

## Mini-review: Heparin and Amyloid $\beta_2$ -Microglobulin

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Heparin is a major component of glycosaminoglycans (GAGs), which are components of tissues in the interstitial spaces of the body. Heparin is the most important anticoagulant used in clinical settings, especially for hemodialysis (HD).

Since Gejo's report in 1985,  $\beta_2$ -microglobulin ( $\beta_2$ -M) has been recognized as the precursor protein in dialysis-related amyloidosis (DRA), which is inevitably associated with long-term HD [1]. In general, this amyloidosis develops in the presence of two essential background conditions, i.e.,

extremely high concentrations of precursor proteins and amyloidogenic conformation of these precursor proteins. Connors [2] first demonstrated the amyloidogenic potential of  $\beta_2$ -M in 1985. Serum  $\beta_2$ -M levels in end-stage renal failure were known to be elevated because the kidney is the main organ related to the metabolism of  $\beta_2$ -M. In addition, we later proved that the conversion of the native  $\beta_2$ -M conformation to the amyloidogenic  $\beta_2$ -M conformation was triggered by unfolding of the C-terminal portion from Ile-92 to Met-99 [3, 4] (Figure 1).

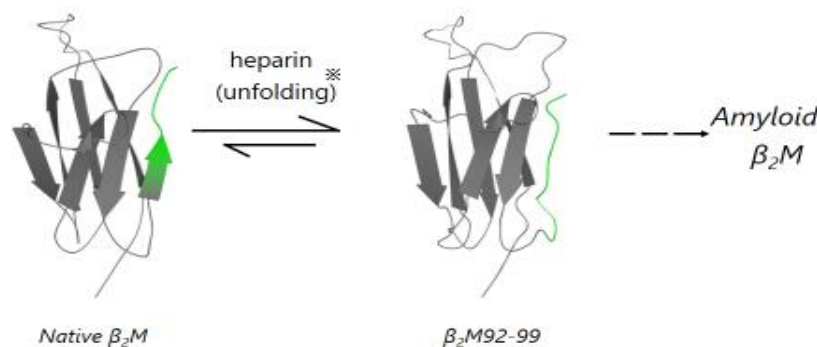


Figure 1

**Figure 1:** C-terminal unfolding : destructuring by loss of intramolecular H-bond involved in C-terminal 92-99

However, how native  $\beta_2$ -M is converted to an amyloidogenic conformer with the unfolded C-terminus in vivo, i.e., in clinical settings such as HD, and how  $\beta_2$ -M concentrations in the interstitial space reach as high as millimolar values despite micromolar serum levels of  $\beta_2$ -M in patients with HD [5] have not been clarified. In this mini-review, we would like to demonstrate that heparin may be a key molecule for the answers to both of these questions.

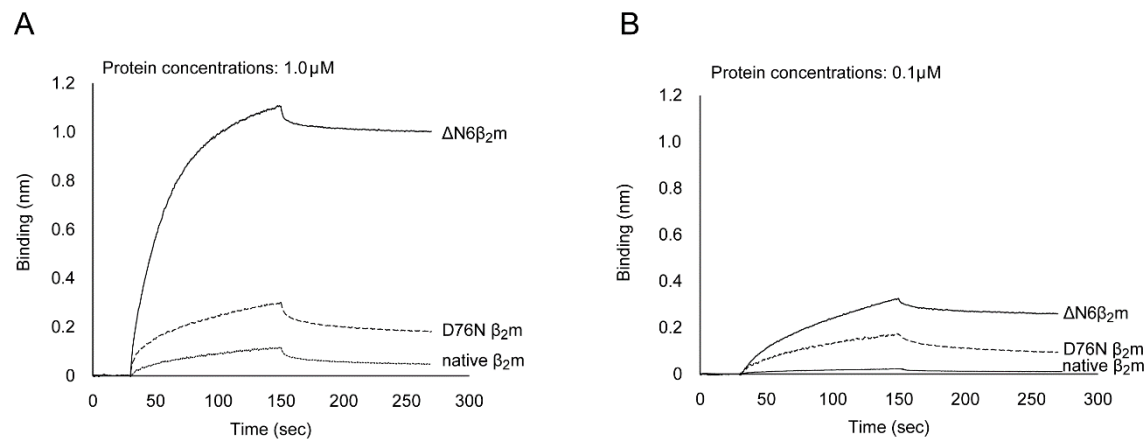
Study demonstrated a clear difference between HMH and LMH in interactions with  $\beta_2$ -M, as follows [7].

Heparin is a mucopolysaccharide with different molecular weights (MWs) when prepared from animal organs, such as the intestine or the lung, and has been used widely as an anticoagulant. In the early 1990s, low-molecular-weight heparin (LMH), less than 6 kD, was introduced in HD to reduce the high risk of bleeding that had been associated with conventional heparin, which had heterogeneous MWs of more than 10 kD, or with high-molecular-weight heparin (HMH) [6]. Our previous

### Binding of Heparin with $\beta_2$ -M

HMH binds in a dose-dependent manner with  $\Delta N6\beta_2\text{-M}$ , a well-known highly amyloidogenic variant of  $\beta_2\text{-M}$  [8]; binds somewhat with D76N

$\beta_2\text{-M}$ , a natural amyloid variant of  $\beta_2\text{-M}$ ; and demonstrates weak binding with native  $\beta_2\text{-M}$  (**Figure 2**).

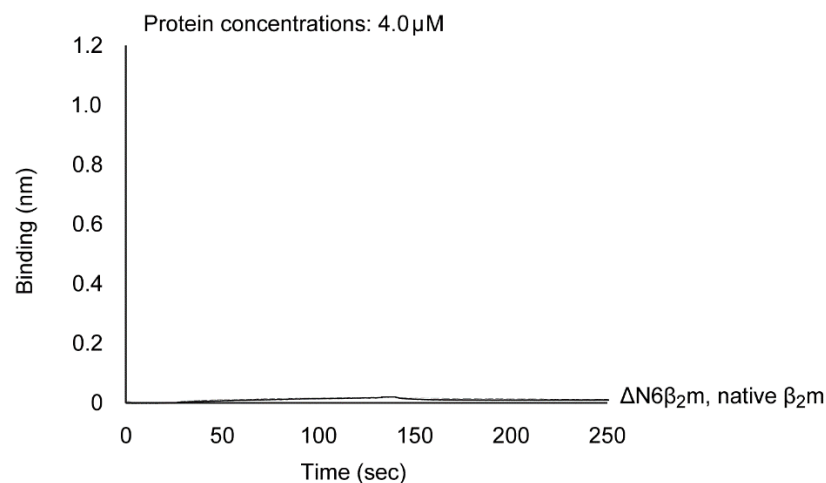


**Figure 2:** Analysis of the interaction of  $\beta_2\text{m}$  variants with high molecular weight heparin.

Biotinylated H.M.H. (0.5 U/mL) was immobilized onto streptavidin biosensor. Protein concentrations were 1.0  $\mu\text{M}$  (A) and 0.1  $\mu\text{M}$  (B), respectively.  $\Delta N6\beta_2\text{m}$  (line, —), D76N  $\beta_2\text{m}$  (dashed line, ----) and native  $\beta_2\text{m}$  (dotted line, ..).

The  $k_D$  values for each  $\beta_2\text{-M}$  species were as follows:  $2.07 \times 10^{-8}\text{M}$ , and  $1.72 \times 10^{-7}\text{M}$ ,  $3.71 \times 10^{-6}\text{M}$  respectively. The  $k_D$  value of  $\Delta N6\beta_2\text{-M}$  was low, consistent with that of its specific aptamer [7].

LMH, however, did not bind with  $\Delta N6\beta_2\text{-M}$  or native  $\beta_2\text{-M}$  (**Figure 3**).



**Figure 3:** Analysis of the interaction of  $\beta_2\text{m}$  variants with low molecular weight heparin.

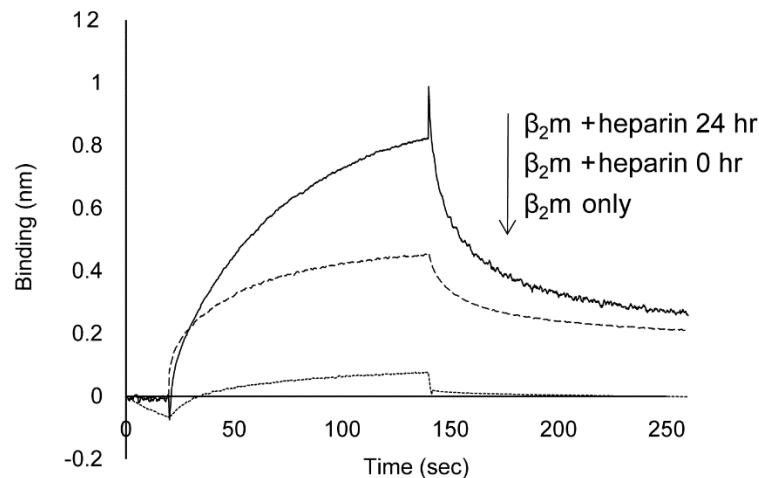
Biotinylated L.M.H. (5.0 U/mL) was immobilized onto streptavidin biosensor.  $\Delta N6\beta_2\text{m}$  (line, —) and native  $\beta_2\text{m}$  (dotted line, ..) were used as the binding partner. Protein concentration was 4.0  $\mu\text{M}$ . The real time binding curves were used to compute equilibrium dissociation constant by globally fitting the rate equation for 1:1 kinetics to the data. Three-independent experiments were performed, respectively.

An intermediate molecule with a partially unfolded structure of precursor proteins was identified as a key molecule in the amyloidogenic process [10]. With regard to  $\beta_2\text{-M}$ , we confirmed that the unfolding at the C-terminus from Ile-92 to Met-99 was key process for the initiation of

Because GAGs in the interstitial space have a rich heparin component,  $\beta_2\text{-M}$  has been trapped in the interstitial space and accumulated in a time-dependent fashion in patients undergoing HD, and their levels of  $\beta_2\text{-M}$  are expected to reach as high as millimolar values in 10 years or more.

### Unfolding of the C-terminus of the $\beta_2\text{-M}$ molecule

amyloidogenicity of this precursor protein (**Figures 1–3**). Our study of heparin demonstrated that 1  $\mu\text{M}$  HMH could trigger the C-terminal unfolding of  $\beta_2\text{-M}$ , which was consistent with the serum concentrations of  $\beta_2\text{-M}$  in patients undergoing HD (**Figure 4**) [7].



**Figure 4:** Analysis of the interaction of mAb92-99 with native  $\beta_2m$  in the presence of H.M.H.

MAb92-99 (10  $\mu\text{g/mL}$ ) was immobilized onto protein a biosensor. After adding native  $\beta_2m$  to the reaction drop holder in the presence or absence of H.M.H., the real-time binding was monitored. Native  $\beta_2m$  incubated with H.M.H. (0.5 U/mL) for 24 h was also used as the binding partner.

In addition, given that  $\beta_2\text{-M}$  concentrations may be much higher in the interstitial space than are concentrations in serum, we may expect more unfolding at the N-terminus than the C-terminus, which is likely to undergo proteolysis at Lys-6 and generate a highly amyloidogenic variant,  $\Delta\text{N6}\beta_2\text{-M}$  [8], because the unfolding process is believed to proceed from the N-terminus of  $\beta_2\text{-M}$  [11].

### GAGs Contain a Majority of the $\text{SO}_3^-$ Moiety

Chemical solvents containing  $\text{SO}_3^-$  such as sodium dodecyl sulfate have induced conformational changes in proteins. For  $\beta_2\text{-M}$ , several studies indicated amyloidogenic conversion that depended on a majority of  $\text{SO}_3^-$  moieties being present in GAGs [12-15]. As is well-known, heparin is a highly negative mucopolysaccharide rich in the  $\text{SO}_3^-$  moiety. Although we had not studied the dose-dependent effect of heparin on the C-terminal unfolding of  $\beta_2\text{-M}$ , we believe that the  $\text{SO}_3^-$  majority contained in heparin may affect the results for HMH and LMH. The conformation of the  $\beta_2\text{-M}$  molecule is maintained by multiple intramolecular hydrogen bonds, which may be partially broken by multiple  $\text{SO}_3^-$  groups.

### Conclusion

Collectively, our studies have indicated two actions of HMH with  $\beta_2\text{-M}$ : direct binding and induction of C-terminal unfolding. The former may result in an accumulation of  $\beta_2\text{-M}$  in the interstitial space and the latter may lead to  $\beta_2\text{-M}$  amyloidogenicity.

LMH showed no clear interactions, even with  $\Delta\text{N6}\beta_2\text{-M}$ , which indicates a superior clinical availability compared with HMH as an anticoagulant during long-term HD, in which DRA is a serious complication.

### Reference

1. Gejyo F, Yamada T, et al; A new form of amyloid protein

associated with chronic hemodialysis was identified as  $\beta_2$ -microglobulin. *Biochem Biophys Res Commun* 29; 701-706, 1985.

2. Connors LH, Shirahama T, Skinner M, Fenves A, Cohen AS; In vitro formation of amyloid fibril from intact beta-2-microglobulin. *Biochem Biophys Res Commun* 31; 1063-1068, 1985.

3. Motomiya Y, Ando Y, et al; Studies on unfolded  $\beta_2$ -microglobulin at C-terminal in dialysis-related amyloidosis. *Kidney Int* 67; 314-320. 2005

4. Motomiya Y, Higashimoto Y, et al; C-terminal unfolding of an amyloidogenic  $\beta_2$ -microglobulin fragment:  $\Delta\text{N6}\beta_2$ -microglobulin. *Amyloid* 22,54-60.2015.

5. van Ypersele de Strihou C, Jadour M, Maldague B, Jamart J, the Working party on dialysis amyloidosis: Effect of dialysis membrane and patient's age on signs of dialysis-related amyloidosis. *Kidney Int* 39, 1012-1019, 1991

6. Schrader J, et al; Comparison of low molecular weight heparin to standard heparin in hemodialysis/hemofiltration. *Kidney Int* 33, 890-896, 1988.

7. Fukazawa K, Higashimoto Y. et al; Influence of heparin molecular size on the induction of C-terminal unfolding in  $\beta_2$ -microglobulin. *Mol Biol Res Com* 5, 225-232, 2016.

8. Esposito G, Michelutti R et al; Removal of the N-terminal hexapeptide from human  $\beta_2$ -microglobulin facilitates protein aggregation and fibril formation. *Protein Sci* 9: 831-845, 2000.

9. Fukazawa K, Higashimoto Y et al; Selection of DNA aptamer that blocks the fibrillogenesis of a proteolytic amyloidogenic fragment  $\beta_2\text{M}$ . *Ther Apher Dial* 22, 61-66, 2017.

10. Chiti F, Mangione P, et al; Detection of two partially structured species in the folding process of the amyloidogenic protein  $\beta_2$ -microglobulin. *J Mol Biol* 307; 379-391, 2001.

11. McParland VJ, Kalverda AP, et al; Structural properties of an amyloid  $\beta_2$ -microglobulin. *Nat Struct Biol* 9: 326-331. 2002.

12. Yamamoto S, Hasegawa K, et al; Low concentrations of sodium

dodecyl sulfate induce the extension of beta 2-microglobulin-related amyloid fibrils at a neutral pH. *Biochemistry* 43, 11075-11082, 2004.

13. Yamamoto S, Yamaguchi I, et al; Glycosaminoglycans enhance the trifluoroethanol-induced extension of beta 2-microglobulin-related amyloid fibrils at a neutral pH. *J Am Soc Nephrol* 15, 126-133, 2004.
14. Borysik AJ, Morten IJ, et al; Specific glycosaminoglycans promote unseeded amyloid formation from beta 2-microglobulin under physiological conditions. *Kidney Int* 72; 174-181, 2007.
15. Relini A, De Stefano S, et al; Heparin strongly enhances the formation of beta 2-microglobulin amyloid fibrils in the presence of type I collagen. *J Biol Chem* 283, 4912-4920, 2008.



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