

Doum Palm Meal (Hyphaene Thebaica) and Partial Maize Substitution: Impact on The Blood Biochemical Indicators of Weaned Pigs

Alagbe Olujimi John

Sumitra Research Institute, Gujarat, India; Department of Animal Nutrition and Biochemistry.

*Corresponding author: Alagbe Olujimi John, Sumitra Research Institute, Gujarat, India; Department of Animal Nutrition and Biochemistry.

Received date: February 02, 2024; Accepted date: February 23, 2024; Published date: March 04, 2024

Citation: Alagbe O. John, (2024), Doum Palm Meal (Hyphaene Thebaica) and Partial Maize Substitution: Impact on The Blood Biochemical Indicators of Weaned Pigs, *J. Nutrition and Food Processing*, 7(3); DOI:10.31579/2637-8914/190

Copyright: © 2024, Alagbe Olujimi John. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract:

This study looked at how weaned pigs' blood biochemical indicators were affected when doum palm meal (DPM) was used in part lieu of maize. After being weaned at 28 days of age and weighing 7.40 ± 0.09 kg at birth, fifty Landrace \times Duroc crossbred pigs were divided into divisions based on their body weights and given five distinct experimental diets. Over the course of a ninety-day fully randomized design experiment. Diets 1, 2, 3, 4, and 5 were fed 0 %, 5 %, 10 %, 15 %, and 20 % DPM in place of maize. The NRC (2012) states that the experimental diet satisfied the pigs' needs, and clean water was provided daily and at will. DPM had the following elements: manganese (0.91 mg/100g), zinc (0.72 mg/100g), copper (0.35 mg/100g), iron (5.60 mg/100g), calcium (371.20 mg/100g), potassium (966.31 mg/100g), phosphorus (206.19 mg/100g), magnesium (150.67 mg/100g), and zinc. The values of total serum protein, albumin, globulin, creatinine, alanine phosphatase, aspartate transaminase, sodium, potassium, calcium, phosphorus, and bicarbonate were not affected ($P > 0.05$) by the treatments, with the exception of the glucose level, which was higher ($P < 0.05$) in D1 than in the other treatments. It has been shown that DPM can partially replace up to 20% of maize without causing metabolic problems or adversely influencing the health of the animals.

Key words: swine; doum palm; maize; serum; minerals; phytochemicals

Introduction

The high cost of other high-energy concentrate feed and the scarcity of grain make swine feed expensive in underdeveloped nations [1]. [2] states that 70–80 % of the overall cost of producing livestock is spent on feed, with a significant portion of this cost being attributable to the high price of maize on the market [3]. A common staple grain in many parts of the world, including Asia, Latin America, and Africa, is maize (*Zea mays* L.). A common feed grain and energy source for cattle, maize is fed to them as part of their diets [3]. When estimating the nutritional worth of other grains, especially their energy content, maize is usually compared to them [4, 5].

Maize is a common cereal grain in Nigeria that is fed to chickens and consumed by humans alike. However, given its increased demand for various processing industries, maize availability in Nigeria both now and in the future is in doubt. The use of cereal grains, especially maize, as a source of starch industry appears to justify the ongoing price increases of standard chicken feeds [6,7]. Given the circumstances, it is necessary to assess additional locally accessible non-conventional feed sources and include the most promising ones into the diets of chickens.

One potential replacement is the doum palm fruit (*Hyphaene thebaica*), which is abundant in essential minerals like potassium, salt, calcium, magnesium, phosphorus, and other nutrients [8]. The plant belongs to the Arecaceae family and is endemic to Egypt, sub-Saharan Africa, and west India, especially Gujarat [9]. The tree is dichotomous and arborescent in nature, and it has been called one of the world's useful plants [10]. Although the leafy stem is used for construction, the foliage is used to make hats, mats, ropes, and baskets [11]. The doum palm is used to treat bilharziasis and its fruit is sometimes chewed to decrease high blood pressure [12, 13]. [14] reported that the doum palm fruit pulp's proximate composition included the following: protein (2.86%), fat (0.92%), ash (6.24%), crude fiber (12.87%), moisture (8.64%), and carbohydrates (68.47%). Similar findings were made by [9] who recorded a crude protein of 2.92 percentage, ether extract (0.49 percentage), crude fiber (15.14 percentage), and metabolizable energy (2254.5 kcal/kg) for DPM. A wealth of information exists regarding the impact of partially substituting doum palm with maize on the serum biochemical indices of weaned piglets, despite the fact that there are numerous papers on the use of doum palm fruit in poultry. Timely assessment is essential to minimize feed costs, boost animal protein, and optimize the use of the test item.

Determining the impact of doum palm meal on the serum biochemical indicators of the weaned piglets was the aim of this investigation.

Methodology

Site of investigation and ethical criteria

The animals division of Sumitra Research Institute, situated in Gujarat, India at 23° 13' N and 72° 41' E, is where the experiment was carried out from March to June, 2023. Every part of management was carried out with the institute's ethical standard (ASF/08A/2023).

Acquiring and preparing dry doum palm meal (DPM)

The doum palm's ripe fruits were gathered from Gujarat's Sumitra Research Farm and arranged in tidy, level iron trays. It was identified and sent to the same institute's Department of Crop Production. To separate the outer part from the kernel, the dried fruits were separated in a mortar and pestle. Each sample was then collected into a separate, sterile plastic bucket. After being further ground into tiny particles in a hammer mill, the mesocarp was kept in an airtight container with a clearly marked label. Before samples were taken to the lab for chemical analysis.

Animal treatment, feed, and design of experiments

At 28 days, 50 Landrace × Duroc piglets, weighing an average of 7.40 ± 0.09 kg at birth, were weaned and relocated to sterile nursery cages from Sumitra Teaching and Research Farm. During a 14-day quarantine, the animals received treatment with Oxytrox® (long-acting antibiotics) at the recommended dosage as well as injections of Ivermectin®, which fights ecto and endo parasites. After the acclimation phase, the pigs were divided into five treatments of ten pigs each, all of whom were housed individually in 1.7-square-meter pens with open sides measuring 10 m × 5-m × 5-m. The pigs were grouped according to their body weight. The pens that each held an animal measured. Five experimental diets were created using different amounts of Doum palm meal (DPM) to substitute

maize on a dry matter basis (Table 2). Diet 1 (D1) included no DPM, while diet 2 (5%), diet 3 (10%), diet 4 (15%), and diet 5 (20%) DPM levels. Diets were developed based on the National Research Council's [15] guidelines for pig needs.

Three feeding times a day were arranged: at 7:00, 12:00, and 17:00. Daily, limitless access to clean water was provided. The weight of the feed supplied the day before was subtracted from the weight of the leftover feed to establish the amount consumed.

Chemical evaluation

The Lionat® near-infrared feed analyzer (Model: CF-007HSD, China) was used to perform a chemical evaluation on DPM and trial diets. After inserting 200 grams of each sample into the sample cap or entry channel, thereafter the cap is placed on the metallic lid of the machine's tray, pressing the scan button allows you to examine the results just in time (1 minutes).

Analysis of minerals was carried out using Labocon atomic absorption spectrophotometer LAAS - 100 series which offer superior baseline stability of 0.002 A/30 minutes with advanced optical system to ensure fast and reliable results. The machine has the following general specification; resolution (< 30 %), baseline stability (0.005 A/30 minutes).

Phytochemical analysis of doum palm meal was evaluated adopting standard laboratory procedures outlined by [23].

Statistical analysis used in the experiment

Using the Statistical Analysis System Software (SAS), all collected on serum biochemical parameters underwent a one-way analysis of variance. The SAS Turkey test was used to separate the means, and significant differences were identified at P<0.05.

Feedstuffs	D1 (0 %)	D2 (5 %)	D3 (10 %)	D4 (15 %)	D5 (20 %)
Yellow maize	53.00	48.00	43.00	38.00	33.00
doum palm meal (DPM)	0.00	5.00	10.00	15.00	20.00
Rice bran	10.00	10.00	10.00	10.00	10.00
S/cake	14.00	14.00	14.00	14.00	14.00
GNM	17.50	17.50	17.50	17.50	17.50
B/meal	3.00	3.00	3.00	3.00	3.00
O/shell	1.55	1.55	1.55	1.55	1.55
Lysine	0.20	0.20	0.20	0.20	0.20
Methionine	0.30	0.30	0.30	0.30	0.30
Premix	0.25	0.25	0.25	0.25	0.25
Salt	0.35	0.35	0.35	0.35	0.35
Total	100.00	100.00	100.00	100.00	100.00
Determined analysis					
Crude protein (%)	18.77	18.67	18.30	18.25	18.20
Energy (Kcal/kg)	2788.5	2700.0	2695.4	2690.0	268.0

Mineral/Vitamin premix supplied per kg diet: - vit A, 8,500 I.U; vit E, 10.91 mg; vit D3, 2500I.U, vit K, 3.2mg; vit B2, 5.0mg; Niacin, 40 mg; vit B12, 25 mg; choline chloride, 100 mg; Mn, 5.0 mg; Zn, 35.1mg; Cu, 2.0g; folic acid, 2.5mg; Fe, 5.8g; pantothenic acid, 10mg; biotin, 30.5g; antioxidant, 56mg; S/cake: soya cake; GNM: groundnut meal; B/meal: bone meal; O/shell: Oyster shell

Table 1(T1): Diet composition chemical analysis

Results and Discussion

Chemical evaluation of DPM

In Table 2, the chemical evaluation of DPM reveals that ether extract and crude protein values of 5.35 % and 1.90 % were less than the 6.09 % and

1.75 % reported by [16]. Because the sample's crude protein concentration is low (less than 30 %) and it cannot be used to replace protein sources in the diet of monogastric according to [15]. The study's 13.51% crude fiber content was more than the 12.87 % reported by [14]. The experimentally reported ash content is consistent with the results of [17], whereas the reported energy level is consistent with those of [18].

Constituents	Composition (percentage)
C/ protein	5.35
C/ fibre	13.51
E/extract	1.10
Ash content	7.83
Metabolizable energy (kcal/kilogram)	2610.7

C/protein: crude protein; C/fibre: crude fibre; E/extract: ether extract

Table 2: Chemical evaluation of DPM

Copper had the lowest concentration (0.35 mg/100g) and potassium had the highest concentration (966.31 mg/100g) in the mineral composition of doum palm meal (Table 3). Calcium, potassium, phosphorus, sodium, magnesium, iron, manganese, zinc, and copper are listed in order of abundance. The results of this experiment showed greater values for

calcium (371.20 mg/100g), phosphorus (206.19 mg/100g), and iron (5.60 mg/100g) than those reported by [19] (336.00 mg/100g, 200.8 mg/100g, and 4.86 mg/100g). The potassium (966.31) value for fresh doum meal was less than that of the reports made by [18]. These differences might result from the test ingredients' species or processing technique [20, 21].

Constituents	Concentrations (mg/100g)
Calcium	371.20
Potassium	966.31
Phosphorus	206.19
Magnesium	150.67
Manganese	0.91
Zinc	0.72
Copper	0.35
Sodium	177.52
Iron	5.60

Table 3: Mineral composition of doum palm meal

Phytochemical composition of doum palm meal is presented in Table 4. Flavonoids had the highest concentration (2705.1 mg/100g) while saponins had the lowest concentration (0.66 mg/100g). These bioactive compounds have therapeutic properties and their concentration in the plant tissues is considered as the main factor to evaluate the therapeutic value and quality of a given herb [22]. For instance, alkaloids are believed to function as defensive elements against predators, especially mammals because of their general toxicity and deterrence capability [23]. Phenolic compounds have antioxidant, anti-inflammatory, anti-carcinogenic and other biological properties and may prevent oxidative stress [24].

Table 4 displays the phytochemical composition of doum palm meal. Flavonoids had the highest concentration (2705.1 mg/100g), whereas saponins had the lowest content (0.66 mg/100g). These bioactive

compounds have therapeutic properties, and it is believed that a herb's concentration in the plant tissues is the main factor determining its quality and therapeutic efficacy [22]. For instance, alkaloids are believed to function as defensive elements against predators, especially mammals, due to their general toxicity and deterring impact [23]. Per [24], phenolic compounds have biological properties that include anti-inflammatory, anti-carcinogenic, antioxidant, and they may help prevent oxidative stress. Pharmacological evidence has revealed the hepatoprotective, antioxidant, and anti-inflammatory effects of terpenoids and flavonoids [25]. Tannins also have antibacterial, antifungal, and immune-stimulating qualities. According to [26], saponins are used as adjuvants in vaccines against the herpes simplex virus (HSV), influenza, and HIV.

Parameters	Concentration (mg/100g)
Alkaloid	1218.3
Flavonoids	2705.1
Saponins	0.66
Tannins	175.8
Terpenoids	2.64
Phenols	561.1

Table 4: Phytochemical components of doum palm meal

Serum biochemical indices of weaned pigs fed DPM

The dietary treatments did not influence ($P>0.05$) total protein, albumin/globulin ratio, creatinine, alanine transaminase and alkaline phosphatase except glucose levels ($P<0.05$) in Table 5. However, all values were within the normal levels for disease free pigs by [27]. Measurements of total protein can reveal nutritional status related to liver or renal disease as well as other medical disorders [28]. Many hydrophobic compounds in the blood, including metals, non-esterified fatty acids, steroids, and thyroxine, are carried by albumin [29]. According to [30], the pigs' normal range of globulin levels indicates that the doum palm meal did not cause any serious health problems. Nutritional and immunological function can be understood by the

albumin/globulin ratio (A/G ratio) [31]. According to [32] it is also an index for screening chronic disorders, including kidney diseases. Diarrhea or extreme dehydration may be the cause of a high A/G ratio [33]. Glycine, arginine, and methionine are the amino acids that are transaminated in the liver, pancreas, and kidneys to produce creatinine [34]. It can also be used as a renal function indicator and is made from creatine and phosphocreatine [35]. T1 had higher ($P<0.05$) glucose levels than the other groups. According to [36], the glucose readings in this trial (100–121.64 mmol/L) fell within the normal range. Stress, inadequate diet, and poor management are all linked to elevated glucose levels (Clack and Coffer, 2008). The study's reported ranges for aspartate transaminase (AST) and alanine phosphatase (ALP) are within the optimal range [37] at 55.79 - 56.50 IU/L and 143.09 - 148.40 IU/L, respectively. According

to [38], low levels of ALP may indicate malnourishment, magnesium and zinc deficiencies, whereas high levels may be indicative of bacterial

infections and liver blockage in animals [39]. Elevated AST levels can be a sign of pancreatitis, cirrhosis, and cardiovascular disorders [40].

Components	D1	D2	D3	D4	D5	SEM	Ref. value
TPP (g/dL)	4.78	4.75	4.73	4.71	4.70	0.13	4.00 - 5.80
ALB (g/dL)	3.15	3.13	3.12	3.11	3.10	0.10	3.10 - 4.80
GLB (g/dL)	1.63	1.62	1.61	1.60	1.60	0.02	0.30 - 1.70
A/G ratio (g/dL)	1.93	1.93	1.94	1.94	1.94	0.01	0.90 - 2.00
Creatinine (mmol/L)	0.95	0.90	0.96	0.90	0.92	0.01	0.50 - 1.10
Glucose (mmol/L)	121.64 ^a	100.70 ^b	100.62 ^b	100.34 ^b	100.00 ^b	1.56	75.0 - 136.0
AST (IU/L)	56.17	56.50	56.01	55.93	55.79	0.75	13.0 - 110.0
ALP (IU/L)	148.40	146.50	143.31	143.84	143.09	9.12	130.0 - 513

a,bwith different superscripts in the same row (P<0.05); diet 1 (D1); with no DPM, D2 (5%), D3 (10%), D4 (15%) and D5 (20%) inclusion levels of DPM; TPP: total protein; ALB: albumin; GLB: globulin

Table 5: Serum biochemical indices of DPM

Serum minerals of weaned pigs fed DPM

Pigs given doum palm meal did not differ in serum concentrations of sodium (Na), potassium (P), chloride (Cl⁻), bicarbonate, calcium (Ca), and phosphorus (P>0.05) (Table 6). All readings, however, fell within the optimal ranges for pigs as reported by [41], indicating that there is no metabolic disease or health issue with the animals. The outcome is

consistent with the research conducted by [42], which found no variations in the calcium, bicarbonate, and sodium levels of weaner pigs fed [41, 43]. According to [44, 45], potassium is essential for heart, neuron, and muscle function. Sodium reflect a part of kidney function and elevated levels of sodium chloride, bicarbonate suggests a metabolic disorder or acidosis [46,47]. Phosphorus helps to regulate chemical reactions in the body [48].

Variables	D1	D2	D3	D4	D5	SEM	Ref. value
Sodium (mmol/L)	146.8	140.5	142.1	143.5	141.9	9.05	131 - 151
Potassium (mmol/L)	3.81	3.93	3.90	3.91	3.95	0.18	3.70 - 6.10
Chloride (mmol/L)	95.67	96.02	96.11	96.72	96.18	1.67	93.0 - 108.0
Bicarbonate (mmol/L)	18.65	18.06	19.10	19.15	19.74	0.92	19.0 - 31.0
Calcium (mmol/L)	10.98	10.95	11.22	11.20	11.27	0.51	9.50 - 12.5
Phosphorus (mmol/L)	6.57	6.54	6.63	6.67	7.00	0.11	6.30 - 11.50

Diet 1 (D1); with no DPM, diet 2 (D2) [5% DPM]; diet 3 (D3) [10% DPM], diet 4 (D4) [15% DPM] and diet 5 (D5) [20% DPM]

Table 6: Serum minerals of DPM

Conclusion

In conclusion, doum palm meal doesn't have any detrimental effects on pigs' serum parameters, making it safe to use as a feed ingredient in swine feed. DPM can also be used to replace up to 20% of maize without having an impact on the pigs' health.

References

1. Wadhwa, M., Bakshi, M.P.S., (2013). Utilization of fruit and vegetable wastes as livestock feed and as substrates for generation of other value-added products. Rap Publication, 4.
2. Faniyi, G.F. (2002). Replacement of wheat offal with untreated citrus pulp in broiler chick diets. Tropical Animal Prod. Invest, 5(95):100.
3. Blandon, J. C., Hamady, G. A. A., Abdel-Moneim M. A. (2015). The effect of partial replacement of yellow corn by banana peels with and without enzymes on broiler's performance and blood parameters. Journal of Animal sciences, 4 (2): 10-19.
4. Ecocrop (2010). Ecocrop database.<http://www.feedipedia.org/node/745>[09/12/2016 15:06:21]
5. Broomhead, J. (2013). The benefits and concerns of utilizing alternative feed ingredients. International Poultry Production, 21(6): 11-13.
6. Ekenyem, B.U., Madubuike, F.N., Dike, O.F. (2006). Effect of Partial Replacement of Yam Peel Meal Dioscorea Spp. for Maize Meal Zeamayson Performance and Carcass Characteristics of Finisher Broiler Chicks. International Journal of Poultry Science. 5 (10): 942-945.
7. Bogart, R., Taylor, R.E. (1983). Scientific Farm Animal Production. 2nd ed., MacMillan Pub. Co., New York, USA.
8. Hsu B, Coupur IM, Ng K. (2006). Antioxidant activity of hot water extract from the fruit of the Doum Palm, *Hyphaene thebaica*. Food Chemistry, Elsevier Science Direct. 98: 317-328.
9. Abdulsalam, I., Magaji, M.Y and Bah, S.U. (2018). Effects of Dietary Levels of Doum Palm Pulp Meal (*Hyphaene thebaica*) Supplementation on the Performance of Broiler Chickens. Asian Journal of Research in Animal and Veterinary Sciences. 2(2): 1-8.
10. Esonu BO, Emenelom OO, Udedibbie A. Anyanwu AB, Herbert U, Ekpor C.F. (2001). Performance and blood chemistry of weaner pigs fed raw mucuna (Velvet bean) meal. Tropical Animal Production Investigation. 4:49-54.
11. Waleed A, Zangh L, Mohammed D, Mohammed, Abubakar M, Elshareif O, Malik T. (2014). Physicochemical, Nutrition and functional properties of the Epicarp, Flesh and Pited sample of Doum palm fruit (*Hyphaene thebaica*). Journal of Food and Nutrition Research. 24:180-186.
12. Reda A.A. (2015). Physicochemical properties of Doum (*Hyphaene thebaica*) fruits and utilization of its flour in formulating some functional foods. Alexandria Journal of Food Science and Technology. 12(2):29-39.
13. Hussein, A.M, Salah, Z.A, Hegazy, N.A. (2010). Physicochemical, sensory and functional properties of wheat-

- Doum fruit flour composite cakes. *Polish Journal of Food and Nutrition Sciences*. 60(3):239- 244.
14. Datti, M., Ibrahim, I. Salihu, Y, M. Abdulhadi, S. M. Muhammad, S. A. Abubakar S., Halima, Ahmad, U and T. Nura (2020). Mineral Content, Proximate Composition and the Antioxidant Properties of the Ethanol Extract of *Hyphaene thebaica* L. from Gezawa Town, Kano State, Nigeria. *Asian Journal of Applied Chemistry Research*. 6(2): 33-40.
 15. NRC (2012). National Research Council. Quality protein maize. National Academy Press, Washington DC. 2012 pp. 1-70.
 16. Alagbe Olujimi John and Anuore, Daniel Nnadozie. (2023), Effect of Doum palm mesocarp meal (*Hyphaene thebaica*) as partial replacement for maize on growth performance and hematological indices of weaned pigs. *Journal of Biotechnology and Bioinformatics Research*. 5(3): 1-6.
 17. Siddeeg A., Salih, Z.A., Al-Farga, A, Ata-Elfadeel, E.M.A and Ali, A. (2019), Physicochemical, Nutritional and Functional Properties of Doum (*Hyphene thebaica*) Powder and Its Application in Some Processed Food Products. *Journal of Nutrition and Food Science Forecast*. 2(1): 1009-1014.
 18. Aboshora W, Idriss SE, Omar KA. (2017) Volatile Flavor Compounds of peel and pulp from Doum (*Hyphaene thebaica* L.) Fruit. *American Journal of Food Science and Nutrition Research*. 4: 165-169.
 19. Bonde, S.D., Agate, V.V., Kulkarni, D.K. (1990). Nutritional composition of the fruits of Doum palms from west coast of India. *Principles*. 34(1): 21-23.
 20. Alagbe, J.O. (2023). Bioactive compounds in ethanolic extract of *Strychnos innocua* root using gas chromatography and mass spectrometry (GC-MS). *Drug Discovery*, 17:e4dd1005.
 21. Agubosi, O.C.P., Oluwafemi, R.A and Alagbe, J.O. (2021). The effect of processing on the proximate, mineral and vitamin composition of Neem leaves (*Azadirachta indica*) grown in Gwagwalada, FCT, Abuja. *Abuja Journal of Agriculture and Environment*, 1(1): 293-299.
 22. Alagbe Olujimi John., Anuore, Daniel Nnadozie., Shittu Muritala Daniel and Ramalan, Sadiq Mohammad. (2023). Growth performance and physiological response of weaned pigs fed diet supplemented with novel a phytochemicals. *Brazilian Journal of Science*. 3(1): 43-57.
 23. Alagbe, J.O., Bamigboye, S., Nwosu, G.C., Agbonika, D.A and Kadiri Mercy Cincinsoko. (2023). Characterization of bioactive compounds in *Luffa aegyptiaca* leaf ethanolic extracts using gas chromatography and mass spectrometry (GC-MS). *Drug Discovery*, 17:e10dd1011.
 24. Alagbe, Olujimi John, Oluchi, C.P Agubosi and Rufus, Adebisi Oluwafemi. (2023). Histopathology of broiler chickens fed diet supplemented with *Prosopis africana* (African mesquite) essential oil. *Brazilian Journal of Science*. 2(9): 49-59.
 25. Alagbe, J.O., Kadiri, M.C., Oluwafemi, R.A., Agubosi, O.C.P and Anorue, D.N. (2023). Analysis of bioactive compounds in ethanolic extracts of *Xylopiya aethiopicum* leaves using gas chromatography and mass spectrometry technique. *American Journal of Science on Integration and Human Development*. 1(1): 1-10.
 26. Alagbe Olujimi John. (2023). Sensory evaluation and fatty acid composition of broiler chickens fed diets containing *Prosopis africana* oil. *Journal of Healthcare and Biomedical Science*. 1(2): 36-45.
 27. Cooper, C.A., Moraes, L.E., Murray, J.D and Owen, S.D. (2014). Hematologic and biochemical reference intervals for specific pathogens free 6 weeks Hampshire and Yorkshire crossbred pigs. *Journal of Animal Science and Biotechnology*. 10(5): 1-5.
 28. Alagbe, Olujimi John, Oluchi, C. Precious Agubosi., Rufus, Adebisi Oluwafemi., Taiwo, Oladoye Akande., Adegoke, Emmanuel Adebite and Emiola, I.A. (2023). Haemato-biochemical indices and intestinal microbial population of broiler chickens fed diet supplemented with *Prosopis africana* (African mesquite) essential oil. *Brazilian Journal of Science*. 2(9): 98-110.
 29. Omokore, E.O and Alagbe, J.O. (2019). Efficacy of dried *Phyllanthus amarus* leaf meal as an herbal feed additive on the growth performance, haematology and serum biochemistry of growing rabbits. *International Journal of Academic Research and Development*. 4(3): 97-104.
 30. Alade, A.A., Adeyemi, M.B., Abimbola, O.O., Babatunde, A.A., Nnamdi, M.A and Leonard, N. A. (2020). effects of dietary inclusion of *Zymomonas mobilis* treated sawdust on haematology, serum biochemistry, carcass characteristics and sensory evaluation of meat of broiler chickens. *Slovak Journal of Animal Science*. 53(4): 168-183.
 31. Oloruntola, O.D., Ayodele, S.O., Adeyeye, S.A., Omoniyi, S.I. (2016) Effect of *Alchomea cordifolia* leaf meal enzyme supplementation on growth, haematology and immunostimulatory response of rabbits. *Asian Journal of Biological and Life Sciences*. 5(2): 190-195.
 32. Adewale, A.O., Alagbe, J.O., Adeoye, Adekemi. O. (2021). Dietary Supplementation of *Rauvolfia Vomitoria* Root Extract as A Phytochemical Feed Additive in Growing Rabbit Diets: Haematology and serum biochemical indices. *International Journal of Orange Technologies*. 3(3): 1-12.
 33. Singh, A.S., Alagbe, J.O., Sharma, S., Oluwafemi, R.A and Agubosi, O.C.P. (2021). Effect of dietary supplementation of melon (*Citrullus linatus*) seed oil on the growth performance and antioxidant status of growing rabbits. *Journal of Multidimensional Research and Reviews*. 2(1): 78-95.
 34. Shittu, M.D., Alagbe, J.O., Adejumo, D.O., Ademola, S.G., Abiola, A.O., Samson, B.O and Ushie, F.T. (2021). Productive Performance, Caeca Microbial Population and Immune-Modulatory Activity of Broiler Chicks Fed Different Levels *Sida Acuta* Leaf Extract in Replacement of Antibiotics. *Bioinformatics and Proteomics Open Access Journal*. 5(1): 000143.
 35. Oluwafemi, R.A., Egwuiyi. G.N and Alagbe, J.O. (2020). Effect of feeding *Polyalthia longifolia* leaf meal as partial replacement of wheat offal. *European Journal of Agricultural and Rural Education*. 1(1): 8-16.
 36. American Metabolic Testing Laboratories, Inc. Chemistry Profile. (2001). www.caprofile.net/t3.html.
 37. Merck Veterinary Manual. Merck Veterinary Manual 10th edition. Merck and Co. Inc. Rahway N.J. 2010
 38. Brockus, C.W., Mahaffey, E.A., Bush, S.E and Krupp Despain, A.W. (2005). Hematologic and serum biochemical reference intervals for Vietnamese potbellied pigs. *Complementary Clinical Pathology*. 13: 162-165.
 39. Dixon, J.L., Stoops, J.D., Parker, J.L., Laughin, M.H., Weisman, G.A. (1999). Dyslipidemia and vascular dysfunction in diabetic pigs fed an atherogenic diet. *Arterioscler Biology*. 19(12): 2981-2983.
 40. Elliott, T.B., Deutz, N.E., Gulani, J., Koch, A., Olsen, C.H., Christensen, C., Chappelli, M. (2014). Gastrointestinal acute radiation syndrome in Gottingen minipigs. *Complementary Medicine*. 64(4): 456-463.
 41. Klem, T.B., Bleken, E., Morberg, H., Thoresen, S.I., Framstad, T. (2010). Hematology and biochemistry reference values for swine for Nowagian crossbred growers pigs. *Veterinary Clinical Pathology*. 39(2): 221-226.

42. Friendship, R.M., Lumsden, J.H., McMillian, I and Wilson, M.R. (1984). Hematology and biochemistry reference values for swine for Ontario swine. *Canadian Journal of Complementary Medicine*. 48(4): 390-393.
43. Blandon, J. C., Hamady, G. A. A., Abdel-Moneim, M. A. (2015). The effect of partial replacement of yellow corn by banana peels with and without enzymes on broiler's performance and blood parameters. *Journal of Animal and Poultry Sciences (JAPSC)*. 4(1): 10-19.
44. Adebayo A and Tukur, A. (2005). Adamawa State in Maps. Paraclete Publishers, Yola, Nigeria 112.
45. Clark, S.G and Coffey, N. (2008). Normal haematology and hematologic disorders in potbellied pigs. *American Exotic Animal Practice*. 11(3): 569-582.
46. Martini, W.Z., Cortez, D.S., Dubick, M.A., Blackbourne, L.H. (2012). Different recovery profiles of coagulation factors, thrombin generation and coagulation function after hemorrhagic shock in pigs, *Trauma Acute Care Surgery*. 73(3): 640-647.
47. Martini, W.Z., Cortez, D.S., Dubick, M.A., Blackbourne, L.H. (2019). Fibrinogen availability and coagulation function after haemorrhage and resuscitation in pigs. *Molecular Medicine*. 17(7-8): 640-647.
48. Rothwell, S.W., Sawyer, E., Dorsey, J., Flournoy, W.S., Settle, T., Simpson, D., Cadd, G. (2009). Wound healing and the immune response in swine treated with hemostatic bandage composed of salmon thrombin and fibrinogen. *Journal of Medicine*. 20(10): 2155-2156.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article Click Here:

Submit Manuscript

DOI:10.31579/2637-8914/190

Ready to submit your research? Choose Auctores and benefit from:

- fast, convenient online submission
- rigorous peer review by experienced research in your field
- rapid publication on acceptance
- authors retain copyrights
- unique DOI for all articles
- immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <https://auctoresonline.org/journals/nutrition-and-food-processing>