

## Short Communication: The Effects Chronic Acetaminophen Treatment on Age-Associated Alterations of Cardiac Function in the Female F344xBN Heart

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Received date: September 09,2018; Accepted date : October 10,2018; Published date: October 16,2018.

Citation for this Article: Kevin M. Rice. (2018) Short Communication: The Effects Chronic Acetaminophen Treatment on Age-Associated Alterations of Cardiac Function in the Female F344xBN Heart, J.Clinical Cardiology and Cardiovascular Interventions. 1(2); Doi:10.31579/2641-0419/10

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### Abstract

#### Background

Although several studies have investigated the age-associated changes in male and female F344 as well as male Fischer 344/NNiaHSD x Brown Norway/BiNia F1 (F344xBN), no study to our knowledge has examined the age-associated changes in structure and function in the female F344xBN heart using echocardiographic measures. This information is crucial in determining whether or not the female F344xBN is an appropriate aging model. Additional studies have also suggested that age-associated increases in levels of oxidative stress may cause cardiac dysfunction and that chronic acetaminophen (APAP) ingestion may be protective against increased oxidative stress. On the basis of these studies we examined the function and structure of the aged female F344xBN heart in the absence and presence of chronic APAP treatment.

#### Methods

To investigate if the aging-related changes in cardiovascular structure and function can be attenuated with APAP treatment, aged (22-month old) female F344xBN rats were administered APAP (30 mg/kg body weight/day) for 8 months in drinking water and echocardiograms assessments were performed.

#### Results

Aging was associated with evidence of diastolic (impaired left ventricle relaxation time) and systolic dysfunction (fractional shortening and end systolic volume). The incidence of arrhythmias was not different with age. However, valvular dysfunction was increased. Chronic APAP treatment did not attenuate the age-associated changes in cardiac structure and function in the female F344xBN. However the occurrence of valve dysfunction with aging was significantly lowered in the APAP treated 30-month female hearts.

### Introduction

The American Heart Association estimates that 80 million Americans have one or more types of cardiovascular disease (CVD). It is estimated that 38.1 million of those afflicted are aged 60 or older. As a person ages he or she has a higher risk of developing myocardial infarction, stroke, atherosclerosis, peripheral occlusive disease, diabetes, hypertension [1-3]. The potential mechanism of increase risk of disease with age is that there is an increase of reactive oxygen species (ROS) which leads to oxidative stress [4]. Recently acetaminophen (APAP), an antipyretic and analgesic, has shown to be protective against oxidative stress in the heart and the vasculature of the brain due to its antioxidant properties [5-7].

Due to these antioxidant effects APAP is considered to be cardioprotective during ischemia/reperfusion, hypoxia/reoxygenation, exogenous peroxynitrite administration, and experimentally induced

myocardial infarction due to its ability to block or reduce the development of mitochondrial permeability transition pores, mitochondrial swelling, cytochrome c release, late stage apoptosis, protein oxidation, production of hydroxyl radicals, arrhythmias, and peroxynitrite induced dysfunction in hearts [5, 8-16].

The Fischer 344/NNiaHSD x Brown Norway/BiNia F1 (F344xBN) has been recommended by the National Institute of Aging as a rat model of aging due to its increased life span and fewer age-associated pathologies compared to the standard model of aging – the Fischer 344 rat. Although several studies have investigated the age-associated changes in male and female F344 rats as well as male F344xBN rats [17-21], no study to our knowledge has examined the age-associated changes in structure and function in the female F344xBN heart using echocardiographic measures.

This information is crucial in determining whether or not the female F344xBN is an appropriate aging model to study the development of cardiovascular disease. Due to evidence that age-associated increase in levels of oxidative stress may cause cardiac dysfunction, we also determined the function and structure of the aged female F344xBN heart with chronic APAP treatment. The purpose of this study was to determine the age-associated changes in the female F344xBN cardiac structure and function and whether chronic APAP treatment would attenuate these effects.

## Materials and Methods

### Animals

All procedures were performed in accordance with the Guide for the Care and Use of Laboratory Animals as approved by the Council of the American Physiological Society and the Animal Use Review Board of Marshall University. All procedures were conducted in strict accordance with the Public Health Service animal welfare policy. Young (6-month), adult (22-month), aged (26-month), and very aged (30-month) female F344xBN rats were obtained from the National Institute for Aging and were housed two per cage in an Association for Assessment and Accreditation of Laboratory Animal Care International (AAALAC) approved vivarium. Housing conditions consisted of a 12 h-12 h light-dark cycle with a temperature maintained at  $22 \pm 2^\circ\text{C}$ . Animals were provided with food and water *ad libitum*. Rats were allowed to recover from shipment for at least two weeks before experimentation. During this time the animals were carefully observed and weighed weekly. Any rat showing signs of failure to thrive, such as precipitous weight loss, disinterest in the environment, or unexpected gait alterations were excluded from the study. Six Female F344xBN rats were randomly assigned to each group (6C – 6-month animal; 22C – 22-month animal; 26C – 26-month animal; 26T – 26-month APAP-treated animal; 30C – 30-month animal; 30T – 30-month APAP-treated animal).

### APAP Treatment

A therapeutic dose of APAP (30 mg/kg/day) was administered through the drinking water for treated animals at 22- months of age. Changes in body weight and water intake were monitored throughout the study to maintain a therapeutic dose. The 6-, 22-, 26-, and 30-month age-matched control groups received tap water and were maintained under the same conditions as the APAP treated groups.

### Echocardiographic procedures

Control (6-, 22-, 26-, and 30-month) and APAP treated (26- and 30-month) rats were anesthetized with an intraperitoneal (ip) injection of ketamine (40 mg/kg) and xyaline (10 mg/kg). The ventral thoraxes were shaved, and the rats were placed either on their backs or left side and covered with ultrasonic transmission gel for adequate sonic transference. A Phillips 5500 ECHO system with a 12 MHz transducer was used to take two-dimensional ECHO measurements, two-dimensional guided M-mode, Doppler M-mode, and other recordings from parasternal long- and short-axis views. Parasternal long- and short axis views were used to determine two-dimensional cardiac structural measurements. The echocardiographic views were then used to position the M-mode echocardiographic line. In order to determine the presence of valve dysfunctions, valvular blood flow velocities were evaluated using pulse wave Doppler with the probe toward the apex (x-axis) and the depth along the y-axis (long axis procedure). Ejection fraction and fractional shortening during systole was calculated using the evaluation of wall structure from short-axis procedures with the probe oriented toward the left ventricle and across the heart. M-mode displays were analyzed by a digital echocardiographic analysis system.

The following measurements were selected for each assessment of cardiac structure and function. The structural parameters included diastolic (IVSd) and systolic (IVSs) left ventricular septal thickness, diastolic (LVIDd) and systolic (LVIDs)-

Left ventricular internal dimension, diastolic (LVPWd) and systolic (LVPWs) left ventricular posterior wall thickness, and right ventricular diastolic internal dimension (RVd). Functional measurement included ejection fraction (EF) and left ventricular fractional shortening during systole (FS).

Additional echocardiographic measurements included mitral valve deceleration (MV decel time), left ventricular mass (LVM), end-systolic volume (ESV), end-diastolic volume (EDV), peak velocity of the A wave (Amax), and peak velocity of the E wave (Emax) were used to evaluate systolic function. Mitral valve deceleration, Emax, Amax, and MV E/A ratio were used to evaluate diastolic function.

### Electrocardiogram (EKG) measurements and Heart Collection

After completion of treatment and echocardiographic procedures, animals were anesthetized with an ip injection of ketamine (40 mg/kg) and xyaline (10 mg/kg) and supplemented as necessary for reflexive responses. Before heart collection, electrocardiograms (EKGs) with leads I, II, and III were measured in age-matched control and treated animals using the Biopac Student Lab software (BIOPAC Systems, Inc., Microsoft). After completion of EKG measurements, the heart was removed after a midline laparotomy and was then placed in Krebs-Ringer bicarbonate buffer (KRB) containing the following: 118 mM NaCl, 4.7 mM KCl, 2.5 mM  $\text{CaCl}_2$ , 1.2 mM  $\text{KH}_2\text{PO}_4$ , 1.2 mM  $\text{MgSO}_4$ , 24.2 mM  $\text{NaHCO}_3$ , and 10 mM a-D-glucose (pH 7.4) equilibrated with 5%  $\text{CO}_2/95\%$   $\text{O}_2$  and maintained at  $37^\circ\text{C}$ . Isolated hearts were quickly massaged to remove any blood from the ventricles, cleaned of connective tissue, weighed, and immediately snap frozen in liquid nitrogen for further analysis.

### Statistical Methods

Results were reported as mean  $\pm$  SEM. Statistical analyses were performed using Sigma Stat 3.5 statistical software (Systat Software, Inc). Age comparisons between ECHO structural, functional parameters, and morphologic indices were evaluated by one-way ANOVA or Kruskal-Wallis one-way Analysis of Variance on Ranks with the Student-Newman-Keuls or Dunn's methods as the post hoc test, respectively. Differences between age-matched control and treated animals were determined by Student's T-test. The level of significance accepted a priori was  $P < 0.05$ .

## Results

### Echocardiographic Structural Parameters

Echocardiographic evaluation of cardiac structural parameters in female F344xBN rats are compared in Table 1. Left ventricular septal thickness (IVS) during systole (IVSs) significantly increased at 26-months ( $0.253 \pm 0.003$  cm) compared to 6-months ( $0.193 \pm 0.008$  cm;  $P < 0.05$ ); however, no age-associated changes during diastole (IVSd) were found. Left ventricular internal dimension during systole (LVIDs) significantly increased at 22- ( $0.340 \pm 0.006$  cm) and 26-months ( $0.331 \pm 0.009$  cm) compared to 6-month ( $0.378 \pm 0.017$  cm;  $P < 0.05$ ). Treatment with APAP significantly decreased LVIDs compared to control animals at 30-months ( $0.344 \pm 0.015$  vs  $0.356 \pm 0.011$  cm, respectively;  $P = 0.004$ ). During diastole LVID did not change with age but significantly increased with APAP treatment at 26-months ( $0.630 \pm 0.009$  cm) when compared to control ( $0.593 \pm 0.009$  cm;  $P < 0.001$ ). Left ventricular posterior wall thickness (LVPW) during systole (LVPWs) and diastole (LVPWd) was significantly increased at 22- ( $0.273 \pm 0.004$  and  $0.183 \pm 0.004$  cm, respectively) and 30-months ( $0.288 \pm 0.009$  and  $0.189 \pm 0.008$  cm, respectively) compared to 6-months ( $0.193 \pm 0.002$  and  $0.153 \pm 0.010$  cm, respectively;  $P < 0.05$ ). APAP treatment increased both LVPWs and LVPWd thickness at 26-months compared to control ( $0.314 \pm 0.007$  and  $0.185 \pm 0.003$  cm, respectively;  $P = 0.002$ ). No changes were found in right ventricular dimension during diastole or left ventricular mass with age or APAP treatment.

Groups	IVSs (cm)	IVSd (cm)	LVIDs (cm)	LVIDd (cm)	LVPWs (cm)	LVPWd (cm)	RVDd (cm)	LVM (g)
6C	0.193 ± 0.008	0.118 ± 0.012	0.378 ± 0.017	0.610 ± 0.022	0.193 ± 0.002	0.153 ± 0.010	0.105 ± 0.016	0.998 ± 0.051
22C	0.244 ± 0.003	0.147 ± 0.002	0.340 ± 0.006 <sup>a</sup>	0.611 ± 0.007	0.273 ± 0.004 <sup>a</sup>	0.183 ± 0.004 <sup>a</sup>	0.135 ± 0.003	1.099 ± 0.017
26C	0.253 ± 0.003 <sup>a</sup>	0.151 ± 0.003	0.331 ± 0.009 <sup>a</sup>	0.593 ± 0.009	0.267 ± 0.007	0.166 ± 0.004	0.138 ± 0.009	1.094 ± 0.017
26T	0.253 ± 0.005	0.144 ± 0.004	0.315 ± 0.007	0.630 ± 0.009 <sup>d</sup>	0.314 ± 0.007 <sup>d</sup>	0.185 ± 0.003 <sup>d</sup>	0.146 ± 0.10	1.121 ± 0.016
30C	0.239 ± 0.013	0.149 ± 0.004	0.356 ± 0.011	0.606 ± 0.10	0.288 ± 0.009 <sup>a</sup>	0.189 ± 0.008 <sup>a</sup>	0.111 ± 0.006	1.144 ± 0.036
30T	0.250 ± 0.004	0.149 ± 0.002	0.334 ± 0.015 <sup>d</sup>	0.644 ± 0.016	0.318 ± 0.013	0.208 ± 0.009	0.152 ± 0.018	1.218 ± 0.031

**Table 1.** Echocardiographic evaluation of cardiac structural parameters in female F344xBN rats (mean ± SEM; n = ## rats/group). 6C – 6-month animal; 22C – 22-month animal; 26C – 26-month animal; 26T – 26-month APAP-treated animal; 30C – 30-month animal; 30T – 30-month APAP-treated animal; <sup>a</sup>P < 0.05 significant difference from 6-month control animal; <sup>d</sup>P < 0.05 age-matched control vs. APAP treated animal.

### Echocardiographic Functional Parameters

Echocardiographic evaluation of cardiac functional parameters in female F344xBN rats can be found in Table 2 and 3. As described in Table 2, ejection fraction was significantly increased at 26-months (82 ± 1.0%) compared to 6-months (74 ± 0.9%; P < 0.05). APAP treatment at 26-months (86 ± 0.8%) also significantly increased the ejection fraction compared to control (P = 0.009). Fractional shortening significantly decreased at 30-months (41.5 ± 1.7%) compared to 26-months (45.6 ± 1.0%; P < 0.05). APAP treatment at 26-months (49.8 ± 1.2%) significantly increased the fractional shortening when compared to control (P = 0.007).

End systolic volume (ESV) was significantly decreased at 26-months (0.082 ± 0.005 mL) when compared to 6-months (0.143 ± 0.017 mL; P < 0.05). APAP treatment significantly decreased ESV at 30-months (0.084 ± 0.001 mL) when compared to control (0.114 ± 0.10 mL; P = 0.034). APAP treatment at 26-months (0.579 ± 0.021 mL) significantly increased end diastolic volume when compared to control (0.492 ± 0.20 mL; P = 0.005). Heart rate did not change with age but significantly increased with APAP treatment at 26- (286.6 ± 5.9 bpm; P = 0.015) and decreased at 30-months (238.7 ± 14.8 bpm; P = 0.029) when compared to controls.

Groups	Ejection Fraction (%)	Fractional Shortening (%)	ESV (mL)	EDV (mL)	Heart Rate (bpm)
6C	74 ± 0.9	38 ± 0.7	0.143 ± 0.017	0.535 ± 0.052	281 ± 16.5
22C	80 ± 0.8	44 ± 0.8	0.09 ± 0.004	0.505 ± 0.017	N.T.
26C	82 ± 1.0 <sup>a</sup>	46 ± 1.0	0.082 ± 0.005 <sup>a</sup>	0.492 ± 0.20	259 ± 8.8
26T	86 ± 0.8 <sup>d</sup>	50 ± 1.2 <sup>d</sup>	0.076 ± 0.005	0.579 ± 0.021 <sup>d</sup>	287 ± 5.9 <sup>d</sup>
30C	78 ± 1.8	42 ± 1.7 <sup>c</sup>	0.114 ± 0.10	0.523 ± 0.022	278 ± 10.1
30T	82 ± 2.5	46 ± 2.5	0.084 ± 0.001 <sup>d</sup>	0.623 ± 0.043	239 ± 14.8 <sup>d</sup>

**Table 2.** Echocardiographic evaluation of cardiac functional parameters in female F344xBN rats with and without APAP (mean ± SEM; n = ## rats/group). 6C – 6-month animal; 22C – 22-month animal; 26C – 26-month animal; 26T – 26-month APAP-treated animal; 30C – 30-month animal; 30T – 30-month APAP-treated animal; <sup>a</sup>P < 0.05 significant difference from 6-month control animal; <sup>c</sup>P < 0.05 significant difference from 26-month control animal; <sup>d</sup>P < 0.05 age-matched control vs. treated animal; N.T. – not tested.

As described in Table 3, no age-associated changes in E-E' ratio was found in the female F344xBN rats. APAP treatment increased the E-E' ratio at 26-months (19.61 ± 1.42) compared to control (14.73 ± 0.64; P = 0.017). Left ventricular isovolumetric relaxation time (LV IVRT) was significantly increased at 22- (0.029 ± 0.001 sec) and 26-months (0.036 ± 0.005 sec) compared to 6-months (0.015 ± 0.002 sec; P < 0.05). Treatment with APAP at 26-months (0.028 ± 0.002 sec) significantly decreased LV IVRT compared to control (P < 0.001). There were no changes in mitral valve deceleration (MV Dec) time with age or APAP treatment.

With age mitral valve peak velocity of the E wave (MV Emax) significantly decreased at 22- (59.8 ± 1.2 cm/sec), 26- (61.5 ± 1.5 cm/sec), and 30-months (55.4 ± 1.4 cm/sec) when compared to 6-months (78.0 ± 3.3 cm/sec; P < 0.05). Aging also significantly decreased peak velocity of the A wave (MV Amax) at 30-months (35.1 ± 1.6 cm/sec) when compared to 6-months (41.5 ± 1.3 cm/sec; P < 0.05). No change in MV Emax or Amax was found with APAP treatment. No changes were found in MV E/A ratio with age or APAP treatment.

Groups	E-E'	LV IVRT (sec)	MV Dec Time (sec)	MV Emax (cm/sec)	MV Amax (cm/sec)	MV E/A
6C	20.8 ± 1.02	0.015 ± 0.002	0.053 ± 0.005	78 ± 3.3	42 ± 1.3	1.75 ± 0.03
22C	16.8 ± 0.68	0.029 ± 0.001 <sup>a</sup>	0.054 ± 0.002	60 ± 1.2 <sup>a</sup>	37 ± 0.8	1.63 ± 0.04
26C	14.7 ± 0.64	0.036 ± 0.005 <sup>a</sup>	0.057 ± 0.002	62 ± 1.5 <sup>a</sup>	36 ± 0.9	1.60 ± 0.06
26T	19.6 ± 1.42 <sup>d</sup>	0.028 ± 0.002 <sup>d</sup>	0.057 ± 0.002	60 ± 1.2	39 ± 1.4	1.56 ± 0.05
30C	21.9 ± 2.63	0.030 ± 0.000	0.063 ± 0.003	55 ± 1.4 <sup>a</sup>	35 ± 1.6 <sup>a</sup>	1.66 ± 0.10
30T	16.6 ± 1.30	0.028 ± 0.001	0.061 ± 0.003	57 ± 2.1	36 ± 1.4	1.56 ± 0.04

**Table 3.** Echocardiographic evaluation of cardiac functional parameters in female F344xBN rats with and without APAP (mean ± SEM; n = ## rats/group). 6C – 6-month animal; 22C – 22-month animal; 26C – 26-month animal; 26T – 26-month APAP-treated animal; 30C – 30-month animal; 30T – 30-month APAP-treated animal; <sup>a</sup>P < 0.05 significant difference from 6-month control animal; <sup>b</sup>P < 0.05 significant difference from 22-month control animal; <sup>c</sup>P < 0.05 significant difference from 26-month control animal; <sup>d</sup>P < 0.05 age-matched control vs. treated animal.

## Valvular Dysfunction

Tricuspid valve dysfunction was higher in 22-, 26-, and 30-month (52.4%, 21.7%, 33.3%) female hearts relative to 6-month. APAP treatment at 26-months (33.3%) had a higher percentage of female rats with tricuspid dysfunction compared to 26-month control group. However, with APAP treatment at 30-months the percentage of female rats with tricuspid dysfunction (33.3%) was the same when relative to the 6-month group. The percentage of female rats with mitral dysfunction also was higher with age (22C: 19.0%; 26C: 17.4%; 30C: 13.3%) relative to 6-month rats. APAP treatment at 26-months had a similar percentage of mitral dysfunction compared to control; however, APAP treatment at 30-months had a lower percentage when compared to the 30-month control group. The presence of aortic dysfunction with age was only seen in the 30-month control group (20%). At 26-months with APAP treatment (26T: 16.7%; 30T: 15.0%) the percentage of female rats with aortic dysfunction was higher compared to control groups (26C: 0%; 30C: 20.0%). The percentage of aortic dysfunction in 30-month APAP rats was lower (20.0%) than the 30-month control rats (15%). Female rats with pulmonary valve dysfunction were higher but this percentage was similar or lower with APAP treatment at 26- and 30-months (26T: 77.8%; 30T: 45.0%) when compared to control groups (26C: 78.3%; 30C: 66.7%).

## Electrocardiogram Analysis

The presence of arrhythmias was not found in the vast majority of female F344xBN rats, and therefore the incidence was not different between control and APAP treatment groups. In the 30-month APAP treated group one, premature ventricular contraction was observed (data not shown).

## Heart Weight, Body Weight, and Heart Weight to Body Weight Ratio

Female F344xBN heart weights and body weights were compared in Table 4. Heart weight increased significantly with age at 26-months ( $0.954 \pm 0.002$  g) compared to 6- and 22-month groups (6C:  $0.721 \pm 0.002$  g; 22C:  $0.787 \pm 0.02$  g;  $P < 0.001$ ). At 30-months ( $1.04 \pm 0.04$  g) heart weight significantly increased compared to all age groups ( $P < 0.05$ ). APAP treatment significantly increased heart weight at 30-months when compared to control ( $P = 0.029$ ). Body weight increased significantly at 26- ( $296.4 \pm 6.5$  g) and 30-months ( $315.6 \pm 8.6$  g) when compared to 6-month ( $235.3 \pm 1.7$  g;  $P < 0.05$ ). When normalized to body weight, heart weights were significantly decreased at 22-months ( $0.272 \pm 0.01\%$ ) compared to 6-months ( $0.307 \pm 0.02\%$ ;  $P < 0.05$ ). However, normalized heart weights at 26- ( $0.321 \pm 0.01\%$ ) and 30-month ( $0.327 \pm 0.01\%$ ) rats was significantly increased compared to 6-months ( $P < 0.05$ ). APAP treatment significantly decreased heart weight after normalization to body weight at 26-months ( $0.295 \pm 0.01\%$ ;  $P = 0.048$ ) but significantly increased at 30-months ( $0.367 \pm 0.01\%$ ;  $P = 0.002$ ) compared to controls.

Groups	N	Heart Weight (g)	Body Weight (g)	HW/BWRatio(%)
6C	4	0.72 ± 0.02	235.3 ± 1.7	0.317 ± 0.02
22C	41	0.79 ± 0.02	290.4 ± 13.0	0.27 ± 0.01 <sup>a</sup>
26C	22	0.95 ± 0.02 <sup>ab</sup>	296.4 ± 6.5 <sup>a</sup>	0.32 ± 0.01 <sup>b</sup>
26T	16	0.96 ± 0.04	325.2 ± 9.6 <sup>d</sup>	0.30 ± 0.01 <sup>d</sup>
30C	10	1.0 ± 0.04 <sup>abc</sup>	315.6 ± 8.6 <sup>a</sup>	0.33 ± 0.01 <sup>b</sup>
30T	11	1.1 ± 0.01 <sup>d</sup>	309.8 ± 6.7	0.37 ± 0.01 <sup>d</sup>

**Table 4.** Total body weight (BW) and heart weight (HW) in female F344xBN rats at 6-, 22-, 26-, and 30-months of age with or without APAP treatment (means ± SEM; n = ## rats/group). 6C – 6-month animal; 22C – 22-month animal; 26C – 26-month animal; 26T – 26-month APAP-treated animal; 30C – 30-month animal; 30T – 30-month APAP-treated animal; <sup>a</sup> $P < 0.05$  significant difference from 6-month control animal; <sup>b</sup> $P < 0.05$  significant difference from 22-month control animal; <sup>c</sup> $P < 0.05$  significant difference from 26-month control animal; <sup>d</sup> $P < 0.05$  age-matched control vs. treated animal.

## Discussion

The present study was conducted to obtain the cardiac function reference values for the aging female F344xBN rats. The F344xBN rat, according to the National Institute of Aging, is the preferred animal model to study age-associated pathophysiological changes due to its longer life span and low occurrence of pathologies [22]. The 6-, 22-, 26-, and 30-month age groups were chosen based off survivability curves from the National Institute of Aging to represent females in the third, seventh, and eighth decade of life [22, 23]. These reference values are critical in order to compare the effects of aging on systolic and diastolic function in order to establish if the female F344xBN rat is an appropriate model for aging cardiac dysfunction.

Previous work in our laboratory has indicated that with aging there is an increase in oxidative-nitrosative stress which may responsible for the age-associated changes in cardiac structure and function [24-26]. This age-associated increase in oxidative-nitrosative stress is attenuated with chronic APAP treatment [20] due to its antioxidative properties. In order to investigate the role of age-associated oxidative-nitrosative stress on cardiac structure and function, APAP was administered to the treatment groups in order to see if it would attenuate these age-associated changes in cardiac function.

Increased risk of diastolic dysfunction with aging appears to occur more often in women compared to men [27]. These diastolic dysfunctions oftentimes precede the development of systolic dysfunction [27]. Diastolic dysfunction in humans is defined by prolonged deceleration time, a high A wave velocity, a low E wave velocity, and prolonged isovolumetric relaxation [27]. In the present study of the female F344xBN heart, we observed age-related increases in left ventricular relaxation time, decrease in E', and a trend in mitral valve deceleration time. Nonetheless, we did not find any changes in the E/A ratio with age nor did APAP treatment produce a change (Table 3). In contrast, Boluyt *et al.* show that aged female F344 animals have increased LV IVRT, decreased E wave, and increased A wave velocity [28]. In our study, as in the Boluyt *et al.* study with age there was a longer left ventricular relaxation time, a reduced E' wave velocity, a trend toward an increased mitral valve deceleration time, and no change in the E/A ratio [28]. Taken together, these data suggest that aging in female F344 and F344xBN rats, much like that seen in humans, is characterized by changes in diastolic function. APAP treatment had no effect on this age related change.

As previously observed in F344 rats, systolic function in female F344xBN rats did not appear to be significantly impaired with aging (Table 2) [28, 29]. Similar to the F344 rats, we found a slight increase in 26-month ejection fraction; this parameter, however, must be interpreted carefully as there was also increased valvular regurgitation at this measurement time [28]. Forman *et al.* find that F344 male rats have higher incidence of mitral regurgitation (MR) relative to female rats [29]. This may explain differences in age related functional changes between male and female rats. More investigation is needed to determine the mechanism of age related differences in cardiac function in male and female F344xBN rats. Nonetheless, our data are consistent with the notion that APAP significantly improved the ejection fraction (and therefore increased the EDV and reduced the ESV) at 26 months and there was trend towards improvement at 30 months (note that the ESV was significantly lower in treated compared to age matched controls at this time point).

Unlike our previous data regarding the presence of age associated arrhythmias in the male F334xBN [17], the aging female F344xBN failed to demonstrate age associated arrhythmias. However, APAP treatment attenuated the age associated incidents of valvular dysfunction.

These data suggest that aging in the female F344xBN rat heart is associated with changes in cardiac structure (Table 1) and function (Tables 2 and 3) and that chronic APAP treatment may diminish the incidence of valvular dysfunction in most situations, but increase the incidence in others (e.g., increased tricuspid dysfunction at 26 months).



APAP appears to result in beneficial changes in ejection fraction (Table 2), but a variable effect on heart rate (i.e., increased at 26 months, but decreased at 30 months compared to control). This study provides reference values for cardiac structure and function for adult female F344XBN rats. Further investigation regarding other parameters of cardiac function is currently underway.

## References

1. Sohal, R.S. and R. Weindruch, Oxidative stress, caloric restriction, and aging. *Science*, 1996. 273(5271): 59-63.
2. Finkel, D., et al., Quantitative genetic analysis of biobehavioral markers of aging in Swedish studies of adult twins. *J Aging Health*, 2000. 12(1): 47-68.
3. Finkel, D. and N.L. Pedersen, Contribution of age, genes, and environment to the relationship between perceptual speed and cognitive ability. *Psychol Aging*, 2000. 15(1): 56-64.
4. Trifunovic, A. and N.G. Larsson, Mitochondrial dysfunction as a cause of ageing. *J Intern Med*, 2008. 263(2): 167-178.
5. Merrill, G.F., Acetaminophen and low-flow myocardial ischemia: efficacy and antioxidant mechanisms. *Am J Physiol Heart Circ Physiol*, 2002. 282(4): H1341-9.
6. Tripathy, D. and P. Grammas, Acetaminophen protects brain endothelial cells against oxidative stress. *Microvasc Res*, 2009. 77(3): 289-296.
7. Tripathy, D. and P. Grammas, Acetaminophen inhibits neuronal inflammation and protects neurons from oxidative stress. *J Neuroinflammation*, 2009. 6: 10.
8. Golfetti, R., VanDyke K, Rork T, Spiler N, Merrill G. Acetaminophen in the post-ischemia reperfused myocardium. *Exp Biol Med (Maywood)*, 2002. 227(11): 1031-7.
9. Golfetti, R., T. Rork, and G. Merrill, Chronically administered acetaminophen and the ischemia/reperfused myocardium. *Exp Biol Med (Maywood)*, 2003. 228(6): 674-682.
10. Merrill, G., McConnell P, Vandyke K, Powell S. Coronary and myocardial effects of acetaminophen: protection during ischemia-reperfusion. *Am J Physiol Heart Circ Physiol*, 2001. 280(6): H2631-648.
11. Merrill, G.F. and E. Goldberg, Antioxidant properties of acetaminophen and cardioprotection. *Basic Res Cardiol*, 2001. 96(5):423-30.
12. Merrill, G.F., et al., Acetaminophen and myocardial infarction in dogs. *Am J Physiol Heart Circ Physiol*, 2004. 287(5): H1913-920.
13. Rork, T.H, Knox Van Dyke, Norell M. Spiler, Gary F. Merrill Acetaminophen in the hypoxic and reoxygenated guinea pig myocardium. *Exp Biol Med (Maywood)*, 2004. 229(11): 1154-1161.
14. Merrill, G.F., Merrill JH, Golfetti R, Jaques KM, Hadzimichalis NS et al, Antiarrhythmic properties of acetaminophen in the dog. *Exp Biol Med (Maywood)*, 2007. 232(9): 1245-1252.
15. Hadzimichalis, N.M., Baliga SS, Golfetti R, Jaques KM, Firestein BL, et al., Acetaminophen-mediated cardioprotection via inhibition of the mitochondrial permeability transition pore-induced apoptotic pathway. *Am J Physiol Heart Circ Physiol*, 2007. 293(6): H3348-3355.
16. Jaques-Robinson, K.M., Golfetti R, Baliga SS, Hadzimichalis NM, et al., Acetaminophen is cardioprotective against H<sub>2</sub>O<sub>2</sub>-induced injury in vivo. *Exp Biol Med (Maywood)*, 2008. 233(10): 1315-1322.
17. Rice, K., Sunil K. Kakarla , Srinivas Thulluri , Nandini D.P.K. Manne , et al., Anti-Arrhythmic Effect of Chronic Acetaminophen Treatment in the Aging F344XBN Rat Involves Diminished Myocardial Fibrosis and Altered Micrornas Regulation : Additive Role of Three Dimensional Echocardiography to Aid in the Diagnosis of Left Ventricular Thrombus. *Heart Circ*, 2017. 1(019).
18. Wu, M., et al., Katta A, Gadde MK, Liu H, Kakarla SK Aging-associated dysfunction of Akt/protein kinase B: S-nitrosylation and acetaminophen intervention. *PLoS One*, 2009. 4(7): e6430.
19. Wu, M., Desai DH, Kakarla SK, Katta A, Acetaminophen prevents aging-associated hyperglycemia in aged rats: effect of aging-associated hyperactivation of p38-MAPK and ERK1/2. *Diabetes Metab Res Rev*, 2009. 25(3): 279-286.
20. Kakarla, S.K., Fannin JC, Keshavarzian S, Katta A, et al., Paturi S Chronic acetaminophen attenuates age-associated increases in cardiac ROS and apoptosis in the Fischer Brown Norway rat. *Basic Res Cardiol*, 2010. 105(4): 535-544.
21. Rice, K.M., et al., Chronic paracetamol treatment influences indices of reactive oxygen species accumulation in the aging Fischer 344 X Brown Norway rat aorta. *Ann Clin Lab Sci*, 2012. 42(2): 152-61.
22. Turturro, A., William W. Witt Sherry Lewis Bruce S. Hass et al., Growth curves and survival characteristics of the animals used in the Biomarkers of Aging Program. *J Gerontol A Biol Sci Med Sci*, 1999. 54(11): B492-501.
23. Lipman, R.D., Clarence E. Chrisp DeWitt G. Hazzard Roderick T. Bronson et al., Pathologic characterization of brown Norway, brown Norway x Fischer 344, and Fischer 344 x brown Norway rats with relation to age. *J Gerontol A Biol Sci Med Sci*, 1996. 51(1):B54-59.
24. Fannin, J., Srinivas Thulluri, Ravi Kumar Arvapalli, Paulette Wehner et al., The Effects of Aging on Indices of Oxidative Stress and Apoptosis in the Female Fischer 344/Nnia X Brown Norway/BiNia Rat Heart. *Open Cardiovasc Med J*, 2013. 7:113-121.
25. Arvapalli, R.K., Paturi S, Laurino JP, Katta A, Kakarla SK et al., Deferasirox decreases age-associated iron accumulation in the aging F344XBN rat heart and liver. *Cardiovasc Toxicol*, 2010. 10(2): 108-116.
26. Asano, S., Katta A, Desai DH, Walker EM, Wehner P et al., Aging influences multiple indices of oxidative stress in the heart of the Fischer 344/NNia x Brown Norway/BiNia rat. *Redox Rep*, 2007. 12(4): 167-180.
27. Hamlin, S.K., et al., Role of diastole in left ventricular function, II: diagnosis and treatment. *Am J Crit Care*, 2004. 13(6): 453-66; quiz 467-468.
28. Boluyt, M.O., Converso K, Hwang HS, Mikkor A, Russell MW. et al., Echocardiographic assessment of age-associated changes in systolic and diastolic function of the female F344 rat heart. *J Appl Physiol*, 2004. 96(2): 822-888.
29. Forman, D.E., Antonio Cittadini, Gohar Azhar, Pamela S Douglas and Jeanne Y Weiet al., Cardiac morphology and function in senescent rats: gender-related differences. *J Am Coll Cardiol*, 1997. 30(7): 1872-1877.