of rapamycin (mTOR) could decrease the expression of NOX1 and NOX4 and podocyte apoptosis [161]. Moreover, targeting NOX4-derived H₂O₂ could alleviate podocyte injury in DN through elevation of podocyte calcium, which was associated with transient receptor potential channel 6 (TRPC6) dependent calcium influx [162]. GLP-1RA liraglutide selectively inhibited TRPC6 expression and subsequently reduced ROS production and protected podocyte structure and function in the DKD rat model [163]. In clinical studies, treatment with GLP-1RA liraglutide for 12 weeks in patients with DN increased the urine level of glutathione peroxidase, an important intrinsic antioxidant enzyme that eliminates oxidative stress [164].

Mediation in natriuresis shows another mechanism by which GLP-1RAs have renal-protective effects in diabetes. Natriuresis is the increased urinary excretion of sodium, which can result in a reduction in blood volume and a decreased workload on the heart. In mechanism, GLP-1RAs phosphorylate and inactivate the Na⁺/H⁺ exchanger 3 in the proximal tubule, consequently decrease Na⁺ reabsorption and promote natriuresis [165]. GLP-1RAs can also promote the secretion of atrial natriuretic peptide (ANP) in cardiac cells to increase urinary excretion of sodium and water [166]. In summary, GLP-1RA is capable of exerting protective effects on podocyte through various factors and it is necessary to further explore its immunity-related mechanism on podocyte.

3.3 Acei/Arb

The functions of the renin-angiotensin-system (RAS) have been well documented in both blood pressure regulation and renal disease development. Angiotensin II (Ang II) is the major bioactive product produced by RAS with the aid of a sequence of enzymatic processes. It exerts various physiologic effects to maintain cell and organ function,

including renal cell growth, mitogenesis, apoptosis, migration and differentiation. Moreover, Ang II also induces the activation of multiple intracellular signaling pathways that are involved in renal damage [167,168].

Angiotensin-converting enzyme inhibitor (ACEI) and angiotensin II receptor blocker (ARB) have been used to control hypertension in clinical treatment. Data from several large clinical trials have shown that in addition to lowering blood pressure and cardiovascular protective effects, ACEI and ARB have impressively reno-protective benefits and have been recommended for the treatment in patients with DKD by several guidelines [169,170]. The mechanisms of reno-protection include anti-inflammatory and anti-fibrotic effects, downregulation of sympathetic activity and reduction of aldosterone secretion.

It has been found that podocyte expresses functional components of the RAS, such as angiotensinogen, renin, ACE type1 and the AT1 and AT2 angiotensin receptor subtypes [171]. It is reported that increased Ang II is a vital risk factor for the progression of renal disease and is associated with the reduction of nephrin and podocin expression in podocyte. The loss of nephrin and podocin is causative for podocyte damage, resulting in marked albuminuria and increased podocyte apoptosis [168,172]. ACE inhibition and the angiotensin blockade in the form of ACEI and ARB have been proven to be useful in preventing the progression of DN and podocyte dysfunction. For instance, Zhu et al. showed that in adult rat model with glomerular sclerosis, treatment with long-term (24 weeks) ARB significantly improved the podocyte density in glomeruli through the improvement of podocyte regeneration from parietal epithelial cells transition, as well as leading to reduction of proteinuria and prolonged rat survival [173]. Another study performed by Kelly et al. displayed a similar result in an ob/ob mouse model with diabetic nephropathy, treatment with ARB increased glomerular podocyte number and lowered proteinuria compared with diabetic ob/ob controls [174]. Research also demonstrated that ACEI therapy could sustain glomerular repair and improve kidney function via the promotion of podocyte repair, limitation of crescent generation, and modulation of progenitor cell proliferation [175]. Taken together, these findings strengthen the concept that ACEI and ARB have beneficial effects on podocyte protection and kidney function, but the mechanisms involved in DN require a multi-pronged strategy.

3.4 Mineralocorticoid receptor antagonists (MRA)

The aldosterone, the physiological main ligand of the mineralocorticoid receptor (MR), is the final component of the renin-angiotensin-aldosterone system (RAAS). Aldosterone and MR systems have been considered pivotal players in the regulation of fluid volume, electrolytes, hemodynamic homeostasis and blood pressure. MR is a subfamily of nuclear receptors that belongs to the steroid hormone receptors family, acting as intracellular receptors and nuclear transcription factors [176]. Accumulating studies implicate the abnormal reaction in aldosterone and MR system is a pathogenic factor in kidney injury independently of its physiologic functions, and the blockade of MR can prevent the detrimental effects on cell injury [177]. MR antagonists (MRAs) elicit benefits for CKD through the inhibition of aldosterone and the MR system [178]. The MR antagonists (MRAs) include steroidal MRAs and non-steroidal MRAs. In the past years, spironolactone and eplerenone were the earlier steroidal MRAs that are still widely used to date. These steroidal MRAs have been proven their benefits and utilities in heart failure and proteinuria in patients treated with an RAAS blockade by

landmark trials. However, these agents remain underutilized in patients with CKD, because of the concern for hyperkalemia and worsening GFR [179].

Finerenone is a non-steroidal MR antagonist (MRA) with great cardiac and renal benefits and less effect on inducing hyperkalemia which is proven by landmark trials [180]. Finerenone is developed using a special chemical structure and optimized to create a bulky MRA without any activity at the L-type calcium channel, which may exert its effects more efficiently than a steroidal MRA [181]. The recently published guidelines from the American Diabetes Association (ADA) and the American Association of Clinical Endocrinology (AACE) recommended the addition of finerenone to standard treatment in patients with DKD [182,183]. In mechanism, the biological actions of aldosterone are mainly through MR. MR expression is present in podocyte and aldosterone appears to have direct deleterious effects on podocyte. Animal research found that rats continuously infused with aldosterone increased ROS production through NADPH oxidase and induced podocyte foot process effacement. Treatment with MRA suppressed the oxidative stress and prevented podocyte effacement [184]. Moreover, in the type2 diabetic mouse model with DN progression, the activation in RAS-related C3 botulinus toxin substrate 1 (Rac1) and MR pathway contributed to podocyte injury, while treatment with finerenone inhibited Rac1-MR pathway and protected against podocyte injury [185]. These evidence support finerenone as a vital therapeutic method in the inhibition of DN development and podocyte injury.

Conclusion

Podocyte plays a crucial role in renal physiology and pathology. For years, the exploration of mechanisms in DN has illustrated the potential for the discovery of innovative therapeutic targets. Due to the podocyte's location in the last barrier of the glomerular capillary, it has been considered as a valuable target to prevent proteinuria in DN, thereby studies focusing on podocyte protection have attracted extensive attention worldwide. Therapy aimed at preventing or limiting podocyte injury have ever been considered as a promising strategy with major potential clinical and economic benefits. Recently, the therapeutic options for podocyte protection mainly include immunosuppressive drugs and novel biological agents. However, immunosuppressive drugs, such as glucocorticoids and calcineurin inhibitors, primarily target immune-mediated inflammation in the kidney, their use may lead to serious side effects, especially after prolonged use, such as infection, hypertension and osteoporosis [186]. Novel biological agents known for its high specificity, minimal side effects, and durable benefits is developed for the treatment of podocyte disease. They also have side effects that can't be ignored including infusion response, production of anti-drug antibodies, immunogenicity and hypogammaglobulinemia. Furthermore, the optimization of parameters for immunotherapy need to improve its clinical applicability and effectiveness [187]. To date, it is still hard to exactly outline the mechanisms of podocyte injury in DN, and agents specifically targeting on podocyte protection have not been fully developed for patients with DN in clinical treatment.

As described earlier in this review, numerous preclinical studies support the promising potential of podocyte protection, especially focusing on immune-related therapies. But moving from proof of concept to clinical evaluation still has a long way to go. The molecular mechanisms of podocyte injury in DN continue to be explored, because except for immune-related injury, glucotoxicity, lipotoxicity and renal

hemodynamic change in diabetes also play vital roles in podocyte injury, and the complicated crosstalk between these factors in DN may exist. These obstacles make researchers difficult to develop agents specially targeting on podocyte protection. Hopefully, further studies will provide more insights into the mechanisms underlying podocyte injury, likely providing improved therapeutics for podocyte protection in DN.

Fund Program: The Construction of Major Subject [Grant no (YNZDXK202401)] of Huadu District People's Hospital of Guangzhou ; Guangzhou Science and technology plan project (202201011649) ; The Internal Fund of Huadu District People's Hospital of Guangzhou (2022C04)

References

- Manpreet K Sagoo, Luigi Gnudi. (2020).Diabetic Nephropathy: An Overview. *Methods Mol Biol.*:2067:3-7.
- American Diabetes Association Professional Practice Committee. Chronic Kidney Disease and Risk Management: Standards of Care in Diabetes-.*Diabetes Care.* Jan 1;47(Suppl 1): S219-S230
- American Diabetes Association Professional Practice Committee. Diabetes Care. Diagnosis and Classification of Diabetes: Standards of Care in Diabetes-2024.2024 Jan 1;47(Suppl 1):S20-S42.
- Rhee CM, Leung AM, Kovesdy CP, Lynch KE, Brent GA, Kalantar-Zadeh K. Updates on the management of diabetes in dialysis patients.
- Federica Barutta, Stefania Bellini, Gabriella Gruden. (2022).Mechanisms of podocyte injury and implications for diabetic nephropathy. *Clin Sci (Lond).* Apr 14;136(7):493-520.
- Jiang S, Luo M, Bai X, Nie P, Zhu Y, Cai H, Li B, Luo P. (2022).Cellular crosstalk of glomerular endothelial cells and podocytes in diabetic kidney disease. *J Cell Commun Signal.* 2022. Sep;16(3):313-331.
- Jamie S Lin, Katalin Susztak. (2016).Podocytes: the Weakest Link in Diabetic Kidney Disease? *Curr Diab Rep.* May;16(5):45.
- Anni Jiang, Anni Song, Chun Zhang. (2022).Modes of podocyte death in diabetic kidney disease: an update. *Nephrol.* 2022 Jul;35(6):1571-1584.
- Pichler R, Afkarian M, Dieter BP, Tuttle KR.(2017). Immunity and inflammation in diabetic kidney disease: translating mechanisms to biomarkers and treatment targets. *Am J Physiol Renal Physiol.* 2017 Apr 1;312(4):F716-F731.
- Pierre-Louis Tharaux, Tobias B Huber. (2012).How many ways can a podocyte die? *Semin Nephrol.* Jul;32(4):394-404..
- Xia H, Bao W, Shi S. (2017).Innate Immune Activity in Glomerular Podocytes. *Front Immunol.* Feb 8;8:122.
- Andreas Goldwisch 1, Miriam Burkard, Martha Olke, Christoph Daniel, Kerstin Amann, Christian Hugo, Christian Kurts, Alexander Steinkasserer, André Gessner. Podocytes are nonhematopoietic professional antigen-presenting cells. *J Am Soc Nephrol.* May;24(6):906-16.
- Shadia A Fathy, Mohamed R Mohamed, Mohamed A M Ali, Ashraf E El-Helaly , Abdulnabi T Alattar.(2019). Influence of IL-6, IL-10, IFN- γ and TNF- α genetic variants on susceptibility to diabetic kidney disease in type 2 diabetes mellitus patients. *Biomarkers.* Feb;24(1):43-55.
- Chia-Chao Wu, Jin-Shuen Chen, Kuo-Cheng Lu, Chun-Chi Chen, Shih-Hua Lin, et.al (2010). Aberrant cytokines/chemokines production correlate with proteinuria in patients with overt diabetic nephropathy. *Clin Chim Acta.* May 2;411(9-10):700-704.
- Zatterale F, Longo M, Naderi J, Raciti GA, Desiderio A, Miele C, Beguinot F. Chronic (2020).Adipose Tissue Inflammation Linking Obesity to Insulin Resistance and Type 2 Diabetes. *Front Physiol.* Jan 29;10:1607.
- Chaochen Wang, Hiroshi Yatsuya, Koji Tamakoshi, Mayu Uemura, Yuanying Li, et.al (2013).Positive association between high-sensitivity C-reactive protein and incidence of type 2 diabetes mellitus in Japanese workers: 6-year follow-up. *Diabetes Metab Res Rev.* Jul;29(5):398-405
- Tashiro K, Koyanagi I, Saitoh A, Shimizu A, Shike T, Ishiguro C, Koizumi .et.al (2002).Urinary levels of monocyte chemoattractant protein-1 (MCP-1) and interleukin-8 (IL-8), and renal injuries in patients with type 2 diabetic nephropathy. *J Clin Lab Anal.* x;16(1):1-4. doi: 10.1002/jcla.2057.
- Bancha Satirapoj, Rattanawan Dispan, Piyanuch Radinahamed, Chagriya Kitiyakara. (2018).Urinary epidermal growth factor, monocyte chemoattractant protein-1 or their ratio as predictors for rapid loss of renal function in type 2 diabetic patients with diabetic kidney disease. *BMC Nephrol.* Sep 21;19(1):246.
- Fiorentino M, Bolignano D, Tesar V, Pisano A, Biesen WV, Tripepi G, D'Arrigo G, Gesualdo L; (2017).Renal biopsy in patients with diabetes: a pooled meta-analysis of 48studies. *Nephrol Dial Transplant.* Jan 1;32(1):97-110
- Xue Tong, Qun Yu, Ghada Ankawi, Bo Pang, Bo Yang, Hongtao Yang. (2020).Insights into the Role of Renal Biopsy in Patients with T2DM: A Literature Review of Global Renal Biopsy Results. *Diabetes Ther.* Sep;11(9):1983-1999
- Packialakshmi B, Stewart JJ, Burmeister DM, Chung KK, Zhou X. (2020).Large animal models for translational research in acute kidney injury. *Ren Fail.* Nov;42(1):1042-1058.
- Betz B, Conway BR. ((2014).).Recent advances in animal models of diabetic nephropathy. *Nephron Exp Nephrol.*;126(4):191-195.
- Cuiping Zhang, Chunchun Xiao, Peng Wang, Wenhua Xu, Aimei Zhang,et.al (2014). The alteration of Th1/Th2/Th17/Treg paradigm in patients with type 2 diabetes mellitus: Relationship with diabetic nephropathy. *Hum Immunol.* Apr;75(4): -96.
- Liang Li, Wei Tang, Yan Zhang, Meng Jia, Limei Wang, Quanxin Li, Qingsheng Han, Xiuping Peng, Yusheng Xie, Jichao Wu, Ziyang Wang, Junhui Zhen, Xiaojie Wang, et.al (2022).tissue-resident memory CD8+ T cells in the kidney is a potential therapeutic strategy to ameliorate podocyte injury and glomerulosclerosis. *Mol Ther.* Aug 3;30(8):2746-2759.
- Xiao-Qian Li, Dong-Yuan Chang, Min Chen, Ming-Hui Zhao. (2019).Deficiency of C3a receptor attenuates the development of diabetic nephropathy. *BMJ Open Diabetes Res Care.* 2019 Nov 15;7(1):e000817.
- Yutian Lei, Satish K Devarapu, Manga Motrapu, Clemens D Cohen, Maja T Lindenmeyer, Solange Moll, Santhosh V Kumar, Hans-Joachim Anders.(2019). Interleukin-1 β Inhibition for Chronic Kidney Disease in Obese Mice With

- Type 2 Diabetes. *Front Immunol.* May 29;10:1223.
27. Ran Nakamichi, Kaori Hayashi, Hiroshi Itoh.(2021). Effects of High Glucose and Lipotoxicity on Diabetic Podocytes. *Nutrients.* Jan 15;13(1):241
 28. Marita Liebisch, Gunter Wolf. (2020).AGE-Induced Suppression of EZH2 Mediates Injury of Podocytes by Reducing H3K27me3. *Am J Nephrol.*;51(9):676-692.
 29. Federica Barutta, Stefania Bellini, Gabriella Gruden.(2022). Mechanisms of podocyte injury and implications for diabetic nephropathy. *Clin Sci (Lond).* Apr 14;136(7):493-520.
 30. Anni Jiang, Anni Song, Chun Zhang. Modes of podocyte death in diabetic kidney disease: an update. *Nephrol.* 2022 Jul;35(6):1571-1584
 31. L Evan Reddick, Neal M Alto. (2014).Bacteria fighting back: how pathogens target and subvert the host innate immune system. *Mol Cell.* Apr 24;54(2):321-328.
 32. Wada J, Makino H.Innate (2015).immunity in diabetes and diabetic nephropathy. *Nat Rev Nephrol.* 2016 Jan;12(1):13-26..
 33. Sydney C W Tang, Wai Han Yiu. (2020).Innate immunity in diabetic kidney disease. *Nat Rev Nephrol.* Apr;16(4):206-222.
 34. Chen C, Xu P. (2023).Cellular functions of cGAS-STING signaling. *Trends Cell Biol.* Aug;33(8):630-648. doi: 10.1016/j.tcb.2022.11.001. Epub 2022 Nov 24.
 35. Odendall C, Kagan JC. Activation and pathogenic manipulation of the sensors of the innate immune system. *Microbes Infect.* 2017 Apr-May;19(4-5):229-237. doi: 10.1016/j.micinf.2017.01.003. Epub 2017 Jan 14.
 36. Behzadi P, García-Perdomo HA, Karpiński TM.(2021). Toll-Like Receptors: General Molecular and Structural Biology. *J Immunol Res.*May 29;2021:9914854. doi: 10.1155/2021/9914854. eCollection
 37. Kang JY, Lee JO. (2011).Structural biology of the Toll-like receptor family. *Annu Rev Biochem.*;80:917-41. doi: 10.1146/annurev-biochem-052909-141507.
 38. Miriam C Banas, Bernhard Banas, Kelly L Hudkins, Tomasz A Wietecha, Masayuki Iyoda, et.al (2008).Alpers. TLR4 links podocytes with the innate immune system to mediate glomerular injury. *J Am Soc Nephrol.* 2008 Apr;19(4):704-13. doi: 10.1681/ASN.2007040395. Epub Feb 6.
 39. Panchapakesan U, Pollock C.(2018). The role of toll-like receptors in diabetic kidney disease. *Curr Opin Nephrol Hypertens.* Jan;27(1):30-34. doi: 10.1097/MNH.0000000000000377.
 40. Wenduona Bao, Hong Xia, Yaojun Liang, Yuting Ye, Yuqiu Lu, Xiaodong Xu, et.al (2016).Shaolin Shi. Toll-like Receptor 9 Can be Activated by Endogenous Mitochondrial DNA to Induce Podocyte Apoptosis. *Sci Rep.* Mar 3:6:22579. doi: 10.1038/srep22579.
 41. Alana A Shigeoka, Todd D Holscher, Andrew J King, Frank W Hall, William B Kiosses, Peter S Tobias,et.al(2016). TLR2 is constitutively expressed within the kidney and participates in ischemic renal injury through both MyD88-dependent and -independent pathways. *J Immunol.* May 15;178(10):6252-8. doi: 10.4049/jimmunol.178.10.6252.
 42. Sharbafi MH, Assadiasl S, Pour-Reza-Gholi F, Barzegari S, Mohammadi Torbati P, et.al(2019).TLR-2, TLR-4 and MyD88 genes expression in renal transplant acute and chronic rejections. *Int J Immunogenet.* Dec;46(6):427-436. doi: 10.1111/iji.12446. Epub 2019 Jul 9.
 43. Mudaliar H, Pollock C, Panchapakesan U. (2014).Role of Toll-like receptors in diabetic nephropathy. *Clin Sci (Lond).* May;126(10):685-94. doi: 10.1042/CS20130267.
 44. Harshini Mudaliar, Carol Pollock, Jin Ma, Huiling Wu, (2014). Steven Chadban, Usha Panchapakesan. The role of TLR2 and 4-mediated inflammatory pathways in endothelial cells exposed to high glucose. *PLoS One.* Oct 10;9(10):e108844. doi: 10.1371/journal.pone.0108844. eCollection 2014.
 45. Jin Ma, Steven J Chadban, Cathy Y Zhao, Xiaochen Chen, Tony Kwan,et.al. (2014).TLR4 activation promotes podocyte injury and interstitial fibrosis in diabetic nephropathy. *PLoS One.* 2014 May 19;9(5):e97985. doi: 10.1371/journal.pone.0097985. eCollection 2014.
 46. Jin Ma, Huiling Wu, Cathy Y Zhao, Usha Panchapakesan, Carol Pollock, Steven J Chadban. (2014).Requirement for TLR2 in the development of albuminuria, inflammation and fibrosis in experimental diabetic nephropathy. *Int J Clin Exp Pathol.* 2014 Jan 15;7(2):481-495.
 47. Fanglin Li, Ningyu Zhang, Zhiming Li, Lihua Deng, Jianjie Zhang, Yunfeng Zhou.(2017). Toll-like receptor 2 agonist exacerbates renal injury in diabetic mice. *Exp Ther Med.* Feb;13(2):495-502.
 48. Saurus P, Kuusela S, Lehtonen E, Hyvönen ME, Ristola M, Fogarty CL, et.al (2015).Podocyte apoptosis is prevented by blocking the Toll-like receptor pathway. *Cell Death Dis.* 2015 May 7;6(5):e1752. doi: 10.1038/cddis.2015.125.
 49. Yinghui Wang, Jiayi Liu, Qingqing Zhang, Weiwei Wang, Qingzhen Liu, et.al (2021).Human umbilical cord mesenchymal stem cells attenuate podocyte injury under high glucose via TLR2 and TLR4 signaling. *Diabetes Res Clin Pract.* Mar;173:108702. doi: 10.1016/j.diabres.2021.108702. Epub 2021 Feb 18.
 50. Luigi Franchi, Jong-Hwan Park, Michael H Shaw, Noemi Marina-Garcia et.al(2008).Intracellular NOD-like receptors in innate immunity, infection and disease. *Cell Microbiol.* 2008 Jan;10(1):1-8. doi: 10.1111/j.1462-5822.2007.01059.x.
 51. Meunier E, Broz P. (2017).Evolutionary Convergence and Divergence in NLR Function and Structure. *Trends Immunol.* 2017 Oct;38(10):744-757. doi: 10.1016/j.it.2017.04.005. Epub May 31.
 52. J, Zhou TJ, Ren GL, Cai L, Meng XM. (2022).Novel insights into NOD-like receptors in renal diseases. *JActa Pharmacol Sin.* Nov;43(11):2789-2806. doi: 10.1038/s41401-022-00886-7. Epub 2022 Apr 1.
 53. Ankit Malik, Thirumala-Devi Kanneganti.(2017). Inflammasome activation and assembly at a glance. *J Cell Sci.* Dec 1;130(23):3955-3963.
 54. Khurram Shahzad, Sameen Fatima, Hamzah Khawaja, Ahmed Elwakiel, Ihsan Gadi, Saira Ambreen, Silke Zimmermann, Peter R Mertens, Ronald Biemann, Berend Isermann.(2022). Podocyte-specific Nlrp3 inflammasome activation promotes diabetic kidney disease. *Kidney Int.* Oct;102(4):766-779. Epub 2022 Jun 30.
 55. Boini KM, Xia M, Abais JM, Li G, Pitzer AL, Gehr TW, Zhang Y, Li PL.(2014). Activation of inflammasomes in podocyte injury of mice on the high fat diet: Effects of ASC gene deletion and silencing. *Biochim Biophys Acta.* 2014 May;1843(5):836-

845. doi: 10.1016/j.bbamcr.2014.01.033. Epub Feb 5.
56. Christgen S, Kanneganti TD. (2019).Inflammasomes and the fine line between defense and disease. *Curr Opin Immunol.* 2020 Feb;62:39-44. doi: 10.1016/j.coi.2019.11.007. Epub Dec 25.
 57. Franz G Bauernfeind, Gabor Horvath, Andrea Stutz, Emad S Alnemri, Kelly MacDonald, David Speert,et.al(2009). Cutting edge: NF-kappaB activating pattern recognition and cytokine receptors license NLRP3 inflammasome activation by regulating NLRP3 expression. *J Immunol.* Jul 15;183(2):787-91. doi: 10.4049/jimmunol.0901363. Epub 2009 Jul 1.
 58. Coll RC, Schroder K, Pelegrín P. (2022).NLRP3 and pyroptosis blockers for treating inflammatory diseases.*Trends Pharmacol Sci.* Aug;43(8):653-668. doi: 10.1016/j.tips.2022.04.003. Epub 2022 May 3.
 59. Sokolova M, Sahraoui A, Høyem M, Ogaard J, Lien E, Aukrust P, Yndestad A, Ranheim T, Scholz H. (2018).NLRP3 inflammasome mediates oxidative stress-induced pancreatic islet dysfunction. *Am J Physiol Endocrinol Metab.* Nov 1;315(5):E912-E923.
 60. Yu JW, Lee MS.(2016). Mitochondria and the NLRP3 inflammasome: physiological and pathological relevance. *Arch Pharm Res.* Nov;39(11):1503-1518. doi: 10.1007/s12272-016-0827-4. Epub 2016 Sep 7.
 61. Heid ME, Keyel PA, Kamga C, Shiva S, Watkins SC, Salter RD. (2013).Mitochondrial reactive oxygen species induces NLRP3-dependent lysosomal damage and inflammasome activation. *J Immunol.* Nov 15;191(10):5230-8. doi: 10.4049/jimmunol.1301490. Epub 2013 Oct 2.
 62. Matthew S J Mangan, Edward J Olhava, William R Roush, H Martin Seidel, Gary D Glick, Eicke Latz. (2018).Targeting the NLRP3 inflammasome in inflammatory diseases. *Nat Rev Drug Discov.* Aug;17(8):588-606. doi: 10.1038/nrd.2018.97. Epub 2018 Jul 20.
 63. Hallie M Blevins, Yiming Xu, Savannah Biby, Shijun Zhang. (2022).The NLRP3 Inflammasome Pathway: A Review of Mechanisms and Inhibitors for the Treatment of Inflammatory Diseases. *Front Aging Neurosci.* Jun 10;14:879021.
 64. Pan Gao, Xian-Fang Meng, Hua Su, Fang-Fang He, Shan Chen, Hui Tang, Xiu-Juan Tian,et.al (2014).Thioredoxin-interacting protein mediates NALP3 inflammasome activation in podocytes during diabetic nephropathy. *Biochim Biophys Acta.* Nov;1843(11):2448-60.
 65. Gao P, He FF, Tang H, Lei CT, Chen S, Meng XF, Su H, Zhang C. (2015).NADPH oxidase-induced NALP3 inflammasome activation is driven by thioredoxin-interacting protein which contributes to podocyte injury in hyperglycemia. *J Diabetes Res.*;2015:504761. doi: 10.1155/2015/504761. Epub 2015 Mar 5.
 66. Khurram Shahzad, Fabian Bock, Wei Dong, Hongjie Wang, Stefan Kopf, et.al (2015), Thati Madhusudhan, Berend Isermann. Nlrp3-inflammasome activation in non-myeloid-derived cells aggravates diabetic nephropathy. *Kidney Int.* Jan;87(1):74-84. doi: 10.1038/ki.2014.271. Epub 2014 Jul 30.
 67. Ming Wu, Zhifen Yang, Chengyu Zhang, Yu Shi, Weixia Han, Shan Song, Lin Mu, Chunyang Du, Yonghong Shi.(2021). Inhibition of NLRP3 inflammasome ameliorates podocyte damage by suppressing lipid accumulation in diabetic nephropathy. *Metabolism.* May;118:154748
 68. Yun Hou, Sixiang Lin, Jun Qiu, Wangnan Sun, Menghua Dong, Yanxiao Xiang, Lin Wang, Pengchao Du. (2020).NLRP3 inflammasome negatively regulates podocyte autophagy in diabetic nephropathy. *Biochem Biophys Res Commun.* 2020 Jan 15;521(3):791-798.
 69. Sophie Carina Kunte, Julian A Marschner, Martin Klaus, Tâmisia Honda, Chenyu Li, et.al(2023), Hans-Joachim Anders. No NLRP3 inflammasome activity in kidney epithelial cells, not even when the NLRP3-A350V Muckle-Wells variant is expressed in podocytes of diabetic mice. *Front Immunol.* Aug 23;14:1230050. doi: 10.3389/fimmu.2023.1230050. eCollection 2023.
 70. Negroni A, Pierdomenico M, Cucchiara S, Stronati L.(2018). NOD2 and inflammation: current insights. *J Inflamm Res.* Feb 12;11:49-60. doi: 10.2147/JIR.S137606. eCollection 2018.
 71. Pengchao Du, Baoxia Fan, Huirong Han, Junhui Zhen, Jin Shang. (2013). NOD2 promotes renal injury by exacerbating inflammation and podocyte insulin resistance in diabetic nephropathy. *Kidney Int.* Aug;84(2):265-276.
 72. Jin Shang, Qiang Wan, Xiaojie Wang, Yiqi Duan, Ziyang Wang. (2015). Identification of NOD2 as a novel target of RNA-binding protein HuR: evidence from NADPH oxidase-mediated HuR signaling in diabetic nephropathy. *Free Radic Biol Med.* Feb: 79:217-227.
 73. Issa W, Njeim R, Carrazco A, Burke GW, Mitrofanova A. (2024). Role of the Innate Immune Response in Glomerular Disease Pathogenesis: Focus on Podocytes. *Cells.* Jul 6;13(13):1157.
 74. Chan KL, Tam TH, Boroumand P, Prescott D, Costford SR, Escalante NK. (2017). Circulating NOD1 Activators and Hematopoietic NOD1 Contribute to Metabolic Inflammation and Insulin Resistance. *Cell Rep.* Mar 7;18(10):2415-2426.
 75. Goubau D, Deddouche S, (2013). Reis e Sousa C. Cytosolic sensing of viruses. *Immunity.* May 23;38(5):855-869.
 76. Rehwinkel J, Gack MU. (2020). RIG-I-like receptors: their regulation and roles in RNA sensing. *Nat Rev Immunol.* Sep;20(9):537-551.
 77. Michifumi Yamashita, Carrie A Millward, Hiroyuki Inoshita, (2013). Paramananda Saikia, Saurabh Chattopadhyay, Ganes C Sen, Steven N Emancipator. Antiviral innate immunity disturbs podocyte cell function. *J Innate Immun.*;5(3):231-241.
 78. Walt E Adamson, Harry Noyes, Paul Johnson, Anneli Cooper. (2024). Phenome-wide analysis reveals epistatic associations between APOL1 variants and chronic kidney disease and multiple other disorders. *EBioMedicine.* Mar: 101:105000.
 79. Ji Fang, Xingmei Yao, Mingqiang Hou, Miao Duan. (2020). ApoL1 induces kidney inflammation through RIG-I/NF-κB activation. *Biochem Biophys Res Commun.* Jun 25;527(2):466-473.
 80. Cristian V Riella, Michelle McNulty, Guilherme T Ribas et.al. (2022). ADAR regulates APOL1 via A-to-I RNA editing by inhibition of MDA5 activation in a paradoxical biological circuit. *Proc Natl Acad Sci U S A.* Nov;119(44): e2210150119.
 81. Kawasaki T, Kawai T. (2019). Discrimination Between Self and Non-Self-Nucleic Acids by the Innate Immune System. *Int Rev Cell Mol Biol.*; 344:1-30.
 82. Lugrin J, Martinon F. (2018). The AIM2 inflammasome:

- Sensor of pathogens and cellular perturbations. *Immunol Rev*. Jan;281(1):99-114.
83. Yong Wu, Huan Yang, Sujuan Xu, Ming Cheng, Jie Gu, et al. (2022). AIM2 inflammasome contributes to aldosterone-induced renal injury via endoplasmic reticulum stress. *Clin Sci (Lond)*. Jan 14;136(1):103-120.
 84. Jinyu Pan, Li Han, Jun Guo, Xuyang Wang, Jingjing Tian, et al. (2018). AIM2 accelerates the atherosclerotic plaque progressions in ApoE^{-/-} mice. *Biochem Biophys Res Commun*. Apr 6;498(3):487-494.
 85. Jung Hwan Bae, Seung Ii Jo, Seong Jin Kim, Jong Min Lee, et al. (2019). Circulating Cell-Free mtDNA Contributes to AIM2 Inflammasome-Mediated Chronic Inflammation in Patients with Type 2 Diabetes. *Cells*. Apr 8;8(4):328.
 86. Zhenwei Gong, Xinyi Zhang, Kai Su, Ruihua Jiang, Zhe Sun, et al. (2019). Deficiency in AIM2 induces inflammation and adipogenesis in white adipose tissue leading to obesity and insulin resistance. *Diabetologia*. Dec;62(12):2325-2339.
 87. Elwira Pyz, Andrew S J Marshall, Siamon Gordon, Gordon D Brown. (2006). C-type lectin-like receptors on myeloid cells. *Ann Med* 38(4):242-251.
 88. Radaev S, Sun PD. (2003). Structure and function of natural killer cell surface receptors. *Annu Rev Biophys Biomol Struct*.;32:93-114.
 89. Alex N Zelensky, Jill E Gready. (2005). The C-type lectin-like domain superfamily. *FEBS J*. Dec;272(24):6179-6217.
 90. Hoving JC, Wilson GJ, Brown GD. (2014). Signalling C-type lectin receptors, microbial recognition and immunity. *Cell Microbiol*. Feb;16(2):185-194.
 91. Zeyu Su, Yujia Li, Hang Lv, Xiaoyang Cui, Min Liu, et al. (2021). CLEC14A protects against podocyte injury in mice with adriamycin nephropathy. *FASEB J*. Jul;35(7): e21711.
 92. Na Yang, Minxiu Wang, Ke Lin, Mengyang Wang, Diyun Xu, et al. (2023). Dectin-1 deficiency alleviates diabetic cardiomyopathy by attenuating macrophage-mediated inflammatory response. *Biochim Biophys Acta Mol Basis Dis*. Aug;1869(6):166710.
 93. Andrea Ablasser, Marion Goldeck, Taner Cavlar, Tobias Deimling, Gregor Witte, (2013). Ingo Röhl, Karl-Peter Hopfner, Janos Ludwig, Veit Hornung. cGAS produces a 2'-5'-linked cyclic dinucleotide second messenger that activates STING. *Nature*. Jun 20;498(7454):380-384.
 94. Hiroki Ishikawa, Zhe Ma, Glen N Barber. (2009). STING regulates intracellular DNA-mediated, type I interferon-dependent innate immunity. *Nature*. Oct 8;461(7265):788-792.
 95. Gao D, Li T, Li XD, Chen X, Li QZ, Wight-Carter M, et al. (2015). Activation of cyclic GMP-AMP synthase by self-DNA causes autoimmune diseases. *Proc Natl Acad Sci U S A*. Oct 20;112(42):E5699-705.
 96. Leticia Corrales, Laura Hix Glickman, Sarah M McWhirter, et al. (2015). Direct Activation of STING in the Tumor Microenvironment Leads to Potent and Systemic Tumor Regression and Immunity. *Cell Rep*. May 19;11(7):1018-1030.
 97. Motwani M, Pesiridis S, Fitzgerald KA. (2019). DNA sensing by the cGAS-STING pathway in health and disease. *Nat Rev Genet*. Nov;20(11):657-674.
 98. Nan Zang, Chen Cui, Xinghong Guo, Jia Song, Huiqing Hu, et al. (2022). cGAS-STING activation contributes to podocyte injury in diabetic kidney disease. *iScience*. Sep 16;25(10):105145.
 99. Alla Mitrofanova, Antonio Fontanella, Matthew Tolerico, et al. (2022). Activation of Stimulator of IFN Genes (STING) Causes Proteinuria and Contributes to Glomerular Diseases. *J Am Soc Nephrol*. Dec;33(12):2153-2173.
 100. Haiying Qi, Gabriella Casalena, Shaolin Shi, Liping Yu, et al. (2016). Glomerular Endothelial Mitochondrial Dysfunction Is Essential and Characteristic of Diabetic Kidney Disease Susceptibility. *Diabetes*. Mar;66(3):763-778.
 101. Gabriella A Casalena, Liping Yu, Roberto Gil, Samuel Rodriguez, et al. (2020). The diabetic microenvironment causes mitochondrial oxidative stress in glomerular endothelial cells and pathological crosstalk with podocytes. *Cell Commun Signal*. Jul 8;18(1):105.
 102. Ilse Daehn, Gabriella Casalena, Taoran Zhang, Shaolin Shi, et al. (2014). Endothelial mitochondrial oxidative stress determines podocyte depletion in segmental glomerulosclerosis. *J Clin Invest*. Apr;124(4):1608-1621.
 103. Barry I Freedman, Sophie Limou, Lijun Ma, Jeffrey B Kopp. (2018). APOL1-Associated Nephropathy: A Key Contributor to Racial Disparities in CKD. *Am J Kidney Dis*. Nov;72(5 Suppl 1): S8-S16.
 104. Tung CW, Hsu YC, Shih YH, Chang PJ, Lin CL. (2018). Glomerular mesangial cell and podocyte injuries in diabetic nephropathy. *Nephrology (Carlton)*. Oct;23 Suppl 4:32-37.
 105. Neeffes J, Jongsma ML, Paul P, Bakke O. (2011). Towards a system understanding of MHC class I and MHC class II antigen presentation. *Nat Rev Immunol*. Nov 11;11(12):823-836.
 106. Hathcock KS, Laszlo G, Pucillo C, Linsley P, Hodes RJ. (1994). Comparative analysis of B7-1 and B7-2 costimulatory ligands: expression and function. *J Exp Med*. Aug 1;180(2):631-640.
 107. Marcela Herrera, Magnus Söderberg, Alan Sabirsh, et al. (2017). Inhibition of T-cell activation by the CTLA4-Fc Abatacept is sufficient to ameliorate proteinuric kidney disease. *Am J Physiol Renal Physiol*. Apr 1;312(4): F748-F759.
 108. Li S, Liu Y, He Y, Rong W, Zhang M, Li L, et al. (2020). Podocytes present antigen to activate specific T cell immune responses in inflammatory renal disease. *J Pathol*. Oct;252(2):165-177.
 109. Liang Li, Wei Tang, Yan Zhang, Meng Jia, Limei Wang, et al. (2022). Targeting tissue-resident memory CD8⁺ T cells in the kidney is a potential therapeutic strategy to ameliorate podocyte injury and glomerulosclerosis. *Mol Ther*. Aug 3;30(8):2746-2759.
 110. Chen A, Lee K, D'Agati VD, Wei C, Fu J, (2018). Bowman's capsule provides a protective niche for podocytes from cytotoxic CD8⁺ T cells. *J Clin Invest*. Aug 1;128(8):3413-3424.
 111. Nakamichi R, Hishikawa A, Chikuma S, Yoshimura A, et al. (2023). DNA-damaged podocyte-CD8 T cell crosstalk exacerbates kidney injury by altering DNA methylation. *Cell Rep*. Apr 25;42(4):112302.
 112. *J Clin Epigenet*, (2021). Single Gene Inactivation with Implications to Diabetes and Multiple Organ Dysfunction Syndrome. *J Clin Epigenet*.; Vol. 3 No. 3:24.

113. J Clin Immunol Res. (2018). Appetite Control and Biotherapy in the Management of Autoimmune Induced Global Chronic Diseases. 2(1): 1-4.
114. Hong Q, Zhang L, Das B, Li Z, Liu B, Cai G, Chen X,et.al.(2018). Increased podocyte Sirtuin-1 function attenuates diabetic kidney injury. *Kidney Int. Jun;93(6):1330-1343.*
115. Tongtong Liu, Liping Yang, Huimin Mao, Fang Ma, Yuyang Wang,et.al. (2022). Sirtuins as novel pharmacological targets in podocyte injury and related glomerular diseases, *Biomedicine & Pharmacotherapy, Volume 155, 113620.*
116. Sirtuin 1, (2018). a Diagnostic Protein Marker and its Relevance to Chronic Disease and Therapeutic Drug Interventions". *EC Pharmacology and Toxicology 6.4: 209-215.*
117. Hiroyasu Yamamoto, Kristina Schoonjans, Johan Auwerx. (2007). Sirtuin functions in health and disease. *Mol Endocrinol. Aug;21(8):1745-1755.*
118. Yoshihisa Nakatani, Reiko Inagi. (2016). Epigenetic Regulation Through SIRT1 in Podocytes. *Curr Hypertens Rev.;12(2):89-94.*
119. Takayama K, Ishida K, Matsushita T, Fujita N, Hayashi S, Sasaki K,et.al.(2009). SIRT1 regulation of apoptosis of human chondrocytes. *Arthritis Rheum. Sep;60(9):2731-2740.*
120. Salminen A, Kaarniranta K. SIRT1: (2009). regulation of longevity via autophagy. *Cell Signal. Sep;21(9):1356-1360.*
121. Rodgers JT, Lerin C, Haas W, Gygi SP, Spiegelman BM, Puigserver P. (2005). Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. *Nature. Mar 3;434(7029):113-118.*
122. Scarpulla RC. (2011). Metabolic control of mitochondrial biogenesis through the PGC-1 family regulatory network. *Biochim Biophys Acta. Jul;1813(7):1269-1278.*
123. Fahmida Rasha, Brianyell McDaniel Mims, Isabel Castro-Piedras,et.al(2020). The Versatility of Sirtuin-1 in Endocrinology and Immunology. *Front Cell Dev Biol. Nov 19;8:589016.*
124. Vera P Mourits, Leonie S Helder, Vasiliki Matzaraki, Valerie A C M Koeken,et.al.(2009). The role of sirtuin 1 on the induction of trained immunity. *Cell Immunol. 2021 Aug;366: 104393.*
125. Xi Chen, Yun Lu, Zhengguo Zhang, Jian Wang, Hui Yang,et.al.(2015). Intercellular interplay between Sirt1 signalling and cell metabolism in immune cell biology. *Immunology. Aug;145(4):455-467.*
126. Hong Q, Zhang L, Das B, Li Z, Liu B, Cai G,et.al.(2018). Increased podocyte Sirtuin-1 function attenuates diabetic kidney injury. *Kidney Int. Jun;93(6):1330-1343.*
127. Mingzhu Jiang, Min Zhao, Mi Bai, Juan Lei, Yanggang Yuan,et.al.(2021). SIRT1 Alleviates Aldosterone-Induced Podocyte Injury by Suppressing Mitochondrial Dysfunction and NLRP3 Inflammasome Activation. *Kidney Dis (Basel). Jul;7(4):293-305.*
128. Shuta Motonishi, Masaomi Nangaku, Takehiko Wada, Yu Ishimoto, Takamoto Ohse, Taiji Matsusaka, Naoto Kubota, Akira Shimizu, Takashi Kadowaki, Kazuyuki Tobe, Reiko Inagi. (2015). Sirtuin1 Maintains Actin Cytoskeleton by Deacetylation of Cortactin in Injured Podocytes. *J Am Soc Nephrol. Aug;26(8):1939-1959.*
129. Dorota Rogacka. (2021). Insulin resistance in glomerular podocytes: Potential mechanisms of induction. *Arch Biochem Biophys. Oct 15;710:109005.*
130. American Diabetes Association Professional Practice Committee. 2. Diagnosis and Classification of Diabetes: Standards of Care in Diabetes-2024. *Diabetes Care. 1;47(Suppl 1): S20-S42.*
131. Vallon V. (2020). Glucose transporters in the kidney in health and disease. *Pflugers Arch. Sep;472(9):1345-1370.*
132. Volker Vallon, Kenneth A Platt, Robyn Cunard, Jana Schroth,et.al.(2011). SGLT2 mediates glucose reabsorption in the early proximal tubule. *J Am Soc Nephrol. Jan;22(1):104-112.*
133. Yaribeygi H, Butler AE, Atkin SL, Katsiki N, Sahebkar A. (2018). Sodium-glucose cotransporter 2 inhibitors and inflammation in chronic kidney disease: Possible molecular pathways. *J Cell Physiol. Jan;234(1):223-230.*
134. Ravindran S, Munusamy S. (2022). Renoprotective mechanisms of sodium-glucose co-transporter 2 (SGLT2) inhibitors against the progression of diabetic kidney disease. *J Cell Physiol. 2022 Feb;237(2):1182-1205.*
135. Kelly L Hudkins, Xianwu Li, Alexander L Holland, Shreya Swaminathan, Charles E Alpers. (2022). Regression of diabetic nephropathy by treatment with empagliflozin in BTBR ob/ob mice. *Nephrol Dial Transplant. Apr 25;37(5):847-859.*
136. Paola Cassis, Monica Locatelli, Domenico Cerullo, Daniela Corna, Simona Buelli,et.al(2018). SGLT2 inhibitor dapagliflozin limits podocyte damage in proteinuric nondiabetic nephropathy. *JCI Insight. Aug 9;3(15): e98720.*
137. Anton I Korbut, Iuliia S Taskaeva, Nataliya P Bgatova, Natalia A Muraleva,et.al. (2020). SGLT2 Inhibitor Empagliflozin and DPP4 Inhibitor Linagliptin Reactivate Glomerular Autophagy in db/db Mice, a Model of Type 2 Diabetes. *Int J Mol Sci. Apr 23;21(8):2987.*
138. Lei Yang, Baozhu Liang, Jingxin Li, Xiaoyan Zhang, Hong Chen,et.al.(2018). Dapagliflozin alleviates advanced glycation end product induced podocyte injury through AMPK/mTOR mediated autophagy pathway. *Cell Signal. Feb: 90:110206.*
139. Ji Fang, Xingmei Yao, Mingqiang Hou, Miao Duan, Lina Xing, et.al.(2020). Yunman Wang, Bingbing Zhu, Qijiang Chen, Hao Wang. ApoL1 induces kidney inflammation through RIG-I/NF- κ B activation. *Biochem Biophys Res Commun. Jun 25;527(2):466-473.*
140. Tao Jiang, Scott E Liebman, M Scott Lucia, Jinping Li, Moshe Levi. (2005). Role of altered renal lipid metabolism and the sterol regulatory element binding proteins in the pathogenesis of age-related renal disease. *Kidney Int. Dec;68(6):2608-2620.*
141. Mengyuan Ge, Judith Molina, Jin-Ju Kim, Shamroop K Mallela,et.al.(2023). Empagliflozin reduces podocyte lipotoxicity in experimental Alport syndrome. *Elife. May 2:12: e83353.*
142. Dong Wang, Yuhuan Luo, Xiaoxin Wang, David J Orlicky,et.al,(2018). Moshe Levi. The Sodium-Glucose Cotransporter 2 Inhibitor Dapagliflozin Prevents Renal and Liver Disease in Western Diet Induced Obesity Mice. *Int J Mol Sci. Jan 3;19(1):137.*
143. Pérez-Morales RE, Del Pino MD, Valdivielso JM, Ortiz A, Mora-Fernández C, Navarro-González JF. Inflammation in

- Diabetic Kidney Disease. *Nephron*. 2019;143(1):12-16.
144. Shadia A Fathy, Mohamed R Mohamed, Mohamed A M Ali, Ashraf E El-Helaly, Abdulnabi T Alattar. (2019). Influence of IL-6, IL-10, IFN- γ and TNF- α genetic variants on susceptibility to diabetic kidney disease in type 2 diabetes mellitus patients. *Biomarkers*. Feb;24(1):43-55.
 145. So Ra Kim, Sang-Guk Lee, Soo Hyun Kim, Jin Hee Kim, Eunhye Choi,et.al.(2020). SGLT2 inhibition modulates NLRP3 inflammasome activity via ketones and insulin in diabetes with cardiovascular disease. *Nat Commun*. May 1;11(1):2127.
 146. Hidjo J L Heerspink, Paul Perco, Skander Mulder, Johannes Leierer, Michael K Hansen, Andreas Heinzl, Gert Mayer. (2019). Canagliflozin reduces inflammation and fibrosis biomarkers: a potential mechanism of action for beneficial effects of SGLT2 inhibitors in diabetic kidney disease. *Diabetologia*. Jul;62(7):1154-1166
 147. Yong-Ping Lu, Hong-Wei Wu, Ting Zhu, Xi-Tong Li, Jiao Zuo,et.al.(2019). Empagliflozin reduces kidney fibrosis and improves kidney function by alternative macrophage activation in rats with 5/6-nephrectomy. *Biomed Pharmacother*. Dec: 156:113947.
 148. Drucker DJ. (2018). Mechanisms of Action and Therapeutic Application of Glucagon-like Peptide-1. *Cell Metab*. Apr 3;27(4):740-756.
 149. Nauck MA, Quast DR, Wefers J, Meier JJ. (2021). GLP-1 receptor agonists in the treatment of type 2 diabetes - state-of-the-art. *Mol Metab*. Apr; 46:101102.
 150. von Scholten BJ, Hansen TW, Goetze JP, Persson F, Rossing P. (2015). Glucagon-like peptide 1 receptor agonist (GLP-1 RA): long-term effect on kidney function in patients with type 2 diabetes. *J Diabetes Complications*. Jul;29(5): 670-674.
 151. B J von Scholten, M Lajer, J P Goetze, F Persson, P Rossing. (2015). Time course and mechanisms of the anti-hypertensive and renal effects of liraglutide treatment. *Clinical Trial Diabet Med*. Mar;32(3):343-352.
 152. Marco Zavattaro, Marina Caputo, Maria Teresa Samà, Chiara Mele, Luisa Chasseur, Paolo Marzullo,et.al. (2015). One-year treatment with liraglutide improved renal function in patients with type 2 diabetes: a pilot prospective study. *Endocrine*. Dec;50(3):620-625.
 153. Chaochen Wang, Hiroshi Yatsuya, Koji Tamakoshi, Mayu Uemura, Yuanying Li,et.al. (2013). Positive association between high-sensitivity C-reactive protein and incidence of type 2 diabetes mellitus in Japanese workers: 6-year follow-up. *Diabetes Metab Res Rev*. Jul;29(5):398-405.
 154. Mohsen Mazidi, Ehsan Karimi, Peyman Rezaie, Gordon A Ferns. (2017). Treatment with GLP1 receptor agonists reduce serum CRP concentrations in patients with type 2 diabetes mellitus: A systematic review and meta-analysis of randomized controlled trials. *J Diabetes Complications*. Jul;31(7):1237-1242.
 155. Bendotti G, Montefusco L, Lunati ME, Usuelli V, Pastore I, Lazzaroni E, Assi E, Seelam AJ, El Essawy B, Jang J, Loretelli C, D'Addio F, Berra C, Ben Nasr M, Zuccotti G, Fiorina P. The anti-inflammatory and immunological properties of GLP-1 Receptor Agonists. *Pharmacol Res*. 2022 Aug; 182:106320.
 157. Jiali Wang, Yanni Zhou, Dan Long, Yucheng Wu, Fang Liu. (2023). GLP-1 receptor agonist, liraglutide, protects podocytes from apoptosis in diabetic nephropathy by promoting white fat browning. *Biochem Biophys Res Commun*. Jul 5:664:142-151.
 158. Xiang Li, Xiao Jiang, Mei Jiang, Zhi-Feng Wang, Tao Zhao,et.al. (2024). GLP-1RAs inhibit the activation of the NLRP3 inflammasome signaling pathway to regulate mouse renal podocyte pyroptosis. *Acta Diabetol*. Feb;61(2):225-234.
 159. Yongjian Ye, Xing Zhong, Na Li, Tianrong Pan. (2019). Protective effects of liraglutide on glomerular podocytes in obese mice by inhibiting the inflammatory factor TNF- α -mediated NF- κ B and MAPK pathway. *Obes Res Clin Pract*. Jul-Aug;13(4):385-390.
 160. Karly C Sourris, Yi Ding, Scott S Maxwell, Annas Al-Sharea,et.al. (2024). Glucagon-like peptide-1 receptor signaling modifies the extent of diabetic kidney disease through dampening the receptor for advanced glycation end products-induced inflammation. *Kidney Int*. Jan;105(1):132-149.
 161. Susana Ravassa, Javier Beaumont, Ana Huerta, Joaquín Barba, Isabel Coma-Canella,et.al.(2015).association of low GLP-1 with oxidative stress is related to cardiac disease and outcome in patients with type 2 diabetes mellitus: a pilot study. *Free Radic Biol Med*. Apr: 81:1-12.
 162. Assaad A Eid, Yves Gorin, Bridget M Fagg, Rita Maalouf, Hanna E Abboud,et.al.(2009). Mechanisms of podocyte injury in diabetes: role of cytochrome P450 and NADPH oxidases. *Diabetes*. May;58(5):1201-1211.
 163. Daria V Ilatovskaya, Gregory Blass, Oleg Palygin, Vladislav Levchenko,et.al.(2018). A NOX4/TRPC6 Pathway in Podocyte Calcium Regulation and Renal Damage in Diabetic Kidney Disease. *J Am Soc Nephrol*. Jul;29(7):1917-1927.
 164. Natalie Youssef, Mohamed Nourdein, Rachel Njeim, Hilda E Ghadieh,et.al.(2021). Reno-Protective Effect of GLP-1 Receptor Agonists in Type1 Diabetes: Dual Action on TRPC6 and NADPH Oxidases. *Biomedicines*. Sep 30;9(10):1360.
 165. Jie Liu, Shanshan Guo, Hui Li, Xu-Ying Liu. Effects of glucagon-like peptide-1 receptor agonists (GLP-1RAs) on podocytes, inflammation, and oxidative stress in patients with diabetic nephropathy (DN). *Pak J Med Sci*. 2022 May-Jun;38(5):1170-1174. doi: 10.12669/pjms.38.5.4719.
 166. Soraya Puglisi, Alessandro Rossini, Roberta Poli, Francesca Dughera,et.al.(2021). Effects of SGLT2 Inhibitors and GLP-1 Receptor Agonists on Renin-Angiotensin-Aldosterone System. *Front Endocrinol (Lausanne)*. Oct 21:12:738848.
 167. Minsuk Kim, Mathew J Platt, Tadao Shibasaki, Susan E Quaggin, Peter H Backx,et.al.(2013). GLP-1 receptor activation and Epac2 link atrial natriuretic peptide secretion to control of blood pressure. *Nat Med*. May;19(5):567-575.
 168. Ames MK, Atkins CE, Pitt B. (2019). The renin-angiotensin-aldosterone system and its suppression. *J Vet Intern Med*. Mar;33(2):363-382.
 169. Christiane Rüster, Gunter Wolf. (2006). Renin-angiotensin-aldosterone system and progression of renal disease. *J Am Soc Nephrol*. Nov;17(11):2985-2991.
 170. Lewis EJ, Hunsicker LG, Clarke WR, Berl T, Pohl MA, Lewis JB, Ritz E, Atkins RC, Rohde R, Raz I;Collaborative Study Group.Renoprotective effect of the angiotensin-receptor antagonist irbesartan in patients with nephropathy due to type 2 diabetes. *N Engl J Med*. 2001 Sep 20;345(12):851-860.

171. Mauer M, Zinman B, Gardiner R, Suissa S, Sinaiko A, Strand T, et al. (2009). Renal and retinal effects of enalapril and losartan in type 1 diabetes. *N Engl J Med.* Jul 2;361(1):40-51.
172. Max C Liebau, D Lang, J Böhm, N Endlich, Martin J Bek, Ian Witherden, et al. (2006). Functional expression of the renin-angiotensin system in human podocytes. *Am J Physiol Renal Physiol.* Mar;290(3): F710-9.
173. Guohua Ding, Krishna Reddy, Aditi A Kapasi, Nicholas Franki, Nora Gibbons, Balakuntalam S Kasinath, Pravin C Singhal. Angiotensin II induces apoptosis in rat glomerular epithelial cells. *Am J Physiol Renal Physiol.* 2002 Jul;283(1): F173-80.
174. Xuejing Zhu, Dan Gao, Vittorio Albertazzi, Jianyong Zhong, Li-Jun Ma, Liping Du Yu Shyr, Valentina Kon, Hai-Chun Yang, Agnes B Fogo. Podocyte-Related Mechanisms Underlying Survival Benefit of Long-Term Angiotensin Receptor Blocker. *Int J Mol Sci.* 2022 May 27;23(11):6018.
175. Kelly L Hudkins, Tomasz A Wietecha, Floor Steegh, Charles E Alpers. (2020). Beneficial effect on podocyte number in experimental diabetic nephropathy resulting from combined atrasentan and RAAS inhibition therapy. *Am J Physiol Renal Physiol.* May 1;318(5):F1295-F1305.
176. Paola Rizzo, Rubina Novelli, Ariela Benigni, Giuseppe Remuzzi. (2016). Inhibiting angiotensin-converting enzyme promotes renal repair by modulating progenitor cell activation. *Pharmacol Res.* 2016 Jun: 108:16-22.
177. Gomez-Sanchez E, Gomez-Sanchez CE (2014). The multifaceted mineralocorticoid receptor. *Compr Physiol.* Jul;4(3):965-994.
178. Akira Nishiyama. (2019). Pathophysiological mechanisms of mineralocorticoid receptor-dependent cardiovascular and chronic kidney disease. *Hypertens Res.* Mar;42(3):293-300.
179. Bertocchio JP, Warnock DG, Jaisser F. (2011). Mineralocorticoid receptor activation and blockade: an emerging paradigm in chronic kidney disease. *Kidney Int.* May;79(10):1051-1060.
180. Zannad F, Gattis Stough W, Rossignol P, Bauersachs J, McMurray JJ, Swedberg K, Struthers AD, Voors AA, Ruilope LM, Bakris GL, O'Connor CM, Gheorghiadu M, Mentz RJ, Cohen-Solal A, Maggioni AP, Beygui F, Filippatos GS, Massy ZA, Pathak A, Piña IL, Sabbah HN, Sica DA, Tavazzi L, Pitt B. Mineralocorticoid receptor antagonists for heart failure with reduced ejection fraction: integrating evidence into clinical practice. *Eur Heart J.* 2012 Nov;33(22):2782-2795.
181. Bakris GL, Agarwal R, Chan JC, Cooper ME, Gansevoort RT, Haller H, et al. (2011). Mineralocorticoid Receptor Antagonist Tolerability Study—Diabetic Nephropathy (ARTS-DN) Study Group. Effect of Finerenone on Albuminuria in Patients With Diabetic Nephropathy: A Randomized Clinical Trial. *JAMA.* 2015 Sep 1;314(9):884-894.
182. Rico-Mesa JS, White A, Ahmadian-Tehrani A, Anderson AS. (2020). Mineralocorticoid Receptor Antagonists: A Comprehensive Review of Finerenone. *Curr Cardiol Rep.* Sep 10;22(11):140.
183. American Diabetes Association Professional Practice Committee. (2024). 11. chronic kidney disease and Risk Management: Standards of Care in Diabetes-2024. *Diabetes Care.* Jan 1;47(Suppl 1): S219-S230.
184. Lawrence Blonde, Guillermo E Umpierrez, S Sethu Reddy, Janet B McGill. American Association of Clinical Endocrinology Clinical Practice Guideline: Developing a Diabetes Mellitus Comprehensive Care Plan-2022 Update. *Endocr Pract.* 2022 Oct;28(10):923-1049. doi: 10.1016/j.eprac.2022.08.002. Epub 2022 Aug 11.
185. Shibata S, Nagase M, Yoshida S, Kawachi H, Fujita T. (2007). Podocyte as the target for aldosterone: roles of oxidative stress and Sgk1. *Hypertension.* Feb;49(2):355-364.
186. Daigoro Hirohama, Mitsuhiro Nishimoto, Nobuhiro Ayuzawa, Wakako Kawarazaki, et al. (2021). Activation of Rac1-Mineralocorticoid Receptor Pathway Contributes to Renal Injury in Salt-Loaded db/db Mice. *Hypertension.* Jul;78(1):82-93.
187. Salvadori M, Tsalouchos A. (2022). How immunosuppressive drugs may directly target podocytes in glomerular diseases. *Pediatr Nephrol.* Jul;37(7):1431-1441.
188. Mallipattu SK, He JC. (2016). The podocyte as a direct target for treatment of glomerular disease? *Am J Physiol Renal Physiol.* Jul 1;311(1): F46-51.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article Click Here: **Submit Manuscript**

DOI:[10.31579/2690-1919/399](https://doi.org/10.31579/2690-1919/399)

Ready to submit your research? Choose Auctores and benefit from:

- fast, convenient online submission
- rigorous peer review by experienced research in your field
- rapid publication on acceptance
- authors retain copyrights
- unique DOI for all articles
- immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <https://www.auctoresonline.org/journals/journal-of-clinical-research-and-reports>